

Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK



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Accredited to
ISO 15189:2022

Liverpool University Hospitals NHS Foundation Trust

Issue No: 010 Issue date: 15 April 2025

Mount Vernon Street
Liverpool
L7 8YE
United Kingdom

Contact: Ines Santos
Tel: +44 (0)151 706 4374
E-Mail: Ines.Santos@liverpoolft.nhs.uk
Website: www.liverpoolcl.nhs.uk/

Testing performed by the Organisation at the locations specified below

Locations covered by the organisation and their relevant activities

Laboratory locations:

Location details	Activity	Location code
Address Clinical Sciences Services Building Royal Liverpool Hospital Liverpool University Hospitals Foundation Trust Mount Vernon Street Liverpool L7 8YE United Kingdom	Local contact James Wingfield 0151 706 2000 james.wingfield@liverpoolft.nhs.uk	Diagnostic cytology(non-Gynae) Special Stains Immunocytochemistry Routine Histopathology Frozen Section Histology Immunofluorescence Molecular Pathology Mortuary Services (body receipt, storage and release)



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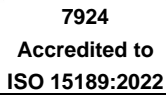
Liverpool University Hospitals NHS Foundation Trust
Trust

Issue No: 010 **Issue date:** 15 April 2025

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Site activities performed away from the locations listed above:

Location details	Activity	Location code
Address FNA Clinic Aintree Hospital Longmoor Lane Liverpool L9 7AL Local Contact Claire Chadwick 0151 706 5623 claire.chadwick@liverpool.nhs.uk	FNA slide preparation, adequacy assessment and reporting	B
Address Mortuary Royal Liverpool University Hospital Mount Vernon Street Liverpool L7 8YE Local Contact Neil White 0151 706 3801 neil.white@liverpoolft.nhs.uk	Mortuary Services (body receipt, storage and release)	A
Address Mortuary Aintree University Hospital Lower Lane Liverpool L9 7AL Local Contact Neil White – as above	Body storage	C
Address Mortuary Broadgreen Hospital Thomas Drive Liverpool L14 3LB Local Contact Neil White – as above	Body storage	D
Address Theatres Royal Liverpool University Hospital Prescot Street Liverpool L7 8XP Local Contact Georgio Bristol 0151 706 4677 georgio.bristol.liverpoolft.nhs.uk	Renal sample adequacy check.	E



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
HUMAN TISSUES AND FLUIDS	<u>Diagnostic Cytology</u>		
CSF, urine, general body fluids, bronchial brushings, EBUS, ERCP	Preparation and examination of cellular material to identify or exclude morphological and cytological abnormalities.	In-house documented procedures using: <u>Preparation of cell samples</u> Sakura Cyto-Tek Centrifuge Thermoscientific centrifuge CP-NCER-SOP-23 Genesis T200 slide processor and Leica ST5020 Multistainer and CV5030 Coverslipper CP-NCER-SOP-4 CP-NCE-SOP-5 Preparation of fluids requiring cytopspin CP-NCER-SOP-1 Fine needle aspiration cytology <u>Fluid preparation using Holigic thin prep CP-NCER-SOP-11</u> <u>Production of cell block</u> CP-NCER-SOP-10 Preparation of an Agar Cell Block	A
CSF, urine, general body fluids, bronchial brushings, EBUS, ERCP, FNA	Preparation and examination of cellular material to identify or exclude morphological and cytological abnormalities.	<u>Staining</u> In-house procedures using Leica stainer ST5020 and May Grunwald Giemsa stain CP-NCER-SOP-4 CP-NCER-SOP-2 May Grunwald Giemsa method	A
FNA	Morphological assessment in order to ascertain adequacy for diagnosis and interpretation/diagnosis	<u>Material/slide preparation and staining</u> May Grunwald stain CP-NCER-SOP-2 and CP-NCER-SOP-1 Fine needle aspiration cytology	B
CSF, urine, general body fluids, bronchial brushings, EBUS, ERCP, FNA		Interpretive Microscopy Non cervical cytology reporting Guidelines CP-NER-SOP-15	A



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<p>HUMAN BODY TISSUE SAMPLES</p> <p>Fixed, fresh and frozen tissue; excisional and incisional biopsies and surgical resection specimens</p> <p>Formalin Fixed Paraffin wax blocks of processed tissue.</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis</u></p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Specimen dissection: Envair Downdraft tables</p> <p>Handling of ocular referral specimens form external HM Coroner and Police forces CP-OPAT-SOP-10</p> <p>Routine Bone decalcification CP-SPE-SOP-20</p> <p>Tissue processing:</p> <p>Leica Peloris II Automated tissue processor CP-HIS-SOP-53</p> <p>Leica Peloris III automated tissue processor CP-HIS-SOP-76</p> <p>Embedding: Thermo Histostar CP-HIS-SOP-8</p> <p>Microtomy: Leica RM2255 automated microtomes CP-HIS-SOP-17</p>	A



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<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>Tissue for frozen sections</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides and frozen sections</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis (cont'd)</u></p> <p>Morphological staining For the detection of:</p> <p>Basophilic and eosinophilic structures</p> <p>Special stains for the detection of:</p> <p>Acid and neutral mucopolysaccharides, glycogen and fungi</p> <p>Acid and Neutral Mucopolysaccharides</p> <p>Microorganisms and haemopoietic cells</p> <p>Connective tissue</p> <p>Fibrin</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Cryotomy Intra-operative Frozen section Leica CM1850 UV Leica CM 1860 UV CP-HIS-SOP-27</p> <p>Interpretive/diagnostic microscopy CP-PATH-POL-2</p> <p>Routine morphological staining using Tissue tek Prisma Stained and Tissue Tek Glas coverslipper CP-HIS-SOP-24</p> <p>Haematoxylin and Eosin (H & E) manual stain CP-HIS-SOP-9</p> <p>Diastase Alcian Blue Periodic acid Schiffs (DABPAS) (ABPAS) CP-HIS-SOP-43</p> <p>Alcian Blue CP-HIS-SOP-61</p> <p>Geimsa CP-HIS-SOP-44</p> <p>Gomori's trichrome (one stage) CP-HIS-SOP-45</p> <p>Martius Scarlet Blue (MSB) CP-HIS-SOP-46</p>	A



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<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides (cont'd)</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis (cont'd)</u></p> <p>Special stains for the detection of: (cont'd)</p> <p>Reticulin</p> <p>Spirochaetes</p> <p>Mast cells</p> <p>Amyloid</p> <p>Leprosy</p> <p>Bacteria</p> <p>Fungi</p> <p>Connective tissue</p> <p>Basement membranes</p> <p>Argentaffin cells, melanin</p> <p>Connective tissue</p> <p>Melanin</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Gordon and Sweets CP-HIS-SOP-47</p> <p>Steiner CP-HIS-SOP-48</p> <p>Toluidine Blue CP-HIS-SOP-50</p> <p>Congo red CP-HIS-SOP-2</p> <p>Fite-Faraco CP-HIS-SOP-3</p> <p>Gram CP-HIS-SOP-4</p> <p>Grocott Silver CP-HIS-SOP-28</p> <p>Haematoxylin Van Gieson (HVG) CP-HIS-SOP-22</p> <p>Jones methanamine silver CP-HIS-SOP-23</p> <p>Masson Hamperl CP-HIS-SOP-30</p> <p>Masson Trichrome CP-HIS-SOP-107</p> <p>Melanin bleach CP-HIS-SOP-31</p>	A



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HUMAN BODY TISSUE SAMPLES (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):	A
FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides (cont'd)	Special stains for the detection of: (cont'd)		
	Elastin	Millers Elastic Van Gieson (EVG) CP-HIS-SOP-21	
	Haemosiderin and ferric iron	Perls CP-HIS-SOP-5	
	Hep B Sag and copper associated protein	Shikata Orcein CP-HIS-SOP-32	
	Calcium	Van Kossa CP-HIS-SOP-51	
	Tubercle bacilli	Ziehl Neelson CP-HIS-COP-6	
Formalin fixed lung tissue on glass slide	Asbestos fibres	Post mortem Asbestos Fibre Count CP-SPE-SOP-31	
FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalins fixed cytopspins slides	Immunohistochemistry to detect the following:	Immunocytochemistry Dako Omnis Basic User SOP: CP-IMM-SOP-33 Immunocytochemistry Dako Omnis Advanced User: CP-IMM-SOP-36 Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18 Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6	
	High molecular weight cytokeratins	34BE12	
	A1AT deficiency	A1AT (Alpha 1 Anti-trypsin)	



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<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalins fixed cytopins slides (cont'd)</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis (cont'd)</u></p> <p>Immunohistochemistry to detect the following: (cont'd)</p> <p>Pan-cytokeratin</p> <p>Androgen receptor in prostate and breast tumours</p> <p>B-Catenin protein in fibromatous lesions</p> <p>BRCA associated protein</p> <p>Diffuse lymphoproliferative disease and neoplastic cells of follicular lymphomas</p> <p>Germinal centre B-lymphs</p> <p>Epithelial cell types</p> <p>Ovarian tumour marker</p> <p>Parafollicular cells in thyroid and medullary thyroid carcinoma</p> <p>Marker of smooth muscle</p> <p>Marker of mesothelioma</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Immunocytochemistry Dako Omnis Basic User SOP: CP-IMM-SOP-33</p> <p>Immunocytochemistry Dako Omnis Advanced User: CP-IMM-SOP-36</p> <p>Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18</p> <p>Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6</p> <p>AE1AE3</p> <p>AR (Androgen Receptor)</p> <p>B-CAT (Beta catenin)</p> <p>BAP-1 (BRCA associated protein)</p> <p>BCL-2</p> <p>BCL-6</p> <p>BEREP4</p> <p>CA125</p> <p>Calcitonin</p> <p>Caldesmon</p> <p>Calretinin</p>	A



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		Immunocytochemistry Dako Omnis Advanced User: CP-IMM-SOP-36	
		Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18	
		Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6	
	Endothelial cells	CD31	
	Endothelial cells and other cell types in various solid tumours	CD34	
	Plasma cells	CD38	
	T-cells and myeloid lineage cells	CD43	
	Lymphocytes and reactive urothelium	CD44	
	T and B lymphocytes	CD45 (Leucocyte common antigen)	
	NK cells, nerves and neuroendocrine differentiation	CD56	
	NK cells	CD57	
	Megakaryocytes and platelets	CD61	
	Macrophages, myeloid cells and monocytes	CD68 (KP1)	
	Macrophages and monocytes	CD68 (PG)	



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		Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6	
	B lymphocytes	CD79a	
	Ewing's sarcoma	CD99	
	Stem cells, melanocytes , mast cells and GIST tumours	CD117 (c-kit)	
	Plasma cells	CD138	
	Lining of blood and lymphatic vessels and mesothelial cells	CD141(thrombomodulin)	
	Intestinal epithelia and GI tract tumours	CDX2	
	Epithelial marker	CEA-M (carcinoembryonic antigen)	
	Neuroendocrine cells	CHROMA (Chromogranin A)	
	Stratified squamous epithelia, basal cells and mesotheliomas	Ck5/6 (Cytokeratins 5 & 6)	
	Normal and neoplastic epithelia	CK7 (Cytokeratin 7)	
	Normal and neoplastic epithelia	CK19 (Cytokeratin 19)	



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		Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6	
	Mucin glycoprotein-1	MUC-1	
	Mucin glycoprotein-5AC	MUC-5	
	Mucin glycoprotein-6	MUC-6	
	Multiple myeloma oncogene-1	MUM-1P	
	Myeloperoxidase in neutrophils and granulocytes	MYELO	
	Rhabdomyosarcomas	MYO-D1	
	Rhabdomyosarcoma and Wilm's tumours	MYOGEN	
FFPE Slides (formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalin fixed cytopspin slides	Pluripotent germ cell tumours, i.e. seminoma	OCT 3/4	
	Epithelial basal cell marker	P63	
	HGPIN and prostatic adenocarcinoma	P504S	



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issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Liverpool University Hospitals NHS Foundation Trust
Trust

Issue No: 010 Issue date: 15 April 2025

Testing performed by the Organisation at the locations specified

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalins fixed cytopspins slides (cont'd)</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis</u></p> <p>Immunohistochemistry to detect the following: (cont'd)</p> <p>Prostatic glandular epithelium and prostate carcinoma</p> <p>Thyroid, renal, Mullerian and thymic tumours</p> <p>Activated T-cells and marker for angioimmunoblastic lymphoma</p> <p>Expression of PD-L1 protein in NSCLC</p> <p>Neuroendocrine cells and neurons</p> <p>Cellular Progesterone receptor expression</p> <p>Germ cell neoplasms and in breast, ovarian and lung carcinomas</p> <p>Normal and neoplastic melanocytes, adipocytes, neural cells and tumours</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Immunocytochemistry Dako Omnis Basic User SOP: CP-IMM-SOP-33</p> <p>Immunocytochemistry Dako Omnis Advanced User: CP-IMM-SOP-36</p> <p>Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18</p> <p>Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6</p> <p>PAPH</p> <p>PAX-8</p> <p>PD-1</p> <p>PD-L1 PD-L1 IHC 22C3 pharmDX kit</p> <p>PGP 9.5</p> <p>PGR</p> <p>PLAP</p> <p>S100</p>	



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<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalins fixed cytopins slides (cont'd)</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis (cont'd)</u></p> <p>Immunohistochemistry to detect the following: (cont'd)</p> <p>Smooth muscle, myoepithelial can other contractile cells</p> <p>Detection of primary and metastatic renal cell carcinomas</p> <p>Melanoma marker</p> <p>Neuroendocrine cells and neurones</p> <p>Marker for normal and malignant thyroid</p> <p>Dermal melanocytes, melanocytic lesions and malignant melanomas</p> <p>Cells of mesenchymal origin and mesenchymal tuours</p> <p>Wilm's tumour, malignant mesothelioma, ovarian cystadenocarcinoma, nephroblastoma and desmoplastic small round cell tumours</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Immunocytochemistry Dako Omnis Basic User SOP: CP-IMM-SOP-33</p> <p>Immunocytochemistry Dako Omnis Advanced User: CP-IMM-SOP-36</p> <p>Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18</p> <p>Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6</p> <p>SMA</p> <p>RCC</p> <p>SOX-10</p> <p>SYN</p> <p>THGLOB</p> <p>TYROS</p> <p>VIM</p> <p>WT-1</p>	<p>A</p>



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<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalins fixed cytopins slides (cont'd)</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)</p> <p>Immunohistochemistry to detect the following: (cont'd)</p> <p>Sebaceous carcinoma</p> <p>Primary liver tumours and yolk sac tumours</p> <p>Myoglobin in skeletal and cardiac muscle</p> <p>Anaplastic lymphoma kinase (CD246) in ALCL</p> <p>Hairy cell leukaemia</p> <p>Dendritic cells</p> <p>Common B-cell marker expressed on B-cells and follicular dendritic cells</p> <p>Follicular dendritic cell</p> <p>IL-3 receptor in monocytes, neutrophils and basophils</p> <p>Macrophages</p> <p>Endogenous C-Myc</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Immunocytochemistry Use of Dako AS Link 48 SOP: CP-IMM-SOP-17 and Immunocytochemistry Use and Operation of PT Link SOP: CP-IMM-SOP-24</p> <p>Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18</p> <p>Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6</p> <p>Adipophilin</p> <p>Alpha fetoprotein (AFP) CP-IMM-COP-17</p> <p>MYOG</p> <p>ALK-1</p> <p>Annexin A1</p> <p>CD11c</p> <p>CD19</p> <p>CD21</p> <p>CD123</p> <p>CD163</p> <p>C-MYC</p>	A



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HUMAN BODY TISSUE SAMPLES (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):	A
FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides, cell blocks and formalins fixed cytopins slides (cont'd)	Immunohistochemistry to detect the following: (cont'd)	Immunocytochemistry Use of Dako AS Link 48 SOP: CP-IMM-SOP-17 and Immunocytochemistry Use and Operation of PT Link SOP: CP-IMM-SOP-24	
		Immunocytochemistry Use of Dako Coverslipper: CP-IMM-SOP-18	
		Interpretive Microscopy procedures: Pathologist Reporting and Cut-up: CP-PATH-POL-2 and PD-L1 Reporting Procedures: CP-PATH-POL-6	
	B-cell centroblastic cell line	DBA44	
	Epstein Barr Virus infection	EBV	
	Erythroid cells	GLYCA (Glycophorin A)	
	Splenic marginal zone lymphoma and B-cell lymphocytic lymphoma	IgD delta chain	
	Mast cells	MCT	
	Neuronal marker	NFP (Neurofilament protein)	
	Marker of molar pregnancy	P57	
	Renal cell oncocyoma tumour	RON-A	
	Detection of SV40 virus in renal transplant samples	SV40	
	Thymic cortical lymphocytes and lymphoblastic lymphomas and leukaemias	TDT	
FFPE slides (formalin fixed paraffin embedded) tissue on glass slides	Cytotoxic T-cells	TIA-1 OCT 2	



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HUMAN BODY TISSUE SAMPLES (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):	A
FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides and cell blocks	Immunohistochemistry to detect the following: (cont'd)	Ventana Bench Mark Ultra CP-IMM-SOP-19	
	ALK protein in NSCLC	ALKD5F3	
	HER2 amplification in breast tumours	HER-2	
	Expression of PD-L1 protein in NSCLC	PD-L1	
	Dermatofibroma	FAC 13A	
		GATA-3	
		NKX3.1	
		ROS1 (clone SP384)	
	Mantle cell lymphoma	BCL-1 (Cyclin -D1)	
	Marker of rejection in renal transplants	CD4	
	Glial fibrillary acidic protein neural cells	GFAP (Glian fibrillary acidic protein)	
	Smooth muscle and myoepithelial cells	SMMS-1	
	Mantle cell lymphoma marker	SOX-11	
FFPE Slides (formalin fixed paraffin embedded) tissue on glass slides		PD-L1 (clone SP142)	
		STAT6	



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HUMAN BODY TISSUE SAMPLES (cont'd) Fresh frozen skin biopsy tissue	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd) Identification of: Immunoglobulins and complement	In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant): Direct Immunofluorescence technique: Documented in-house procedures in conjunction with manufacturer's instructions using DAKO FITC-conjugated antibodies Manual techniques CP-IMM-SOP-16 IgA IgG C3	A



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<p>HUMAN BODY TISSUE SAMPLES (cont'd)</p> <p>Stained slides prepared as above</p> <p>Breast tissue</p> <p>FFPE tissue, cell blocks, cytospin slides</p> <p>FFPE slides (Formalin fixed paraffin embedded) tissue on glass slides</p>	<p><u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)</p> <p><u>Molecular Pathology</u></p> <p>Morphological assessment and interpretation/diagnosis</p> <p>Fluorescent in-situ hybridisation (FISH)</p> <p>Determination of HER 2 gene amplification</p> <p>Chromogenic in situ hybridisation (CISH)</p> <p>Epstein Barr Virus RNA</p>	<p>In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p>Automated techniques using Dako Omnis CP-MOL-SOP-17</p> <p>Fluorescence microscope Olympus BX 61 CP-MOL-SOP-15</p> <p>Interpretive/diagnostic microscopy CP-IMM-SOP-11</p> <p>Evaluation and reporting of HER 2 tests CP-IMM-SOP-11</p> <p>ROS 1IQFISH Break-apart probe</p> <p>Automated techniques using Ventana Benchmark Ultra CP-IMM-SOP-19 and CISH probes</p> <p>INFORM EBER probe</p>	A



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HUMAN BODY TISSUE SAMPLES (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	In house documented procedures using manual methods or analysers in conjunction with manufacturer's instructions for the following methods (where relevant):	A
	<u>Molecular Pathology</u> (cont'd)		
	Morphological assessment and interpretation/diagnosis (cont'd)		
Tissue block	BRAF mutation screening	Automated techniques using Biocartis Idylla CP-MOL-SOP-25	
	EGFR mutation screening		
FFPE samples, vitrous fluid, whole blood	DNA extraction	Automated technique using QIAcube CP-MOL-SOP-23	
Genomic DNA from the above sample types	DNA quantification	Automated technique using Qubit 4.0 Fluorimeter and Qubit dsDNA BR assay kit CP-MOL-SOP-24	
Genomic DNA from the above sample types	Detection of B and T cell gene mutation	Clonality Assay using ABI 3500 CP-MOL-SOP-20 CP-MOL-SOP-19	
Genomic DNA extracted and quantified from FFPE using above techniques	Non-Small Cell Lung Cancer (NSCLC)	Next Generation Sequencing using Genexus Integrated Sequencer and Oncomine Precision Assay GX Data analysis using Oncomine Precision GX Solid Tumour DNA and Fusions workflow CP-MOL-SOP-32	
END			