


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <p>Accredited to ISO 15189:2022</p>	Buckinghamshire Healthcare NHS Trust	
	Issue No: 011 Issue date: 05 December 2025	
	Department of Cellular Pathology Wycombe Hospital Queen Alexandra Road High Wycombe HP11 2TT	Contact: Anna Clarke Tel: +44 (0)1494 426056 E-Mail: Anna.Clarke11@nhs.net Website: www.buckshealthcare.nhs.uk
Testing performed by the Organisation at the locations specified below		

Locations covered by the organisation and their relevant activities

Laboratory locations:

Location details	Activity	Location code
Address: Wycombe Hospital Queen Alexandra Road High Wycombe HP11 2TT Local contact Anna Clarke	Histopathology, Diagnostic Cytopathology Rapid frozen section for intra-operative diagnosis Andrology (fertility and PVSA) OSNA (One Step Nucleic Acid Amplification) for the detection of breast cancer metastasis.	A

Site activities performed away from the locations listed above:

Location details	Activity
Wycombe Hospital Mortuary Queen Alexandra Road High Wycombe HP11 2TT Anna Clarke	Receipt and storage of bodies
Stoke Mandeville Hospital Mortuary Mandeville Road Aylesbury HP21 8AL Anna Clarke	Receipt and storage of bodies Mortuary services



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<p>HUMAN TISSUES AND FLUIDS</p> <p>Body fluids: Fine needle aspirations and FNA material on slides Serous effusions Respiratory samples Bronchial washings Bronchial Brushings Transbronchial FNA) Bone aspirate Sputum Urine Gastrointestinal brushings Biliary brushing Synovial Fluid Cerebrospinal fluid Bile Fluid Cyst Fluids Hydrocele Fluid Nipple Discharge</p>	<p><u>Diagnostic Cytology</u></p> <p>Preparation and examination of cytological specimens to detect cytological abnormality to aid clinical diagnosis</p>	<p>Preparation/centrifugation In-house procedures: CPDC SOP 1 Diagnostic Cytology</p> <p>Shandon Cytospin 3 Centrifuge Thermo Scientific Sorvall ST 16</p>	A
<p>Slides prepared in-house from sample types listed above</p>	<p>Papanicolaou staining</p>	<p>Manual using in-house procedures: CPDC SOP 1 Diagnostic Cytology Automated using Sakura Prisma Plus using in-house procedures:</p>	A
<p>Slides prepared in-house from sample types listed above</p>	<p>Diff Quik Romanowski staining</p>	<p>Manual using in-house procedures: CPDC SOP 1 Diagnostic Cytology</p>	A
<p>Slides prepared in-house from sample types listed above</p>	<p>Morphological assessment and interpretation/diagnosis</p>	<p>Microscopy (qualitative analysis) In-house procedures: CPDC SOP 1 Diagnostic Cytology using microscopes as follows: Nikon Eclipse 80i Nikon Eclipse Ni-U Leica DMLB Olympus BX51 Olympus BX50 Nikon Eclipse 80i Leica DM4000 Nikon Eclipse E400</p>	A



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Diagnostic Cytology</u> (cont'd)		
Synovial Fluids	Detection and identification of Calcium pyrophosphate and monosodium urate crystals	Polarising light microscopy (qualitative analysis) using in-house procedures: CPDC SOP 1 Diagnostic Cytology Nikon Eclipse E400 microscope	A
HUMAN TISSUES AND FLUIDS	<u>Histopathology</u>	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	
Formalin fixed tissues	Macroanalysis and dissection	Manual method using CP HCC SOP 1 CP HCC SOP2 CP HCC SOP3 CP HCC SOP5 CP HCC SOP7 CP HCC SOP8 CP HCC SOP9 CP HCC SOP10 CP HCC SOP11 CP HCC SOP12 CP HCC SOP13 CP HCC SOP14 CP HCC SOP16 CP HCC SOP18 CP HCC SOP19 CP HCC SOP20 CP HCC SOP21 CP HCC SOP22 CP HCC SOP23 CP HCC SOP24 CP HCC SOP25 CP HCC SOP26 CP HCC SOP27 CP HCC SOP28 CP HCC SOP29 CP HCC SOP30 CP HCC SOP31 CP HCC SOP32 CP HCC SOP33 CP HCC SOP34	A



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Preparation and examination to identify or exclude morphological & cytological abnormalities</u>	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	
Formalin fixed tissues	Macroanalysis and dissection	Manual method using CP HCC SOP35 CP HCC SOP36	A
Formalin fixed tissues	Decalcification	Manual method using CP HCU SOP 6	A
Formalin fixed tissues	Tissue Processing	Automated method using Thermo Excelsior AS and ES tissue processors and Sakura VIP CP HCU SOP 14	A
Formalin fixed tissues	Embedding	Manual method using Leica EG1150 Leica EG1160 Thermo Histostar Leica Arcadia CP HML SOP 1	A
Formalin fixed tissues	Microtomy	Manual method using Leica RM2235 Microm HM325 Thermo Finesse Leica Histocore Biocut x 2. CP HML SOP 3 CP HML SOP 4 CP HML SOP 5 Leica RM2135	A
Fresh tissue Frozen tissue	Rapid frozen section Intra-operative diagnosis Basophilic and eosinophilic tissue structures	Cryotomy and manual and automated Haematoxylin & Eosin (H&E) staining method using Thermo Cryostat Cryotome HM 525 CP HSS SOP 1 Appx 11 CP CG SOP 7 CP HCU SOP 20	A



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology</u>	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	
FFPE tissue (cont'd)	Basophilic and eosinophilic tissue structures	<u>Routine Haematoxylin & Eosin (H&E) staining and coverslipping</u> Automated method using Gemini stainer AS CP HSS SOP 1 Appx 11	A
FFPE tissue (cont'd)	<u>Special stains</u> Detection of the following:	Automated Staining using Roche Ventana Benchmark BS CP HSS SOP 5 Manual staining CP HSS SOP 1 Appx 1-29	A
	Acid and sulphated mucins	Alcian Blue	
	Acidic and neutral mucins	Alcian Blue Periodic Acid-Schiff (AB PAS)	
	Elastic connective tissue fibres	Elastic van-Gieson (EVG) (Miller's)	
	Connective tissue and fibrin	Lendrum's Martius, Scarlet, Blue (MSB)	
	Campylobacter and Helicobacter sp.	Modified Giemsa stain	
	Organisms	Gram technique	
	Fungal hyphae and spores	Grocott Hexamine Silver	
	Pneumocystis sp.	Grocott	
	Chromatin	Haematoxylin and Eosin	
	Iron / haemosiderin	Perls Prussian Blue	
	Melanin, lipofuscin and melanosis	Mallory Bleach	
	Melanin and argentaffin granules	Masson Fontana silver	



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology</u> (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	A
FFPE tissue (cont'd)	Special stains Detection of the following: (cont'd)	Automated Staining using Roche Ventana Benchmark BS CP HSS SOP 5 Manual staining CP HSS SOP 1 Appx 1-29	
	Muscle fibrin and connective tissue	Masson Trichrome	
	Copper associated protein and Hepatitis B surface antigen	Orcein (Shikata)	
	Glycogen, mucopolysaccharide components, fungi and basement membranes	Periodic Acid-Schiff (PAS) technique	
	Glycogen	Diastase Periodic Acid-Schiff (D PAS) technique	
	Glycogen, acid and neutral mucins	Periodic Acid Schiff Diastase Alcian Blue (DABPAS)	
	Fibrin, muscle, and collagen	Picro Mallory	
	Reticulin fibres	Reticulin (Gomori)	
	Mast cells	Thionin	
	Collagen connective tissue fibres	Haematoxylin/Van-Gieson technique	
	Leprosy bacilli	Wade Fite technique	
	Acid and alcohol-fast bacilli	Ziehl-Neelsen (ZN) technique	



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<p>HUMAN TISSUES AND FLUIDS (cont'd)</p> <p>FFPE tissue (cont'd)</p>	<p><u>Histopathology</u> (cont'd)</p> <p>Preparation and examination to identify or exclude morphological & cytological abnormalities</p> <p><u>Immunohistochemistry</u></p> <p>Squamous epithelium. Stains all cell layers of epithelium. Prostate basal cells.</p> <p>Pan-keratin (Simple & complex epithelia)</p> <p>Germ Cell tumours e.g. embryonal Ca and yolk sac tumour. Hepatocellular Ca.</p> <p>Smooth Muscle cells and myoepithelial cells</p> <p>B-catenin protein, binds to cytoplasmic tail of E-cadherin</p> <p>For ID of follicular lymphoma</p> <p>Cell membrane glycoproteins expressed on healthy epithelia and in various carcinomas</p> <p>Apical brush border of epithelial cells of fallopian tube. ID of serous ovarian Ca and mesothelioma</p> <p>Calcium binding protein structurally related to S100 and inhibin</p> <p>Stains cells of epithelial origin.</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p>Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4</p> <p>34BE12</p> <p>AE1/AE3</p> <p>Alpha Fetoprotein</p> <p>Actin</p> <p>B-catenin</p> <p>BCL2</p> <p>BerEp4</p> <p>CA125</p> <p>Calretinin</p> <p>CAM 5.2</p>	<p>A</p>



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology</u> (cont'd) Preparation and examination to identify or exclude morphological & cytological abnormalities (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	A
FFPE tissue (cont'd)	<u>Immunohistochemistry</u> (cont'd)	Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4	
	T cells. Membrane and cytoplasmic. For ID of T cell neoplasms/lymphomas	CD3	
	B and T cell malignancies.	CD5	
	ID of ALL, Burkitts Lymphoma and Follicular Germinal Centre Lymphoma	CD10	
	Hodgkin cells and Reed-Sternberg cells	CD15	
	B cells. For ID of B cell lineage/lymphoma	CD20	
	For ID of Reed-Sternberg cells in Hodgkins lymphoma.	CD30	
	Endothelial cells in vascular and lymphatic spaces.	CD31	
	Endothelial cells in vascular and Lymphatic vessels	CD34	
	Lymphocytes, Histiocytes and macrophages	CD45	
	Peripheral nerves. ID of NK cells, NK-like T cells. Small cell lung Ca.	CD56	



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FFPE tissue (cont'd)	<u>Immunohistochemistry</u> (cont'd)	Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4	
	Islet cells of pancreas. Lymphocytes in tonsil. Squamous epithelial cells.	CD99	
	GIST tumours, mast cells and interstitial cells of Cajal in GI tract.	CD117	
	Enterocytes in the mucosa of appendix. Basal and luminal cells. For ID of GI adenocarcinomas and carcinoids	CDX2	
	Colonic epithelial cells. Expressed in colon Ca and lung adeno Ca.	CEA	
	Neuroendocrine cells in epithelial surface of colon. Nerve/ganglion cells.	CHROMO	
	Squamous epithelium in tonsil. Basal cells of the prostate.	CK5/6	
	Epithelial cells in pancreatic ducts and in breast epithelial cells. For ID of tumours of unknown origin	CK7	



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FFPE tissue (cont'd)	<u>Immunohistochemistry</u> (cont'd)	Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4	
	Stains placental villi, cytoplasmic staining	FACTOR 13a	
	Gross cystic disease fluid protein (Apocrine cells)	GCDFP	
	Glial fibrillary acidic protein (Neural cells)	GFAP	
	Stains trophoblastic elements in germ cell tumours.	HCG	
	Hepatocytes. For differential diagnosis of hepatocellular tumours, clear cell hepatocellular Ca from other clear cell malignancies.	HEPATOCYTE	
	Human melanoma black 45 antigen (Activated melanocytes; melanocytic tumours)	HMB45	
	Sertoli cells (diffuse and strong), granulosa cells, prostate, brain, adrenal	Inhibin	
	Plasma cells and related lymphoid cells containing Kappa light chains	Kappa light chains	
	For ID of proliferating cells in normal and neoplastic tissues.	KI67	



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FFPE tissue (cont'd)	<u>Immunohistochemistry</u> (cont'd)	Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4	
	Plasma cells and related lymphoid cells containing Lambda light chains	Lambda light chains	
	Differentiation antigen (Normal, activated & neoplastic melanocytes)	Melan A	
	ID of breast ductal Ca	MAMMAGLOBIN	
	ID of tumours of epithelial origin.	MNF 116	
	Reacts with an epithelial antigen. Stains a variety of lung tumours and other tumours.	MOC 31	
	Enolase gamma subunit (neuronal & neuroendocrine cells)	NSE	
	Pluripotent germ cell tumours	Oct3/4	
	Stains HPV associated squamous cell carcinoma in oropharyngeal specimens. Stains squamous intraepithelial lesions of the cervix.	P16	
	Carcinomas; many neoplasms and normal cells	p53 tumour suppressor protein	



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FFPE tissue (cont'd)	<u>Immunohistochemistry</u> (cont'd) Normal proliferating cells; poorly differentiated squamous cell carcinoma, basal cells; abluminal cells	Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4 p63	
	Prostate adeno Ca	P504S	
	ID of prostatic ductal epithelial cells	PAP	
	Placental Alkaline Phosphatase in germ cell tumours, particularly seminomas; also, breast, lung, ovarian carcinomas	PLAP	
	Cellular progesterone receptor expression	Progesterone receptor (PR)	
	Prostate specific antigen	PSA	
	ID of RCC vs non-renal cell tumours.	RCC	
	Normal and neoplastic melanocytes; macrophages; adipocytes; nerve fibres; many tumour types	S100 protein	
	Smooth muscle myosin	SMM	



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FFPE tissue (cont'd)	<u>Immunohistochemistry</u> (cont'd)	Automated methods using Roche Bench Ultra: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4	
	Synaptic vesicle protein (neurones; neuroendocrine cells)	Synaptophysin	
	Stains cells lining blood and lymphatic vessels and syncytiotrophoblasts of placenta.	THROMBOMODULIN	
	ID of tumours of thyroid origin.	THYROGLOBULIN	
	Thyroid transcription factor-1 (Thyroid follicular & C cells)	TTF-1	
	Vimentin intermediate filament	Vimentin	
	ID of malignancies involving differentiated plasma cells.	VS398c	
	WT1 expression	WT1	
	<u>Molecular</u>		
Fresh tissue: Sentinel lymph nodes (axillary)	Expression of Cytokeratin 19 (CK19) mRNA OSNA	Manufacturer's instructions for Sysmex RD-210 one step nucleic acid amplification analyser in conjunction with documented in-house methods: Detection and amplification of CK 19 mRNA CP HOS SOP 1	A



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HUMAN BODY FLUIDS	<u>Andrological examination activities for the purposes of clinical diagnosis</u>		
Semen	<u>Post vasectomy:</u> Detection of sperm	Manual method based on 2016 Laboratory guidelines for post vasectomy semen analysis: Association of Biomedical Andrologists, the British Andrology Society and the British Association of Urological Surgeons using large volume chambers/ phase microscopy and documented in-house procedure CPAND SOP 2	A
Semen	<u>Fertility testing:</u> Volume pH Sperm morphology excluding TZI scoring Sperm concentration Sperm motility	Based on WHO laboratory manual for the examination and processing of human semen (Sixth edition 2021) and manual method using documented in-house procedures CPAND SOP1 CPDC SOP 1 Volume/weight Colorimetric strips Stained slides (Papanicolau) microscopy (Bright field) Improved Neubauer chambers and microscopy (Phase contrast) Microscopy (Phase contrast)	A
END			