

# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <p>Accredited to ISO 15189:2022</p>	<b>Oxford University Hospitals NHS Foundation Trust</b>	
	<b>Issue No: 013 Issue date: 10 October 2025</b>	
	<b>Cellular Pathology</b> <b>John Radcliffe Hospital</b> <b>Headley Way</b> <b>Headington</b> <b>Oxford</b> <b>OX3 9DU</b>	<b>Contact: Sharon Roberts-Gant</b> <b>Tel: +44 (0)1865-220494</b> <b>E-Mail: sharon.roberts-gant@ouh.nhs.uk</b> <b>Website: www.ouh.nhs.uk</b>
<b>Testing performed by the Organisation at the locations specified below</b>		

### Locations covered by the organisation and their relevant activities

#### Laboratory locations:

Location details	Activity	Location code	
John Radcliffe Hospital Headley Way Headington Oxford OX3 9DU	Sharon Roberts-Gant (contact details above)	Diagnostic Cytopathology Histopathology (inc. routine & special staining, frozen sections) Immunohistochemistry In-situ hybridisation Mortuary services - body receipt, storage & release	JRH
Churchill Hospital Old Road Headington Oxford OX3 7LE	Sharon Roberts-Gant (contact details above)	Slide production, staining & mounting of fine needle aspirates for diagnostic cytology. Mortuary Services - Body receipt, storage & release	CHF
Nuffield Orthopaedic Centre Oxford University Hospitals NHS Foundation Trust Headington Oxford OX3 7HE	Contact: Louisa Brook Tel: +44 (0)1865-738138 Fax: +44 (0)1865-738140 E-Mail: louisa.brook@ouh.nhs.uk Website: www.ouh.nhs.uk	Histopathology (inc. routine & special staining) Immunohistochemistry	NOC

#### Site activities performed away from the locations listed above:

Location details	Activity	Location code	
Horton General Hospital Oxford Road Banbury Oxfordshire OX16 9AL	Sharon Roberts-Gant (contact details above)	Mortuary Services - Body receipt, storage & release	HH



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<p>HUMAN TISSUES AND FLUIDS</p> <p>Fluids Brushings Fine needle aspirates slide preparations for cytological analysis</p> <p>Slides prepared in house from samples listed above</p>	<p><u>Diagnostic cytopathology</u></p> <p>Examination to identify or exclude cytological abnormalities for the purpose of diagnosis</p>	<p>Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p><u>Preparation/centrifugation</u> using procedures: NG02, NG03 in conjunction with manufacturer's instructions using: ThermoShandon Cytospin4, Heraeus Labofuge 400: NG08, NG40 Cytoblocks Kit NG32</p>	JRH
	<p>Examination to identify or exclude cytological abnormalities for the purpose of diagnosis</p>	<p><u>Staining</u> using in-house procedures for (as appropriate): Papanicolaou: NG19 May Grunwald Giemsa: NG20 Quick Diff: NG21 in conjunction with manufacturer's instructions using: Shandon Varistain 24-4: NG02, NG19</p>	JRH
		<p>Automated staining Papanicolaou Use and Maintenance of Tissue Tek Prisma™ Automated Staining Machine, H272</p>	JRH
	<p>Morphological assessment and interpretation/diagnosis</p>	<p><u>Microscopy (qualitative analysis)</u> Using procedures: CPSOP27 in conjunction with manufacturer's instructions using microscopes Leitz Dialux, Labourlux K, Laborlux S, Orthoplan, Orthomat II, Nikon Eclipse 50i, Ci-L, E200, E400 &amp; E600, Optishot Olympus BH2, BHTU, BH-U, BHS312, BHS313, BX40, BX41, BX50, BX51, BX53</p>	JRH CHF



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HUMAN TISSUES	<u>Histopathology</u>	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
Tissues: biopsy, excision, resection and post-mortem specimens	Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis	<u>Macroanalysis &amp; dissection</u> using: Cut up and Assisting H004,	JRH
		Specimen Handling at Cut Up, H005	JRH
		Decalcification and Tissue Softening, H008	JRH
		Frozen Section and Fresh Tissue Handling, H010	JRH
		Handling of POC/Ectopic Cases Following Specimen Dissection (HTA team), H294	JRH
		Use of Menarini MacroPath Pro-x Digital Camera, H305	JRH
		Skin Dissection Guidelines, MO03	JRH
		Gastrointestinal Dissection Guidelines, MO07	JRH
		Urological Dissection Guidelines, MO08	JRH
		Breast Dissection Guidelines, MO09	JRH
Cardio-Respiratory Dissection Guidelines, MO10 Head and Neck Dissection Guidelines, MO11	JRH		
Haematological Dissection Guidelines, MO13 Endocrine Dissection Guidelines, MO15	JRH		



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HUMAN TISSUES (cont'd)	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
		Cut up pf Gynaecological Specimens, MO16	JRH
		Use of Circular Saw, MO43	JRH
		Gynaecological Dissection Guidelines for BMS's, MO49	JRH
Benign soft tissue tumour Malignant soft tissue tumour Other soft tissue Bone Bone tumours	Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis	<u>Macroanalysis, dissection &amp; decalcification</u> using:  NOC Specimen reception and handling SOP0001  NOC Specimen Preparation for Dissection, Assisting with Cut-up, and dissection using the Dragon voice recognition system SOP0004  Instructions for the Medoc 250 band saw SOP0014  <u>NOC Instructions for the cassette printer</u> SOP0021  Photography of specimens SOP0057  PathLite-VividPath macro camera  Decalcification of bone specimens SOP0064  NOC Histopathology Dissection Manual HDM0001	NOC
		<u>X-Ray imaging</u> and Hologic PathVision digital Faxitron Reader using: NOC Specimen X-ray SOP0006	NOC



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Fresh Tissue for Frozen techniques	Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis	<u>Frozen Section and Fresh Tissue Handling H010</u>	JRH
		<u>NOC Preparation of frozen sections for urgent diagnosis and use of cryostats SOP0003</u>	NOC
		<u>Sectioning &amp; slide production</u> using: Leica Cryostats, CM1950 & CM1860 UV	JRH & NOC
Frozen Tissue Tissue on Glass Slides	Stains to detect the following:  Basophilic and eosinophilic tissue structures	Cryostat Use and Maintenance, H201	JRH
		Immunofluorescence Cutting and Staining, H040	JRH
		Manual H&E Method and Quality Monitoring of H&E Stain H171	JRH
		H&E staining Manual method using: SSP0014 (H&E)	NOC
Frozen sections of fresh tissue	Special stains to detect the following:  Ganglion cells  Neutral lipids  Alkaline Phosphatase	<u>Special staining</u> Manual methods using procedures: Special Stains Workload Organisation and Quality Control Procedure H264 and stains as appropriate:	JRH
		ACHE (Acetylcholinesterase) Staining Method H194	JRH
		Oil red O Technique H092	JRH
		NOC ALP SOP0005	NOC
		Immunohistochemistry to detect the following:	Immunofluorescence Cutting and Staining SOP H040
Frozen sections of fresh tissue	Complement factor (fluorescent)  Complement factor (fluorescent)  Fibrinogen (fluorescent)	C1Q FITC	JRH
		C3 FITC	JRH
		Fibrinogen FITC	JRH



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<p>HUMAN TISSUES (cont'd)</p> <p>Tissues: biopsy, excision, resection and post-mortem specimens</p> <p>Benign soft tissue tumour Malignant soft tissue tumour Other soft tissue Bone Bone tumours</p>	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):		
	Immunoglobulin (fluorescent)	IgA FITC	JRH	
	Immunoglobulin (fluorescent)	IgG FITC	JRH	
	Immunoglobulin (fluorescent)	IgM FITC	JRH	
	Light chain (fluorescent)	Kappa FITC	JRH	
	Light chain (fluorescent)	Lambda FITC	JRH	
		<u>Processing</u> using: Sakura VIP: Processors: Use & Maintenance of the Tissue-Tek Vacuum Infiltration Processor 5, H159		
		Use and Maintenance of Tissue-Tek VIP6, H273	JRH	
		Leica Peloris Processors : Use and Maintenance of the Leica Peloris Tissue Processor, H288	JRH	
		Use and Maintenance of the Leica Peloris III, H301	JRH	
	<u>Processing</u> using: <u>VIP automated tissue processors:</u>  <u>NOC Instructions on how to use the Sakura Tissue-Tek VIP6 tissue processor SOP0022</u>		NOC	



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HUMAN TISSUES (cont'd)	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
Tissues: biopsy, excision, resection and post-mortem specimens		<u>Embedding</u> using: Thermoshandon Histostar & Leica embedding centres, Leica Histocore Arcadia embedding centre. Tissue Embedding, Daily Worksheet and Block Organisation, H021	JRH
Benign soft tissue tumour Malignant soft tissue tumour Other soft tissue Bone Bone tumours		<u>Embedding</u> using: Leica Arcadia Embedding station  NOC Embedding SOP0007	NOC
Formalin Fixed Paraffin wax Embedded tissue (FFPE)	Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis	<u>Sectioning &amp; slide production</u> using: Leica Rotary Microtomes, RM2135, RM2235 & RM2255 Leica Histocore Biocut microtomes: Microtomy H022  NOC Microtomy SOP0008	JRH  NOC
FFPE sections on glass slides	<u>Stains to detect the following:</u> Basophilic and eosinophilic tissue structures	Automated Methods Routine Haemotoxylin and Eosin (H&E) staining & coverslipping Use and Maintenance of Tissue Tek Prisma™ Automated Staining Machine, H272 Use and maintenance of the Tissue Tek Glas g2 automated coverslipper: CPSOP32	JRH



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HUMAN TISSUES (cont'd)	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
Benign soft tissue tumour Malignant soft tissue tumour Other soft tissue Bone Bone tumours		Instructions for the DiaPath Giotto Autostainer SOP0025 NOC Haematoxylin and Eosin SSP0014 Clearvue coverslipper NOC Coverslipping glass slides SOP0023	NOC
FFPE sections on glass slides	<u>Special stains to detect:</u>	Automated staining method	
	Dermal Acidic Mucins	Alcian Blue H&E Technique, H136	JRH
	Acidic mucins, PAS positive substances (neutral mucins, glycogen) and mixed mucins	Alcian Blue Periodic Acid Schiff's (ABPAS) Technique, H054	JRH
	Acidic mucins, PAS positive substances (not glycogen) and mixed mucins	Diastase Alcian Blue Periodic Acid Schiff's (DABPAS) Technique, H296	JRH
	Glycogen evidenced by loss of staining after diastase, Selective glycogen staining	Diastase Periodic Acid Schiff's (DPAS) Technique, H097	JRH
		NOC Periodic Acid Schiff's (PAS/Diastase PAS (DPAS)/ PAS Fungi (PASF)) SSP0026	NOC
	Mucins, Glycogen, Basal Lamina, PAS positive substances	Renal Periodic Acid Schiff's (RPAS) Technique, H098	JRH
	<i>Helicobacter pylori</i>	Toluidine Blue for <i>H. Pylori</i> , H071	JRH
FFPE, fresh and frozen sections on glass slides	<u>Stains to detect the following:</u>	Manual staining I methods using procedures:	
	Basophilic and eosinophilic tissue structures	Manual H&E Method and Quality Monitoring of H&E Stain, H171	JRH
		Manual Coverslipping & Repairing Slides, H027	JRH



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HUMAN TISSUES (cont'd)	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
	Acidic mucins	Manual Special Stain methods: Alcian blue (AB) H053	JRH
	Acidic mucins, PAS positive substances, (neutral mucins, glycogen) and mixed mucins	Alcian Blue Periodic Acid Schiffs (ABPAS) H054	JRH
	Amyloid	Congo red method H174	JRH
		SSP0004	NOC
	Alcidic mucins, PAS positive substances (not glycogen) and mixed mucins	Diastase Alcian blue Periodic Acid Schiffs (DABPAS) H296	JRH
	Glycogen evidenced by loss of staining after diastase, selective glycogen staining	Diastase Periodic Acid Schiff's (DPAS) Technique H097	JRH
	Dermal acidic mucins	Alcian blue haematoxylin for skin H136	JRH
	Elastic, Collagen, RBCs & other tissue elements	Elastic van Gieson (EVG) H087	JRH
		NOC Millers Elastic van Gieson SSP0022	NOC
FFPE, fresh and frozen sections on glass slides (cont'd)	Bile pigment, muscle / cytoplasm and collagen	Fouchet - Technique, H063	JRH
		Staining manual methods using procedures:	
	Collagen	Fast green - Sirius Red Technique, H133	JRH
	Haemopoietic cells, Bone marrow cells	Giemsa method H064	JRH



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<p>HUMAN TISSUES (cont'd)</p> <p>FFPE, fresh and frozen sections on glass slides (cont'd)</p> <p>FFPE sections on glass slides</p>	<p><u>Histopathology</u> (cont'd)</p>	<p>Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):</p>	
	<p>Staining manual methods using procedures (cont'd):</p>		
	<p>Mineralised bone, osteoclast/osteoblast activity</p>	<p>Goldner's Trichrome SSP010</p>	<p>NOC</p>
	<p>Gram positive &amp; negative microorganisms</p>	<p>Gram's stain HO69</p> <p>NOC Gram Stain SSP0011</p>	<p>JRH</p> <p>NOC</p>
	<p>Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis</p>		
	<p><u>Special stains to detect the following:</u></p>	<p>Special staining Manual methods:</p>	
	<p>Reticulin fibres, collagen</p>	<p>Reticulin stain (Gordon &amp; Sweets technique) H068</p>	<p>JRH</p>
	<p>Fungi, <i>Pneumocystis carinii</i> / <i>jirovecii</i></p>	<p>Grocott's Hexamine Silver H075</p> <p>NOC Grocott SSP0013</p>	<p>JRH</p> <p>NOC</p>
	<p>Collagen, Muscle, RBCs</p>	<p>Haematoxylin Van Gieson H077</p>	<p>JRH</p>
	<p>Glomerular Basement membranes, basal lamina</p>	<p>Jones Hexamine silver technique H079</p>	<p>JRH</p>
<p>Fibrin, Muscle, Collagen, RBCs</p>	<p>Martius Scarlet Blue (MSB) technique H089</p>	<p>JRH</p>	
<p>Collagen, connective tissue, muscle, RBCs, Fibrin</p>	<p>Masson's trichrome technique H086</p>	<p>JRH</p>	
<p>Melanin, Argentaffin, Chromaffin, Lipofuscin</p>	<p>Masson Fontana and melanin bleach H084</p>	<p>JRH</p>	
		<p>NOC Massons Fontana SSP0019</p>	<p>NOC</p>



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HUMAN TISSUES (cont'd)  FFPE sections on glass slides (cont'd)	<u>Histopathology</u> (cont'd)  Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
	<u>Special stains to detect the following:</u> (cont'd)  Hepatitis B surface antigen (Hep BsAg),, Copper Associated Proteins (CAP) & Elastin	Special staining Manual methods  Orcein shikata stain H094	JRH
	Mucins, Glycogen, PAS positive substances – (such as fungal hyphae)	Periodic Acid Schiffs (PAS) Technique H096  SSP0026	JRH  NOC
	Glycogen evidenced by loss of staining after diastase	Diastase Periodic Acid Schiffs (DPAS) technique H097	JRH
	Haemosiderin, Ferric iron, Ferruginous bodies	Perls Prussian Blue H099  NOC Perls Technique SSP0027	JRH  NOC
	Basal lamina, PAS positive substances	Renal Periodic Acid Schiffs (RPAS) techniques (H098)	JRH
	<i>Helicobacter pylori</i>	Toluidine blue for Helicobacter Pylori H071	JRH
	Cartilage and mast cells	Toluidine blue SSP0033	NOC
	Calcium	Von Kossa SSP0035	NOC
	Acid fast bacilli – <i>mycobacterium leprae</i>	Wade-Fite technique H120	JRH
	Acid fast bacilli <i>mycobacterium sp</i>	Ziehl Neelsen technique H113  NOC Ziehl Neelsen (ZN) for TB SSP0037	JRH  NOC



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HUMAN TISSUES (cont'd)	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
FFPE sections on glass slides	<u>Special stains to detect the following:</u>  Acidic mucins, neutral mucins, PAS positive substances  Acidic mucins, PAS positive substances  Glycogen evidenced by loss of staining after Diastase	Special staining Automated methods  Alcian Blue Periodic Acid Schiffs (PAS)  Diastase Alcian blue PAS  Diastase PAS  Immunohistochemistry Workload Organisation H235  Anitbody Dilution and Optimisation, H151	JRH  JRH  JRH  JRH & NOC
Fixed, fresh and frozen tissue	Target antigens in Human Tissues and fluids	Flexible scope protocols: CPP40 Flexible Scope of Accreditation Management  Bond III staining machine H236  Ventana BenchMark ULTRA staining machine H295	JRH & NOC



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<p>HUMAN TISSUES (cont'd)</p> <p>FFPE sections on glass slides</p>	<p><u>Histopathology</u> (cont'd)</p> <p>Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</p> <p>In-situ hybridisation to detect the following:</p> <p>Epstein Barr virus</p> <p>Lambda light chain</p> <p>Kappa light chain</p>	<p>Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):</p> <p><u>In-situ hybridisation</u> Automated using procedures: H236 and Leica Bond III and probes as appropriate:</p> <p>EBER</p> <p>Lambda</p> <p>Kappa</p>	<p>JRH</p> <p>JRH</p> <p>JRH</p> <p>JRH</p>



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HUMAN TISSUES (cont'd)	<u>Histopathology</u> (cont'd) Examination to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
Head and neck tissues	In-situ hybridisation to detect the following:	Use of Bond III staining machine H236	JRH
	High risk human papilloma virus genotypes	RNAscope ISH	JRH
Breast and Gastrointestinal tissues	Dual colour, dual hapten brightfield in situ hybridisation (DDISH) to detect:	Use of Ventana BenchMark ULTRA Staining Machine, H295	JRH
	Human Epidermal growth factor Receptor 2 (HER2), breast and gastrointestinal cancers	HER2	JRH
Slides prepared in house as above	Morphological assessment and interpretation/diagnosis	<u>Microscopy (qualitative analysis)</u> Using procedures: CPSOP27 in conjunction with manufacturer's instructions using microscopes Leica DM4000B, Leitz Dialux, Labourlux K, Laborlux S, Orthoplan, Orthomat II, Nikon Eclipse 50i, Ci-L, E200, E400 & E600, Optishot Olympus BH2, BHTU, BH-U, BHS312, BHS313, BX40, BX41, BX50, BX51, BX53 and microscopes listed above	JRH & NOC
		<u>NOC sending out slides to the Pathologists SOP0011</u>	NOC



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Digital images from slides prepared in house as above	Morphological assessment and interpretation/diagnosis	Using digital pathology (slide scanners and viewing software):  Philips Intellisite Digital Pathology Suite, H307 Use and Maintenance of Philips™ Slide Scanners: <ul style="list-style-type: none"> <li>• Philips UFS Ultra Fast Whole Slide Imaging Scanner</li> <li>• Philips UVS Ultra Versatile Whole Slide Imaging scanner</li> <li>• Philips IMS Image Management and Storage System</li> </ul>	JRH & NOC
		Slide Scanning Use_and_Maintenance_of _Philips_Slide_Scanners H307	JRH
		Slide Scanning using Philips SG60 Pathology Scanner	NOC
		Slide Reporting Histodata entry and authorisation using approved equipment IT05	JRH & NOC

END