

# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK



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ISO 15189:2022

### Shrewsbury and Telford Hospital NHS Trust

Issue No: 007 Issue date: 10 June 2025

Department of Pathology  
Royal Shrewsbury Hospital  
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Shrewsbury  
SY3 8XQ

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#### Locations covered by the organisation and their relevant activities

##### Laboratory locations:

Location details: Department of Cellular Pathology		Activity
<b>Address:</b> Department of Cellular Pathology Royal Shrewsbury Hospital Mytton Oak Road Shrewsbury SY3 8XQ	<b>Local contact</b> Victoria Banton	Histopathology Rapid frozen section for intra-operative diagnosis Diagnostic Cytopathology

##### On / off site activities performed away from the locations listed above:

Location details		Activity
Mortuary Royal Shrewsbury	Victoria Banton (as above)	Body Store and Mortuary
Mortuary Princess Royal Hospital	Victoria Banton (as above)	Body Store
Radiology Head & Neck Clinic Princess Royal Hospital	Victoria Banton (as above)	FNA material/slide preparation



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
<p><b>HUMAN TISSUES AND FLUIDS</b></p> <p>Tissue samples: Formalin fixed, fresh and frozen, including Bone Marrow trephines Non-gynae cytology cell blocks</p>	<p><u>Histopathology</u></p> <p>Preparation and examination of tissue samples to identify or exclude morphological &amp; cytological abnormalities</p> <p>Specimen dissection and biopsy transfer</p> <p>Decalcification of bony samples prior to histopathological examination</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p><u>Macroanalysis and dissection</u> Manual method using: HISurgCutUp HISBMSCutUp HISCutUP CPPrintMate TURPSampling ThermoFisher PrintMate</p> <p><u>Decalcification</u> Manual method using: HISDecal</p> <p><u>Tissue Processing</u> Automated method using: HISProcessing HISExcelcior and: Thermo Fisher Excelcior</p> <p><u>Embedding</u> Manual method using: HISEmbed and: Sakura Tissue Tek V Thermo Scientific Histostar Embedding centre</p> <p><u>Microtomy</u> Manual method using: HISMicrotomy HISMegaBlock HISSpares and: Leica RM2235</p>



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology</u> (cont'd)  Preparation and examination of tissue samples to identify or exclude morphological and cytological abnormalities (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)
Frozen tissue	Identification of nuclei, cell cytoplasm and connective tissue in cells and tissues for the purposes of clinical diagnosis using Haematoxylin and Eosin	<u>Rapid frozen section</u> Cryotomy and manual staining method using:  HISStainRapidHE and: Leica CM 1860
Fresh tissue: Renal	Attendance at ultrasound guided renal biopsy to confirm adequacy of sample and ensure use of appropriate fixative	<u>Renal Biopsy Procedure</u> CPRenalBX using Wild Heerbugg M32 Dissecting Microscope
Formalin fixed, paraffin embedded (FFPE) tissue	Identification of nuclei, cell cytoplasm and connective tissue in cells and tissues for the purposes of clinical diagnosis using Haematoxylin and Eosin	<u>Routine Haematoxylin &amp; Eosin (H&amp;E) staining and coverslipping</u> Automated method using: HISAutoStain HISCoverslip and: Leica Autostainer XL Leica Coverslipper CV5030
Formalin fixed, paraffin embedded (FFPE) tissue	Detection of the following:  Acid and sulphated mucins  Acid and neutral mucins  Neutral mucins and fungi  Differentiating between haemopoietic cells	<u>Special staining</u> Manual methods using: HISSpecial CPSSControls and the following stains:  Alcian Blue Technique using HISStainAB  (Diastase) Alcian Blue PAS Using HISStainABPAS  (Diastase) Periodic Acid Schiff's HISStainDPAS  Giemsa HISStainGIEM



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FFPE tissue (cont'd)	Detection of the following: (cont'd)  Amyloid  <i>Helicobacter pylori</i> identification  Differentiating between Gram positive and Gram negative microorganisms  Fungi  All tissue structures  Melanin (and removal of pigment prior to staining)  Collagen and smooth muscle  Connective tissue and fibrin  Elastic fibres  Copper associated protein  Basement membranes	<u>Special staining</u> (cont'd)  Congo Red HISStainCR  Giemsa for Helicobacter HISStainGiemsaHP  Gram with Twort Counterstain for Bacteria HISStainGrmTwt  Grocott's Method for Fungi HISStainGroc  Haematoxylin and Eosin Stain HISStainHE  Mallory Bleach for Melanin HISStainMalBleach and Masson Fontana for Melanin HISStainMasFont  Masson Trichrome HISStainMasTri  Martius, Scarlet, Blue (MSB) HISStainMSB  Miller's Elastin Stain HISStainMillers  Orcein for Copper Associated Protein HISStainOrcein  PA Silver HISStainPASilver



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FFPE tissue (cont'd)	Detection of the following: (cont'd)  Haemosiderin  Reticulin fibres  Mast cells  Connective tissue  Calcium salts  Atypical Mycobacterium  Spirochaetes  Acid fast bacilli, e.g. <i>M. tuberculosis</i>	<u>Special staining</u> (cont'd)  Perls Prussian Blue for Ferric Iron HISStainPerls  Gordon & Sweet's Method for Reticulin HISStainRetic  Toluidine Blue for Mast Cell Granules HISStainTolBlue  Van Gieson Method HISStainVG  Von Kossa HISStainVKossa  Wade Fite HISStainWF  Warthin- Starry for Spirochaetes HISStainWarStar  ZN Stain for Acid Fast Bacilli Using HISStainZN



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FFPE tissue	Detection of the following:  Pan cytokeratin Germ tumours Anaplastic lymphoma kinase Prostate grading Colon adenocarcinoma Mantle zone B lymphocytes, interfollicular T lymphocytes Germinal centre B lymphocytes Epithelial cell types, basal cell carcinoma Malignant epithelial cells Glandular cells Parafollicular C cells thyroid Mesothelioma marker Langerhan's cells, interdigitating reticulum cells T lymphocytes	<u>Immunohistochemistry</u> Automated methods using: IPBondMax and IPPrimaryABS and: Leica Bond III Immunostainer and the following antibodies  AE1/AE3 AFP ALK-1 AMACR B-CATENIN Bcl-2 Bcl-6 Ber-EP4 CA125 CAM 5.2 Calcitonin Calretinin CD1a CD3



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FFPE tissue (cont'd)	Immunohistochemistry to detect the following: (cont'd)  T lymphocytes (helper and inducer)  Germinal centre T lymphocytes, some B  Suppressor/ cytotoxic T cells  Mature B lymphocytes, renal cell carcinoma  Hodgkin and Reed Sternberg  B lymphocytes  B lymphocytes, some T lymphocytes  Hodgkin and Reed Sternberg cells. Plasma cells  Endothelial cells, plasma cells, activated B and T cells  Endothelial cells, fibroblasts, haemopoietic precursor cells  Peripheral nerves, NK cells, neuroendocrine differentiation  Macrophages, monocytes, granulocytes  Mast cells, marker for GIST	<u>Immunohistochemistry</u> (cont'd)  CD4  CD5  CD8  CD10  CD15  CD20  CD23  CD30  CD31  CD34  CD56  CD68  CD117



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FFPE tissue (cont'd)	Immunohistochemistry to detect the following: (cont'd)  Plasma cells, differentiation of B lymphocytes  Intestinal epithelial cells  Adenocarcinoma, epithelial cells of the glandular lumen  Neuroendocrine cells and tumours  Basal cells, stratified squamous epithelia  Simple glandular and transitional epithelia  Epithelia, thyroid papillary carcinoma  Gastrointestinal epithelia  Cytomegalovirus  Mantle cell lymphoma  Normal & Abnormal epithelial cells  Smooth and striated muscle cells  Normal duct epithelial cells, ductal carcinoma	<u>Immunohistochemistry</u> (cont'd)  CD138  CDX2  CEA  Chromogranin A  CK5/6  CK7  CK19  CK20  CMV  Cyclin D1  Cytokeratin HMW (HMWCK)  Desmin  E-Cadherin





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FFPE tissue (cont'd)	Immunohistochemistry to detect the following: (cont'd)  Glandular and ductal epithelia  Cell nuclei expressing oestrogen, normal glands and neoplastic cells  Papillary carcinoma differentiation  Identifies breast carcinomas and urothelial carcinomas  Placental trophoblasts  Melanoma, melanocyte differentiation  Sertoli cells, sex chord stromal tumour  Marker of proliferation  B and T lymphocytes  Normal, activated and neoplastic melanocytes  Multiple myeloma oncogene  Thyroid, renal, Mullerian and thymic tumours  Normal proliferating cells, basal cells	<u>Immunohistochemistry</u> (cont'd)  EMA  ER  Galactin-3  GATA-3  HCG  HMB45  Inhibin  Ki67  LCA  Melan-A  MUM-1  PAX8  p63



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FFPE tissue (cont'd)	Immunohistochemistry to detect the following: (cont'd)  Germ cell tumours  Nuclei of cells showing progesterone receptor expression  Prostate secretory and ductal epithelia  Schwann cells, nerve processes, normal and neoplastic melanocytes  Smooth muscle and myoepithelial cells  Smooth muscle and myoepithelial cells  Neuroendocrine cells  Thyroglobulin containing cells  Thyroid transcription factor-1, lung tumours  Cells of mesenchymal origin  Epithelial cells and serous ovarian adenocarcinoma	<u>Immunohistochemistry</u> (cont'd)  PLAP  PR  PSA  S100  SMA  SMM  Synaptophysin  Thyroglobulin  TTF-1  Vimentin  WT-1



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Slides prepared in-house from sample types listed above	Morphological assessment and interpretation/diagnosis	<u>Microscopy (qualitative analysis)</u> In-house procedures: CPConsReporting CPLightMicroSOP using: Laborlux S Leica DM2000 Leica DM3000 Leica Diaplan Nikon Eclipse NiU 401 Nikon Eclipse 80i Nikon Eclipse E400 Olympus BX51
Fresh, frozen tissue of renal, skin and mucosal biopsies	<u>Direct Immunofluorescence</u>  Identification of antibodies in various autoimmune diseases	Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)
	Gamma chains of human IgG	<u>Immunofluorescence and fluorescent microscopy</u> Manual in house procedure using: DIFProcedure DIFPipette  IgG/ FITC
	Mu chain of human IgM	IgM/ FITC
	Alpha chains of human IgA	IgA/ FITC
	Native C3	C3/ FITC
	Conjugated antibodies	C1q/FITC



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<p><b>HUMAN TISSUES AND FLUIDS (cont'd)</b></p> <p>Slides prepared in-house from sample types listed above</p> <p>Body fluids: Cerebrospinal fluid (CSF) Urine General body fluids Bronchial brushings &amp; washings Sputum</p>	<p><u>Histopathology (cont'd)</u></p> <p>Preparation and examination of tissue samples to identify or exclude morphological and cytological abnormalities (cont'd)</p> <p>Morphological assessment and interpretation/diagnosis</p> <p><u>Examination of non-gynaecological specimens for the purpose of clinical diagnosis</u></p> <p>The examination of body fluid/ FNA samples to exclude or identify morphological and cytological abnormalities</p> <p>Staining of slides prepared as above for examination of body fluid/ FNA samples to exclude or identify morphological and cytological abnormalities</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p><u>Fluorescent microscopy (qualitative analysis)</u> Following in-house procedures:</p> <p>DIFPhoto using Leitz Laborlux S with UV lamp housing</p> <p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p>Preparation of direct smears and cytospin slides using : CYTNGSpecs CYTFluids CYTBronch CYTCSF CYTEUS&amp;EBUS CYTUrine CYTSputum CYTHistopaque CYTImmuno and Class II Telstar Bio II-A Microbiological safety cabinet Thermo Fisher Cytospin 4</p> <p>Automated Papanicolaou staining method using : CYTPapStain and Thermo Fisher Raymond Lamb Stainmate Myreva SS-30 staining machine CYTPAPSTAIN</p>



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END		