# **Schedule of Accreditation**

issued by

# **United Kingdom Accreditation Service**

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK



8840

Accredited to ISO 15189:2022

# **Cambridge University Hospitals NHS Foundation Trust**

Issue No: 012 Issue date: 27 November 2025

Cambridge Genomics Laboratory Cambridge University Hospitals NHS Foundation Trust

**Cambridge Biomedical Campus** 

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#### Testing performed at the above address only

#### **DETAIL OF ACCREDITATION**

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND BODY FLUIDS	Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer	Documented in house procedures incorporating manufacturer's instructions (where relevant)
	Sample preparation, DNA extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long term storage	Sample preparation:
Whole Blood Bone Marrow		Red Cell lysis and storage in Qiagen Nucleon B, RLT or Tri- Reagent CU-EX-SOP-34, CU-EX-SOP-23, CU-EX-SOP-5 and CU-EX-SOP-8
Whole Blood		Separation of mononuclear cells from peripheral blood or bone marrow using Lymphoprep CU-EX-SOP-31
		Manual and automated DNA extraction and quantification using:
Whole Blood		Automated Extraction: Automated DNA extraction using Flex Star
		CU-EX-SOP-13

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HUMAN TISSUES AND BODY FLUIDS (cont'd)	Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Whole Blood,Buffy Coat CVS/Amniocentesis Tissues		Automated DNA extraction using Qiagen EZ1
Guthrie Spots		CU-EX-SOP-9
Whole Blood Bone Marrow Body Fluid		Automated DNA extraction using QIACube
Tissue Lineage specific fractions		CU-EX-SOP-3 CU-EX-SOP-5
		Manual extraction:
Body fluid and human tissue (fresh, frozen or fixed) Formalin fixed parrafin embedded tissue Bone Marrow		Manual DNA extraction usingQiagen DNEASY kit CU-EX-SOP-15 CU-EX-SOP-16 CU-EX-SOP-23
Bone Marrow		Extraction of DNA from DNA lysis buffer (Q-AL or nucleon B) CU-EX-SOP-22
Saliva and Buccal swabs		Manual DNA extraction using- Oragene Purifyer CU-EX-SOP-11
Genomic DNA extracted in-house from the sample types listed above		DNA Quantification for QC purposes:
or received as primary sample type from an external source		Nanodrop ND-1000 spectrophotometer and Qubit fluorometer
		CU-EQ-SOP-9 CU-EX-SOP-21

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HUMAN TISSUES AND BODY FLUIDS (cont'd)	Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)
DNA Clean up		Qiagen QiaAmp DNA micro kit CU-EX-SOP12
Whole Blood, bone marrow, body fluid and human tissue (fresh, frozen or fixed)		Automated Extraction:  Automated RNA extraction and conversion to cDNA using QIACube, NJ Research (Biorad) thermal cycler and Syngene Genius Bioimaging System  CU-EX-SOP-3  CU-EX-SOP-19  CU-EX-SOP-32
Whole Blood, bone marrow, body fluid and human tissue (fresh, frozen or fixed)	RNA extraction and preparation of cDNA, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long term storage	Manual and automated RNA extraction, conversion to cDNA and quantification using:  Manual Extraction  Extraction of RNA from RLT Buffer CU-EX-SOP-24

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HUMAN TISSUES AND BODY FLUIDS (cont'd)	Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA and cDNA extracted in-house from the sample types listed above or received as primary sample type from an external source	Detection of nucleic acid sequence variants - SNVs and small indels-	Sanger Sequencing Sanger Sequencing Analysis using Mutation Surveyor by SoftGenetics CU-TS-SOP-29
Genomic DNA and cDNA received as primary sample type from an	Detection of nucleic acid sequence variants - SNVs and small indels-	Sanger Sequencing Using:
external source	See CU-UK-DTA-3	Standard Primer Design methodology: CU-BI-SOP-2
		PCR/ Thermal cyclers, ABI 3730, Biomek robotics
		CU-CM-SOP-5 CU-CM-SOP-13 CU-TS-SOP-28 CU-CM-SOP-31 CU-TS-SOP-45 CU-TS-SOP-65 CU-TS-SOP-58, CU-TS-SOP-62
Illumina TriSight One data previously generated in-house	SNVs and small indels See CU-UK-DTA-7	Analsyis using Dias Bioinformatics software CU-BI-MAN-1
Genomic DNA received as primary sample type from an external source	Next Generation Sequencing for Small Variants	TWIST WES Comprehensive exome panel CU-NG-SOP-8 CU-BI-MAN-2 CU-BI-MAN-1 CU-BI-MAN-6 CU-NG-SOP-14 CU-NG-SOP-10 CU-RP-SOP-9

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Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Next Generation Sequencing	TWIST Core, Endocrine, Neurology (CEN) CU-NG-SOP-14 CU-BI-MAN-1 CU-NG-SOP-8 CU-BI-MAN-2 CU-RP-SOP-9
Nucleic acid extraction	Manual DNA extraction using Tissue Lyser and Qiagen Mini kit CU-EX-SOP-17 CU-TS-MAN-6
Detection of SNVs and small indels	Real Time PCR
See CU-UK-DTA-8	Using Applied biosystems Realtime 7500 analyser
	CU-CM-SOP-21
	Using Rotogene 6000 series – QIAgen CU-CM-SOP-16 CU-CM-SOP-26 CU-CM-SOP-22
Sample preparation, DNA and RNA extraction, quantification and quality check for subsequent in-house analysis, referral to specialist centres and long term storage.	Manual or Automated Extraction of nucleic acids from formalin fixed paraffin embedded (FFPE) tissue using MagMax CU-EX-SOP-33
	Measured/Range of measurement  Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)  Next Generation Sequencing  Nucleic acid extraction  Detection of SNVs and small indels  See CU-UK-DTA-8  Sample preparation, DNA and RNA extraction, quantification and quality check for subsequent in-house analysis, referral to specialist

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Materials/Products tested	measured/Range of measurement	Equipment/Techniques used	
HUMAN TISSUES AND BODY FLUIDS (cont'd)	Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)	
Genomic DNA received as primary sample type from an external source	Determination of fragment length size and detection of deletions, fusion deletions, known SNVs and indels, microsatellites, repeat expansion, linkage markers for the	size and detection of deletions, fusion deletions, known SNVs and indels, microsatellites, repeat expansion, linkage markers for the	Using:
	purpose of genotyping at specific known gene loci  See CU-UK-DTA-8	Fluorescent Fragment Analysis using ABI 3730 and ABI 3500 xl genetic analysers and GeneMarker software	
		CU-TS-SOP-34	
		Commercial Methods: Fluorescent Fragment Assays:	
		Fluorescent ARMS using the Elucigene kits CU-TS-SOP-17 and CU-TS-SOP-9	
		QF-PCR kit (QF-PCR) - Devyser Kits CU-TS-SOP-44 CU-TS-SOP-43	
		And:	
		In house Fluorescent Fragment Assays using PCR and allele specific PCR and PCR of linked markers	
		CU-CM-SOP-14 CU-TS-SOP-21 CU-TS-SOP-23 CU-TS-SOP-30 CU-TS-SOP-36 CU-TS-SOP-41	
		Fluorescent Fragment Analysis using and 3730, 3500 genetic analysers and GeneMarker or Genemapper software	
		CU-TS-SOP-34 and CU-RP-SOP-29	

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HUMAN TISSUES AND BODY FLUIDS (cont'd)	Genomic analysis for the purpose of clinical diagnosis of rare and inherited disease and cancer (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA received as primary sample type from an external source	Determination of fragment size length, Detection of deletions, fusion deletions, known SNVs, repeat expansions and inversions for the purpose of genotyping at specific known gene loci -  See CU-UK-DTA-4	Gel electrophoresis based Fragment Analysis In house methods for: Long range PCR, MS-PCR, inverse shifiting PCR, multiplex PCR followed by gel electophoresis.  SOP: CU-CM-SOP-15 CU-CM-SOP-44 CU-TS-SOP-23
Genomic DNA received as primary sample type from an external source	Detection of large repeat expansions See CU-UK-DTA-6	In house methods for: Southern blot hybridization analysis using labelled probes.  CU-TS-SOP-20 CU-TS-SOP-7
Genomic DNA extracted in-house from human body fluids and tissues or received as primary sample type from an external source	Single Nucleotide Polymorphism (SNP) genotyping (C9ORF72 gene)	Nimagen SNP profile kit for identity checking on ABI 3500 and ABI 3730 genetic instruments  CU-WG-SOP-14
Genomic DNA extracted in-house from human body fluids and tissues or received as primary sample type from an external source	Determination of fragment length size and detection of deletions, fusion deletions, known SNVs and indels, microsatellites, repeat expansion, linkage markers for the purpose of genotyping at specific known gene loci in gene C9ORF72.	Fluorescent fragment analysis using ABI 3500 and ABI 3730 genetic instruments: Frontotemporal Dementia (FTD) and/or Amyotrophic Lateral Sclerosis (ALS) (C9ORF72 gene)  CU-TS-SOP-34 CU-RP-SOP-33

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HUMAN TISSUES AND BODY FLUIDS (cont'd)	Cytogenetic examinations, neoplastic genetics including haemato-oncology, - detection of sub microscopic chromosomal imbalance (gains and losses) expressed as changes to copy number and copy neutral loss/absence of heterozygosity	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Sequencing data received from internal and external sources	Detection of genomic variation with potential clinical utility, including small variants (SNV & indel) and structural variants (CNV, insertion, inversions and translocations resulting in fusion, ITD and PTD)	Detection of variants for clinical reporting of patients with rare disease and cancer: alignment and variant calling. GH-BI-POL-1, GH-BI-POL-2, GH-BI-POL-
Whole blood Amniotic fluid Chorionic villus samples Bone Marrow Biopsy Material Body Fluids	G-Banding/Karyotyping  Detection of chromosomal rearrangements or aberrations arising from:  Prenatally detected Disorders Developmental disorders Reproductive medicine disorders Haematological/Oncology disorders  (Preparative Pre-examination steps listed first)	Manual culturing and processing of human tissue to provide metaphase cells: using commercial media and:  CU-CT-SOP-2. CU-CT-SOP-22, CU-CT-SOP-8, CU-CT-SOP-14, CU-CT-SOP-17, CU-CT-SOP-14, CU-CT-SOP-20  Manual harvesting using in house methods: CU-CT-SOP-3 (whole blood and prenatal and tissue cultures CU-CT-SOP-7). CU-CT-SOP-13  Slide Preparation, coverslipping and storage: CU-CT-SOP-4 and CU-CT-SOP-19  Microscopic Analysis and reporting of G banded chromosomes: Using: Brightfield microscopes and GenASIs image capture and anlaysis software  CU-CT-SOP-25, CU-CT-SOP-26, CU-CT-SOP-27, CU-RP-SOP-38, CU-CT-SOP-15 and CU-RP-SOP-21, CU-RP-SOP-27

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HUMAN TISSUES AND BODY FLUIDS (cont'd)	Cytogenetic examinations (cont'd)	Documented in house procedures incorporating manufacturer's instructions (where relevant)
Genomic DNA received as primary samples from external sources	Detection of pathogenic and likely pathogenic DNA copy number variants and copy number neutral loss of heterozygosity (in diagnostic samples)	SNP Array Using Infinium Global Screening Array (GSA) v 3 kit with Illumina Infinium HTS Assay and Tecan Robot Analysis using Illumina iScanner with Illumina Genome Studio software and NxClinical software CU-MA-SOP-1 CU-RP-SOP-4
Genomic DNA received as primary sample type from an external source	Detection of whole exon deletions/duplications and specific methylation abnormalities	Multiplex Ligation Probe Analysis (MLPA) and methylation specific MS-MLPA
	See CU-UK-DTA-5	using: in house or commercial MRC Holland kits
		Thermal cyclers and 3500 / 3730 Genetic Analysers
		Analysis using GeneMarker
		CU-TS-SOP-37 CU-TS-SOP-16 CU-TS-SOP-42
Fixed cell slides from human tissues and body fluids:	Detection and analysis of genomic rearrangements and imbalances. Confirmation of genomic	Fluorescent in-situ hybridisation (FISH)
Amnoitic fluid	rearrangements - using	Using:
CVS Stemcells Wholeblood Bone Marrow CSF	locus specific probes:  Deletion Copy Number/Amplification	Cytocell and Vysis commercial kits, Vysis HYBrite hotplate and applied imaging (Leica)
Biopsy Material Body fluids FFPE Tissue	See CU-UK-DTA-2	Analysis using: Cytovision analysis software and and fluorescent microscope system
		CU-FI-SOP-6, CU-FI-SOP-5, CU-FI-SOP-12, CU-FI-SOP-11, CU-FI-SOP-4, and CU-FI-SOP-8
END		

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