


# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 8952 Accredited to ISO 15189:2022	<b>Belfast Health and Social Care Trust</b>	
	<b>Issue No:</b> 008 <b>Issue date:</b> 08 April 2025	
	<b>Regional Molecular Diagnostics Service</b> Belfast City Hospital 51 Lisburn Road Belfast BT9 7AB United Kingdom	<b>Contact : Louise McArt</b> <b>Tel : +44(0) 2895 040822</b> <b>E-Mail: <a href="mailto:louise.mcart@belfasttrust.hscni.net">louise.mcart@belfasttrust.hscni.net</a></b> <b>Website: <a href="https://belfasttrust.hscni.net/service/laboratory-services/">https://belfasttrust.hscni.net/service/laboratory-services/</a></b>
Testing performed by the Organisation at the locations specified below		

### Locations covered by the organisation and their relevant activities

#### Laboratory locations:

Location details	Activity	Location code
<b>Address</b> Tower Block, Belfast City Hospital 51 Lisburn Road Belfast BT9 7AB  <b>Local contact</b> Head of Germline Dr Shirley Heggarty <a href="mailto:Shirley.Heggarty@belfasttrust.hscni.net">Shirley.Heggarty@belfasttrust.hscni.net</a> 028 95045264	Regional Molecular Diagnostics Service Germline Somatic	A
<b>Address</b> Health Sciences Building, Queen's University Belfast 97 Lisburn Road Belfast BT9 7AE  <b>Local contact</b> Head of Somatic Dr Mark Catherwood <a href="mailto:Mark.Catherwood@belfasttrust.hscni.net">Mark.Catherwood@belfasttrust.hscni.net</a> 028 95048138	Regional Molecular Diagnostics Service Somatic	B



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**DETAIL OF ACCREDITATION**

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<b>HUMAN BODY FLUIDS / TISSUES</b>  Whole blood Cultured cells CVS Cord blood Buccal cells Products of Conception Fresh / frozen tissue (including skin, muscle, spleen) Guthrie cards Saliva Urine Amniotic Fluid	<u>Molecular Genetics (Germline)</u>  Detection of nucleic acid sequence variants and/or copy number changes for the purpose of clinical diagnosis  DNA extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long-term storage	Documented in-house procedures using techniques and kits in combination with manufacturers instructions	A
		Manual and Automated DNA Extraction and Quantification          Manual Extraction  Using: iGEN Biotech New iGENatal Kit/ Qiagen Gentra Puregene Blood Kit/ DNA Genotek prepIT.L2P Kit  SOPs: LP 352 003 iGENatal LP 364 012 Puregene LP 364 014 prepIT.L2P	A
		Automated Extraction  Using: Promega Maxwell CSC / Perkin Elmer Chemagic MSM1  SOPs: LP 366 051 Maxwell CSCLP 364 020 Chemagic	A
		DNA Quantification  Using: IMPLEN Nanophotometer Pearl/ Invitrogen Qubit 4 Fluorometer  SOPs: LI 366 020 Nanophotometer PD 305 307 Qubit	A



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Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	DNA amplification by PCR for fragment analysis or downstream use in sanger sequencing	DNA Amplification by PCR  Using: In-house methods/ MegaMix Gold/ Eurogentec Primers/ Beckman Coulter Biomek i5 Automated Workstation/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Syngene G:BOX F3 Gel Documentation System  SOPs: LI 362 104 PCR Conditions LP 366 001 Gel Electrophoresis LP 366 013 PCR LP 366 016 G:BOX F3 LP 366 032 Biomek i5	A
Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Detection of nucleic acid sequence variants (SNVs and Indels) in genes specified in definitive list and those identified through research and/or family (trio) NGS sequencing work [Definitive list in RMDS Test Accreditation List held by this laboratory]	Sanger Sequencing  Using: In-house methods/ Applied Biosystems BigDye Terminator Cycle Sequencing Kit/ Beckman Coulter Agencourt AMPure XP PCR Purification System/ Beckman Coulter Agencourt CleanSEQ Dye-Terminator Removal/ Thermocyclers [Definitive list in Current Dept Equipment List/ Beckman Coulter Biomek i5 Automated Workstation/ Applied Biosystems 3500DX DNA Analyzer/ Applied Biosystems 3500 Genetic Analyzer/ SoftGenetics Mutation Surveyor Software/Alamut Visual Plus Software SOPs: LI 362 102 CleanSEQ LI 366 008 AMPure LP 366 008 Sanger Sequencing LP 366 028 Mutation Surveyor LP 366 032 Biomek i5LP 366 038 ABI 3500 LP 366 043 Variant Classification	A



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Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Determination of copy number changes and methylation specific (MS) dosage changes [Definitive list in RMDS Test Accreditation List held by this laboratory]	Multiplex Ligation Probe Amplification (MLPA) (to include MS-MLPA)  Using: MRC Holland SALSA MLPA Reagents and Probemixes/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Applied Biosystems 3500 Genetic Analyzer/ SoftGenetics GeneMarker Software  SOPs: LP 363 026 MS-MLPA LP 366 025 MLPA Set Up LP 366 026 MLPA Analysis LP 366 038 ABI 3500	A
Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Detection of nucleic acid point variants, small insertions and deletions [Definitive list in RMDS Test Accreditation List held by this laboratory]	Amplification Refractory Mutation System (ARMS)  Using: Elucigene CF4 & CF-EU2/ Elucigene DPYD/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Applied Biosystems 3500DX DNA Analyzer/ Applied Biosystems 3500 Genetic Analyzer/ Applied Biosystems GeneMapper Software/ SoftGenetics GeneMarker Software  SOPS: LI 362 081 CF Neonatal Reporting LP 362 083 CF Neonatal LP 363 032 CF Routine LP 363 073 DPYD genotyping LP 366 029 GeneMarker LP 366 034 GeneMapper LP 366 038 ABI 3500	A



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Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Detection of nucleic acid sequence repeat expansions [Definitive list in RMDS Test Accreditation List held by this laboratory]	Repeat Fragment Detection  Using: In-house methods/ MegaMix Gold/ Eurogentec Primers / Asuragen AmplideX PCR/CE FMR1 Kit/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Applied Biosystems 3500DX DNA Analyzer/ Applied Biosystems 3500xL Genetic Analyzer/ Applied Biosystems GeneMapper Software/ SOPs: LP 363 020 Friedreich Ataxia LP 363 021 Huntington Disease LP 363 022 Kennedy Syndrome LP 363 023 Fragile X Syndrome LP 363 025 Myotonic Dystrophy LP 363 039 Dominant Ataxias LP 366 034 GeneMapper LP 366 038 ABI 3500	A
Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Detection of nucleic acid short tandem repeats	Short Tandem Repeat Detection  Using: Promega PowerPlex 16 System/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Applied Biosystems 3500DX DNA Analyzer/ Applied Biosystems 3500 Genetic Analyzer/ Applied Biosystems GeneMapper Software  SOPs: LP 363 036 Promega PowerPlex 16 LP 366 034 GeneMapper LP 366 038 ABI 3500	A



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Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Gene screening of large gene Panels and whole exome/virtual panels for genetic variants: SNVs CNVs Indels Trios [Definitive list in RMDS Test Accreditation List held by this laboratory]	Next Generation Sequencing (NGS)	A
		Library Preparation Using: Roche KAPA HyperPlus Kit/Roche Avenio Edge DNA HyperPlus Kit/Roche KAPA HyperCapture Reagents/Roche Avenio Edge Target Enrichment Reagents/ Hamilton Microlab STAR Liquid Handling System/Roche Avenio Edge System Invitrogen Qubit 4 Fluorometer/ Agilent 4200 TapeStation System/ Roche LightCycler480 Instrument II/ Thermocyclers [Definitive list in Current Dept Equipment List]	A
		SOPs: LF 362 109 Library Quantification LF 366 047 Library Preparation LI 362 109 Library Quantification LP 366 045 Library Preparation (Hamilton) LP 366 052 Library Preparation (Avenio)	
		Next Generation Sequencing Set Up  Using: NextSeq 500/550 High Output Kit/Illumina NextSeq 550DX  SOPs: LF 366 047 Library Preparation LP 306 010 Targeted Cancer Panel LP 363 059 NGS Exome PD 305 305 NextSeq 550DX PD 305 306 NextSeq 500 and NextSeq 550 Libraries Guide	A



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Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Gene screening of large gene Panels and whole exome/virtual panels for genetic variants: SNVs CNVs Indels Trios [Definitive list in RMDS Test Accreditation List held by this laboratory]	Next Generation Sequencing Analysis  Using: Germline Variant Calling and Annotation Pipeline (benchmarked against NHSE Pipeline Tool using Coriell Reference DNA)/ Congenica (for tertiary analysis)  SOPs: LP 363 059 NGS Exome LP 306 010 Targeted Cancer Panel LP 307 004 GitHub LP 307 005 Microsoft Azure LP 307 007 Germline Pipeline LP 307 008 Congenica Analysis	A
EDTA Blood	Loop-mediated Isothermal Amplification for the detection of: Haemochromatosis	Loop-mediated Isothermal Amplification (LAMP)  Using: LaCAR LAMP Human Haemochromatosis Kit/ Beckman Coulter Biomek 4000 Liquid Handler/ Roche LightCycler 480  SOPs: LP 363 074 HFE Genotyping LP 366 032 Biomek PD 305 302 LightCycler 480	A
Genomic DNA	Human Factor II (Prothrombin) G20210A mutation Human Factor V Leiden G1691A mutation	Cobas Mutational Analysis  Using Roche cobas Factor II and Factor V Test  SOP: LP 363 080 Cobas Factor II and Factor V	



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Genomic DNA	HAEMOPHILIA A - F8 Gene: Intron 1 Inversion	DNA Amplification by PCR & Detection by Gel Electrophoresis  Using: Horizontal Gel Electrophoresis Tank Electrophoresis Power Supply Thermocyclers [Definitive list in Current Dept Equipment List] Syngene G:BOX F3 Gel Documentation System MegaMix Gold Eurogentec Primers  SOP: LP 363 077 F8 Intron 1 Inversion	
Genomic DNA	HAEMOPHILIA A - F8 Gene: Intron 22 Inversion	DNA Amplification by PCR & Detection by Gel Electrophoresis  Using: QINSTRUMENTS BioShake iQ Heatblock Horizontal Gel Electrophoresis Tank Electrophoresis Power Supply Thermocyclers [Definitive list in Current Dept Equipment List] Syngene G:BOX F3 Gel Documentation System MegaMix Gold Promega Bcl1 restriction enzyme Promega T4 DNA Ligase Eurogentec Primers  SOP: LP 363 078 F8 Intron 22 Inversion	



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<b>HUMAN BODY FLUIDS / TISSUES</b>  Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	<b>Cytogenetics (Germline and Somatic)</b>  Diagnosis of aneuploidy in products of conception, intrauterine deaths, neonatal deaths, stillbirths and by rapid prenatal diagnosis	Quantitative Fluorescence-Polymerase Chain Reaction (QF-PCR)	A
		Products of Conception, Intrauterine Deaths, Neonatal Deaths and Stillbirths  Using: Elucigene QST*R-PL Kit/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Applied Biosystems 3500xL Genetic Analyzer/ SoftGenetics GeneMarker Software  SOPs: LP 352 001 QF-PCR Tissue Set Up LP 352 002 QF-PCR Tissue Analysis LP 366 038 ABI 3500xL	A
		Rapid Prenatal Diagnosis  Using: Elucigene QST*R Plus Kit/ Elucigene QST*R-13 Kit/ Elucigene QST*R-18 Kit/ Elucigene QST*R-21 Euplex Kit/ Elucigene QST*R-XY Kit/ Applied Biosystems 3500xL Genetic Analyzer/ SoftGenetics GeneMarker Software/ Applied Biosystems GeneMapper Software  SOPs: LP 352 003 QF-PCR AF Set Up LP 352 004 QF-PCR AF Analysis LP 366 038 ABI 3500xL	A



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Whole blood Amniotic fluid CSF Cord blood Bone marrow Tissues	Chromosome analysis for the purpose of clinical diagnosis  Detection of chromosomal rearrangements or aberrations arising from: Prenatally detected disorders Developmental disorders Reproductive medicine disorders Haematological/Oncology disorders	Documented in-house procedures using techniques and kits in combination with manufacturers' instructions:  Karyotyping and G-Banding  Cell Culturing and Harvesting  Using: Manual methods  SOPs: Prenatal LP 345 001 AF Culture LP 345 002 AF/Tissue LP 345 003 Solid Tissue Culture Developmental Delay LP 345 016 Blood Culture LP 345 034 Blood Harvest Haematological/Oncology LP 345 029 BM & Blood Culture LP 345 030 BM & Blood Harvest	A  A



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		<p>Chromosome Preparation and G-Banding</p> <p>Using: ADSBiotec Hanabi-PVI Metaphase Spreader/ Thermo Scientific ClearVue Coverslipper</p> <p>SOPs: LP 345 033 Slide Making QA LP 346 011 G-Banding LP 351 005 Hanabi</p>	A
		<p>Microscopic Analysis of G-Banded Chromosomes</p> <p>Using: Leica Microsystems DM5500B Microscope/ Leica Biosystems GSL-120 Slide Loader/ Leica Biosystems CytoVision Image Analysis Capture System</p> <p>SOPs: LI 348 003 GSL120 Cancer Analysis LI 348 004 GSL120 Constitutional Analysis LP 347 033 DM5500B Use LP 348 005 GSL120 Set Up LP 348 008 Cancer Chromosome Analysis LP 348 011 Cytovision Scanning Classifiers LP 348 034 Constitutional Chromosome Analysis</p>	A



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Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
Whole blood Amniotic fluid CVS CSF Cord blood Bone marrow Tissues Buccal cells Products of conception	Detection of chromosomal aberrations in the diagnosis of constitutional disorders and haematological malignancy using a range of probe types including: Break-apart probes Fusion probes Deletion probes Insertion probes Copy Number Amplification probes [Definitive list in RMDS FISH Probe List held by this laboratory]	Fluorescence In Situ Hybridisation (FISH)  Using: A range of probe suppliers including: Abbott Vysis/Cytocell/Empire Genomics/ [Definitive list in FISH Probe List] Stemcell EasySep Human CD138 Positive Selection Kit/ Stemcell EasySep Human CD19 Positive Selection Kit/ StemCell RoboSep-S Cell Separator/ Abbott StatSpin ThermoBrite S500/ Leica Microsystems DM5500B Microscope/ Leica Biosystems GSL-120 Slide Loader/ Leica Biosystems CytoVision Image Analysis Capture System/  SOPS: LI 347 007 ThermoBrite LI 347 034 Cytovision Spot Counting Analysis LP 345 037 CD138+ Selection LP 345 038 CD19+ Selection LP 347 001 Buccal Swabs Interphase FISH LP 347 002 FISH Set Up LP 347 003 Blood/BM FISH LP 347 005 Post Hybridisation FISH Washes LP 347 033 DM5500B Use LP 348 005 GSL120 Set Up LP 348 011 Cytovision Scanning Classifiers LP 348 038 FISH Analysis	A



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### Belfast Health and Social Care Trust

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Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Genomic analysis for detection of copy number changes or loss of heterozygosity arising from: Prenatally detected disorders Developmental disorders Reproductive medicine disorders	Single Nucleotide Polymorphism (SNP) Microarray  Using: Illumina Infinium CytoSNP-850K BeadChip Kit/ Illumina NextSeq 550 Instrument/ VWR Hybridisation Oven/ Tecan Te-Flow Unit/ Thermocyclers [Definitive list in Current Dept Equipment List]/ Illumina BlueFuse Multi Software  SOPs: LP 350 006 Microarray Set Up LP 350 012 Microarray Analysis PD 305 318 Recommendations for Microarray in Pregnancy	A



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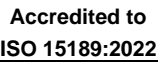
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HUMAN BODY FLUIDS / TISSUES	Solid Tumours (Somatic)	Documented in-house procedures using techniques and kits in combination with manufacturers instructions:	B
	Detection of specific gene alterations and antigen expression, in a range of cancer types to facilitate targeted treatment decisions		
	DNA extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long-term storage	Manual and Automated DNA Extraction and Quantification	B
Plasma	Extraction of cfDNA	Manual Extraction	
		Using: Roche Cobas cfDNA Sample Preparation Kit	
		SOPs: LP 390 033 Cobas cfDNA Extraction	



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Formalin Fixed Paraffin Embedded Tissue	Immunohistochemistry for the detection of: Breast cancer – PD-L1 Gastric cancer – HER2 Lung cancer - PD-L1	Immunohistochemistry (IHC)  Using: Roche VENTANA PD-L1 (SP142) Assay/ Roche VENTANA anti-HER2/neu (4B5) Rabbit Monoclonal Primary Antibody/  Roche VENTANA PD-L1 (SP263) Assay/  Roche Ventana Benchmark Ultra/ Sakura Tissue-Tek Prisma E2D Automated Slide Stainer and Coverslipper Film E2/ Olympus BX53 Light Microscope  SOPs: LP 390 237 IHC Set Up LP 390 236 IHC Analysis PMC-SOP-120 Ventana Benchmark Ultra PMC-SOP-42 Tissue-Tek PMC-SOP-55 Olympus BX53	B
Formalin Fixed Paraffin Embedded Tissue	Fluorescent Insitu Hybridisation for the detection of: Breast cancer – ERBB2	Fluorescent Insitu Hybridisation (FISH)  Using: Leica HER2 FISH system/ Leica BOND RX/ Zeiss AxioImager Z2  SOPs: LP 390 183 ERBB2 FISH Set Up LP 390 142 ERBB2 FISH Analysis LI 390 107 ERBB2 Tumour Annotation PMC-SOP-34 BOND RX PMC-SOP-52 AxioImager Z2	B



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Genomic DNA extracted in house from the sample types listed above and received as primary samples from external sources	Cobas Mutational Analysis for the detection of: Lung cancer – EGFR (DNA/cfDNA) Melanoma cancer – BRAF & NRAS	Cobas Mutational Analysis  Using: Cobas BRAF/NRAS Mutation Test/ Cobas EGFR Mutation Test/ Cobas z 480 Analyser  SOPs: LP 390 035 BRAF/NRAS Cobas LP 390 038 EGFR Cobas PMC-EXT-303 Cobas z 480 Analyser	B
Formalin Fixed Paraffin Embedded Tissue	Tumour Annotation	Tumour Annotation  Using: Olympus BX53 Light Microscope  SOPs: PMC-SOP-15 Tumour Content and Cellularity PMC-SOP-55 Olympus BX53	B
Genomic DNA	Pancancer Panel	Next Generation Sequencing  Using:	A



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		<p>Library Preparation</p> <p>Equipment: Invitrogen Qubit 4 Fluorometer Agilent 4200 TapeStation System Thermocyclers [Definitive list in iPassport Equipment Module]</p> <p>Reagents: Roche KAPA HyperPlus Kit Roche KAPA HyperCap Roche KAPA Hyperchoice Target Enrichment Probes</p> <p>SOP: PMC-SOP-74 Library Preparation</p> <p>Next Generation Sequencing Set Up</p> <p>Equipment: Illumina NovaSeq 6000</p> <p>SOP: PMC-SOP-24 NovaSeq-6000 PMC-SOP-56 Sequencing on the NovaSeq 6000</p> <p>Next Generation Sequencing Analysis</p> <p>Equipment: PanCancer Workflow Package for Analysis of Targeted Sequencing Data</p> <p>SOP: PMC-SOP-107 Analysis and Reporting PMC-SOP-79 Manual Analysis Using Integrative Genomics Viewer PMC-SOP-108 PanCancer Pipeline PMC-SOP-154 NGS Data Management</p>	
END			