Schedule of Accreditation

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK



8176

Accredited to ISO 15189:2022

Birmingham Women's and Children's NHS Foundation

Issue No: 013 Issue date: 04 June 2025

Birmingham Women's Hospital

Mindelsohn Way **Edgbaston**

Birmingham

B15 2TG

Contact: Jennie Bell Tel: +44 (0)1214 721377 E-Mail: jennie.bell1@nhs.net

Website: www.bwc.nhs.uk

Testing performed by the Organisation at the locations specified below

Locations covered by the organisation and their relevant activities

Laboratory locations:

Location details		Activity
Birmingham Women's Hospital West Midlands Regional Genetics Laboratory Mindelsohn Way Edgbaston Birmingham B15 2TG	Local contact Jennie Bell	Molecular Genetics

Site activities performed away from the locations listed above:

Location details		Activity
Birmingham Research Park Limited Institute of Research and Development Birmingham Research Park Vincent Drive Edgbaston Birmingham B15 2SQ	Local contact Jennie Bell	Reporting

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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
	Sample processing, DNA/RNA extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long term storage	
Whole Blood		Lineage specific cell separation Automated using Automacs
Bone Marrow		Miltenyi Biotech autoMACS®Pro Separator
		SOP: PP 02.01.95
		Manual and automated DNA extraction and quantification using:
Whole Blood Bone Marrow		Qiagen QIASymphony SP platform with:
Saliva (Oragene self collection kit) Plasma		QIAsymphony DNA Midi Kit QIAsymphony DSP virus kit
		SOPs:
		PP 03.01.21 PP 03.01.13 PP 03.01.48
Whole Blood Bone Marrow		Qiagen QIAcube with Qiagen QIAamp DNA Blood Mini Kit
		SOP: PP 03.01.15

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
FFPE Dried Blood Spots		Promega Maxwell 16 with Promega Maxwell 16 FFPE Plus LEV DNA Purification Kit and LEV Blood Kit
		PP 03.01.43 PP 03.01.46
MACS separated cell fractions Amniotic Fluid		Qiagen EZ1 using EZ1Tissue Kit
CVS		PP 03.01.08: PP 03.01.14: PP 03.01.10:
Fresh frozen solid tissue*		Qiagen EZ1 using EZ1Tissue Kit with manual homogenation
		PP 03.01.07
Fresh tissue Foetal Tissue POC		Qiagen EZ1 using EZ1Tissue Kit with tissue abruption using Precellys 24 Homogeniser
		PP 03.01.19
		Manual DNA Extraction:
Peripheral Blood		Qiagen Gentra Puregene Kit
Bone Marrow Foetal Blood Mouth Wash Cultured Cells		PP 03.01.02 PP 03.01.06 PP 03.01.05
Peripheral Blood		Biorad Instagene matrix
		PP 03.01.12
Peripheral Blood		Phenol Chloroform extraction
Bone Marrow, Foetal Blood CVS,Fixed Cells		PP 03.01.03, PP 03.01.04 PP 03.01.11

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Peripheral Blood Bone Marrow Foetal Blood		Phenol Chloroform extraction – High Risk protocol
Foetal blood		PP 03.01.24
Saliva (Oragene self collection kit)		DNA extraction using Genotek Prep-IT kit
		PP.03.01.13
Genomic DNA extracted inhouse from the sample types listed above or received as primary sample type from an external source		DNA Quantification for QC purposes Nanodrop 8000 Qubit 2.0/4.0 fluorometerPicogreen quantification process for DNA samples using Fluroskan
		SOPs: PP 03.01.27 PP 03.01.36 PP 03.01.50
	Sample processing, DNA/RNA extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long term	Automated dual DNA and RNA extraction and quantification using:
FFPE	storage	Dual DNA and RNA extraction using Promega Maxwell 16 LEV RNA FFPE Purigication Kit and Promega Maxwell IVD
		MP 01.01.55

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
	Sample processing, DNA/RNA extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long term storage	Automated RNA extraction using:
Whole Blood Bone Marrow		Extraction of total RNA using the Promega Maxwell IVD and Promega Maxwell 16 LEV simply RNA Blood Kit
		PP 03.01.47
RNA extracted inhouse from the sample types listed above or received as primary sample type		Reverse Transcritpion Manual -High Capacity cDNA Reverse Transcriptase kit (Applied Biosystems)
from an external source		PP 03.01.26:
RNA extracted inhouse from FFPE		RNA Quantification for QC purposes Qubit
		PP 03.01.36
		Fluorescence based ARMS PCR
Genomic DNA extracted in-house from the sample types listed and	Detection of SNVs and Small indels	Multiplexing Luminex commercial CF XTAG kit:
received as primary samples from external sources	[definitive list QA 01.02.60]	Equipment: Luminex 200
		Analysis using:
		Luminex integrated software
		SOP: FRAG 01.01.05

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA and RNA extracted in-house from the sample types	Detection of nuclei acid sequence variants – SNVs, small indels and	Sanger Sequencing
listed and received as primary	splice site mutations	Using
samples from external sources	[definitive list QA 01.02.60]	Standard primer design methodology and PCR amplification (where relevant to internal samples and confirmatory processes)
		SOP: SEQ 01.01.26 PP 03.01.35
		And:
		PCR blocks, ABI 3730 Capillary electrophoresis instruments
		Analysis and interpretation of variants by Mutation Surveyor software.
		SOP: 01.01.10 SEQ 01.01.25
		Fragment Length Analysis
		Fragment Length Analysis using Capillary Electrophoresis
Genomic DNA extracted in-house from the sample types listed and received as primary samples from external sources	Detection of fragment length size, deletions, known SNVs and small indels, gene rearrangements, repeat expansions, linkage makers,	PCR, methylation specific PCR, Triplet Repeat PCR using in-house methods
CAROTTAL SOCIOCS	short tandem repeats, microsatellites and methylation status	Equipment: PCR blocks and resolution using an ABI 3500XL capillary sequencer:
	[definitive list QA 01.02.60]	Analysis using GeneMapper and GeneMarker
		SOPs: FRAG 01.01.07 FRAG 01.01.13

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA extracted in-house from the sample types listed	Determination of copy number changes	Quantitative Fluorescence PCR (QF-PCR)
	[definitive list QA 01.02.60]	Using:
		In-house methods for trisomy screen
		And: Thermocyclers. Resolution by capillary electorphesis using ABI 3500XL.
		Analysis using Genemarker and Genemapper
		SOP: FRAG 01.01.16 PN 01.01.18
Genomic DNA and RNA extracted in house from the sample types listed above and received as	Qualitative Genotype analysis for SNVs, indels and fusion transcripts	Qualitative Reverse Transcriptase PCR (RTPCR) (Including nested)
primary samples from external sources (cont'd)	[definitive list QA 01.02.60]	Using:
		Agarose gel electrophoresis
		SOPs:TP 01.01.67 TP 01.01.61 TP 01.01.60
RNA extracted in house from the sample types listed above and	For the quantitative detection of common fusion transcripts	Quantitative Real Time PCR (RQ-PCR)
received as primary samples from external sources (cont'd)	[definitive list QA 01.02.60]	Using:
		In house methodology ABI 7500 Real time PCR system
		SOPs: QPCR 01.01.11 HOA 01.01.57 QPCR 01.01.27 HOA 01.01.61

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA extracted in-house from the sample types listed and received as primary samples from external sources	Determination of methylation status and copy number – deletions and duplications	Multiplex Ligation-dependent Probe Amplification (MLPA) and methylation specific (MS) MLPA.
	[definitive list QA 01.02.60]	Using
		Commercial MRC Hollandkits, thermocyclers and ABI3500xI
		Analysis using GeneMarker and Genemapper
		SOPs:FRAG 01.01.04 FRAG 01.01.10
Genomic DNA extracted in-house from the sample types listed above	Post-transplant Chimaerism analysis - % donor engraftment	Microsatellite analysis using in house PCR methodology and ABI 3500 XL.
		Analysis using; ChimerMarker Software
		SOPs: TP 01.01.63, HOA 01.01.14
Genomic DNA extracted in-house from the sample types listed above	Detection of known SNVs and indels at high sensitivity	Droplet Digital PCR
nom the sample types listed above	[definitive list QA 01.02.60]	Using:
	[definitive list Q/V01.02.00]	BioRAD commercial Kits and inhouse methodology
		BioRAD QX200 Droplet Reader
		BioRAD AutoDroplet Generator
		BioRAD PCR Plate Reader
		Analysis using integrated software
		SOPs: QPCR 01.01.01, QPCR 01.01.31

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
		Next Generation Sequencing
Cell-Free DNA extracted in-house from plasma	CNV/(Rapid screening for aneuploidy (T13,18,21)	VeriSeq Solution v 2.0
nom plasma		For
		Non-invasive Prenatal Testing (NIPT) using
		With
		Automated cfDNA extraction and Library Preparation using Hamilton Star Liquid Handler and with massively paralleled sequencing on a NextSeq 550 with VeriSeq NIPT LRM module for anlaysis.
		Receipting, Processing and Reporting of Lucina NIPT referrals PN 01.01.43
		And
		SEQ 01.01.69 NIPT Veriseq Technical protocol
Genomic DNA and RNA extracted in house from the sample types listed and samples received as	Gene screening of large gene panels for genetic variants –	Next Generation Sequencing:
primary samples from external sources (cont'd)	[definitive list QA 01.02.60]	Sequencing by MiSeq
sources (cont u)	SNVs/indels	KAPA Hyper plus/ NimbleGen (Roche) SOP:PN 01.02.10 PN 01.02.04
	SNVs/indels	Multiplex PCR (QIAgen) SOP:PN 01.02.02 PN 01.02.03

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA and RNA extracted in house from the sample types listed and samples received as primary samples from external sources (cont'd)		Next Generation Sequencing (cont):
		Analysis using:
		Agilent 2200 Tapestation, Qubit 2.0 Fluorometer, thermal cycler and 2.0 Fluorometer, Illumina MiSEQ
		Analysis using:
		BI 01.01.08: NIPD RHDO Bioinformatics Pipeline BI 01.01.07: NIPD Bespoke Bioinformatics Pipeline
		SEQ 01.01.22
DNA obtained from whole blood	DYPD testing	PCR with Maldi-TOF Agena MassArray PG 01.01.03

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Materials/Products tested HUMAN TISSUES AND BODY FLUIDS Whole Blood Amniotic Fluid CVS Foetal blood Products of Conception Bone Marrow Tissues/Skin Biopsy	Type of test/Properties measured/Range of measurement Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer G-banding/Karyotyping: Detection of chromosomal rearrangements or aberrations arising from: (e.g) Prenatal Diagnosis Reproductive Medicine Disorders Developmental Disorders Haemto/Oncology Disorders Chromosome Breakage Disorders (preparative pre-examination steps listed first)	Standard specifications/ Equipment/Techniques used Documented in house procedures incorporating manufacturer's instructions (where relevant) Culturing and processing of human tissue/cells using inhouse and commercial media to provide interphase cells: Cell Separation using autoMACS (see processesing section) Cell Harvesting Automated process using Multiprep Cell Sprint Robotic Harvester SOP TP 01.01.15 Manual Process SOP: PP 02.01.40 Chromosome analysis, Microscopic and Macroscopic analysis of G banded chromosomes using Manual Processing and Varistain Banding Instrument and microscopes Analysis using Metasystems Icaris SOPs: PP 02.01.92 PP 02.01.92 PP 02.01.93 PP 02.01.94 KARYO 01.01.06 GL 01.01.19 HOA 01.01.05	

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HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Formalin fixed paraffin embedded tissue (FFPE) Peripheral Blood Bone Marrow Fixed culture cells (more specific – cultured, uncultured PB, Marrow, AFs etc) Amniotic fluid CVS- Chronic villius samples	Detection of chromosomal aberrations in the diagnosis of haematological malignancy, bone marrow failure syndromes, non-haematological malignancies and constitutional disorders, solid tumours and companion testing Break-apart probes Fusion products Deletion Insertion Copy Number / Amplification	Fluorescence in situ hybridisation (FISH) Culturing and processing of human tissue/cells using in-house and commercial media to provide interphase/metaphase cells: Preparation and harvesting as for G-banding Manual Process PP 02.01.40 And commercial and in house developed probes. Hybridisation using Hybrite/Thermobrite Analysis Fluorescence microscope and metasystems ISIS SOPs FISH 01.01.04,FISH 01.01.95, FISH 01.01.97, GI 01.01.06
END		

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