


# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <b>Accredited to ISO 15189:2022</b>	<b>Leeds Teaching Hospitals NHS Trust</b>	
	<b>Issue No: 016    Issue date: 10 June 2025</b>	
	<b>Leeds General Infirmary Old Medical School Great George Street LS1 3EX</b>	<b>Contact: Helen Pritchett Tel: +44 (0)113 39 25697 E-Mail: <a href="mailto:h.pritchett@nhs.net">h.pritchett@nhs.net</a> Website: <a href="https://www.leedsth.nhs.uk/a-z-of-services">https://www.leedsth.nhs.uk/a-z-of-services</a></b>
<b>Testing performed by the Organisation at the locations specified below</b>		

### Locations covered by the organisation and their relevant activities

#### Local Contact information:

Contact information		Department
Wendy Boote	E-Mail: <a href="mailto:wendy.boote@nhs.net">wendy.boote@nhs.net</a> Tel: +44 (0) 113 206 67609	Cellular Pathology
James Steer	E-Mail: <a href="mailto:leedsth-tr.dna@nhs.net">leedsth-tr.dna@nhs.net</a> Tel: +44 (0)1132 065205	Genetics



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**Laboratory locations:**

Location details	Department	Activity	Location code
Address  Chancellor Wing St James's University  Hospital Beckett Street LS9 7TF	Cellular Pathology	non-gynaecological cytology Reporting of the above Muscle / nerve histopathology, enzyme histochemistry	SJUH
Bexley Wing St James' University Hospital Beckett Street Leeds LS9 7TF	Cellular Pathology     Genetics	Routine diagnostic service for histopathology (including neuropathology and paediatric pathology) Reporting of the above and non gynae cytology Out of hours on call service for urgent frozen sections / diagnosis  FISH testing, DNA extraction and quantification, NGS library preparation	BEX
Seacroft Hospital York Road Leeds West Yorkshire LS14 6UH	Cellular Pathology	Andrology (post vasectomy and infertility investigation).	SEA
Yorkshire and North East Genomic Laboratory Hub Central Lab Ashley Wing St James's University Hospital Leeds LS9 7TF United Kingdom	Genetics	Molecular and Cytogenetics testing	ASH



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**Additional site activity performed away from the laboratory:**

Location details	Activities
On-site Mortuary and Body Store, St James' University Hospital  On-site Mortuary and Body Store, Leeds General Infirmary	Storage of bodies Viewing facilities Record keeping (body identification and audit trail)



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
HUMAN BODY TISSUE AND FLUIDS	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	
Body fluids / Fine needle aspirations, urines and bronchial/ sputum samples	<u>Diagnostic Non Gynae Cytopathology</u>	Sample preparation (Rotina 380 centrifuge, , Shandon cytospin 4):  CPATH-SOP-CYTO-2 Preparation of Non Gynae Samples  CPATH-SOP—HP-107 Preparation of Cell blocks	SJUH   SJUH
	Visualisation of nuclear detail in cells present	CPATH-SOP-CYTO-15 H&E (automated using Jung Autostainer)  CPATH-COSHH-10 Papanicolaou (automated using Jung Autostainer)  CPATH-COSHH-12 Rapid Romanowsky Stain	SJUH
Slides prepared in house as above	Morphological assessment and interpretation/diagnosis	CPATH-SOP-CYTO-8 Medical reporting of Non-Gynae cytology	BEX



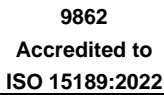
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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	BEX
Tissue / organs	<u>Histopathology</u>	Tissue dissection:  CPATH-SOP-HP-156 Cut Up of Specimens by Biomedical Scientists  HPSOP 0305 Cut Up of Specimens by Medical Staff CPATH-SOP-HP-125  CPATH-REC-527 Guidelines For Inflating A Lung  CPAYH-SOP-HP-65 Handling of CJD Specimens  CPATH-SOP-HP-125 Further Blocks  CPATH-REC-526 Guidelines for Opening a Fresh Uterus  CPATH-REC-527 Guidelines on Opening a Kidney  CPATH-SOP-HP-66 Renal Biopsy Procedure  CPATH-SOP-MED-1 Dissection of gall bladder resection specimens  CPATH-SOP-MED-2 Dissection of appendiceal resection specimens  CPATH-SOP-MED-30 Cut up of gynae specimens	



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Tissue / organs (cont'd)	<u>Histopathology (cont'd)</u>	Tissue processing:  CPATH-SOP-HP-160 Processing tissue using the Sakura X-Press (small urgent biopsies and neuropath specimens)  Embedding: CPATH-SOP-HP-109 Embedding paraffin wax impregnated tissues  Microtomy:  CPATH-SOP-HP-114 Sectioning using rotary microtome  CPATH-SOP-HP-28 Sectioning using automated microtome  CPATH-SOP-HP-28 Sectioning of Problem Blocks  CPATH-SOP-HP-27 Sections/levels required for particular tissue types  Frozen Sections:  CPATH-SOP-HP-22 Leica 1950 cryostat  CPATH-SOP-HP-23 Specimens Requiring Frozen Sections  CPATH-SOP-HP-18 Preparation of Split Skin Biopsies for Immunofluorescence	



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Tissue / organs (cont'd)	<u>Histopathology (cont'd)</u>	Frozen Sections:  CPATH-SOP-HP-23 Processing and Embedding of Alopecia Cases and other Paraffins  CPATH-SOP-MED-43 Protocol for Transplant Donor Lesion Biopsies – Frozen Sections	
Tissue	Identification of basophilic and eosinophilic structures	Routine Haematoxylin and Eosin Staining  Automated: CPATH-SOP-HP-63 Use and maintenance of Spectra Autostainer/coverslipper  Manual: CPATH-SOP-HP-4 Mayers haematoxylin and eosin for rapid method for frozen sections	
Tissue	<u>Histopathology - Identification of:</u>  Acid and neutral mucins  Diastase resistance acidic and neutral mucins  Mucins, glycogen  Diastase resistance mucin  Basement membranes, fungi	Automated: CPATH-SOP-HP-54HPSOP 657 Use of theAgilent Artisan Link Pro for Special stain techniques  ABPAS  ABPAS+D  PAS  DPAS  ExPAS	BEX



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Tissue (cont'd)	<u>Histopathology - Identification of: (cont'd)</u>	Automated: CPATH-SOP-HP-54 Use of the Artisan Link Pro for Special stain techniques	
	Basement membranes, fungi	ExDPAS	
	Fungi	Grocott	
	Iron (ferric)	Perls	
	Reticulin fibers	Retic	
	Spirochetes, H.Pylori	Warthin & Starry	
	Amyloid	Congo Red	
	Copper/Hepatitis B	Orcein	
	Sulphated acid mucins	CPATH-SOP-HP-29 Alcian blue pH1.0	
	Acid mucins	CPATH-SOP-HP-30 Alcian blue pH2.5	
	Acid mucins/connective tissue	CPATH-SOP-HP-31 Alcian blue van gieson	
	Acid mucins/connective tissue/elastic	CPATH-SOP-HP-53 Alcian blue elastic van gieson	
	AL amyloid	CPATH-SOP-HP-32 Congo red potassium permanganate	
	Nissl granules and neurones	CPATH-SOP-HP-33 Cresyl violet	
	Elastic/connective tissue	CPATH-SOP-HP-34 Elastic van gieson	



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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	BEX
Tissue (cont'd)	<u>Histopathology - Identification of (cont'd):</u>	Manual special staining:	
	H Pylori	CPATH-SOP-HP-35      Giemsa-modified	
	Bone marrow	CPATH-SOP-HP-37      Giemsa-overnight	
	Gram positive bacteria	CPATH-SOP-HP-37      Grams technique	
	Gram positive and negative bacteria	CPATH-SOP-HP-38      Gram twort	
	General tissue morphology	CPATH-SOP-HP-4      Mayers Haematoxylin and Eosin	
	Collagen/muscle	CPATH-SOP-HP-39      Haematoxylin van gieson	
	Acid mucin	CPATH-SOP-HP-52      Hales colloidal iron	
	Myelin	CPATH-SOP-HP-40      Luxol fast blue Haematoxylin and Eosin	
	Melanin	CPATH-SOP-HP-41      Masson Fontana	
	Muscle/collagen	CPATH-SOP-HP-42      Masson Trichrome	
	Paired sections identifies melanin pigment	CPATH-SOP-HP-43      Melanin bleach	



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Tissue (cont'd)	<u>Histopathology - Identification of (cont'd):</u>	Manual special staining:	
	Fibrin/connective tissue	CPATH-SOP-HP-44 MSB	
	Copper	CPATH-SOP-HP-45 Rhodanine technique	
	Fat	CPATH-SOP-HP-46 Sudan IV	
	Mast cells	CPATH-SOP-HP-48 Toluidine blue	
	Calcium	CPATH-SOP-HP-49 Von Kossa	
	<i>Mycobacterium leprae</i>	CPATH-SOP-HP-50 Wade fite	
	Tubercle bacilli	CPATH-SOP-HP-50 Ziehl Neelsen	
	Lipids	CPATH-SOP-HP-46 Sudan IV staining (frozen sections only)	



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Tissue and cell blocks	<u>Histopathology – identification of:</u>	Automated Immunohistochemical staining, PTLINK/Autostainer CPATH-SOP-HP-55 Preparation of Primary antibodies and associated reagents CPATH-SOP-HP-56 DAKO Autostainer Link 48 CPATH-SOP-HP-57 DAKO PTLINKs for antigen retrieval Using the following monoclonal, polyclonal and cocktail antibodies:	
	Amyloid deposits	AMYLOID-A	
	Lung cancer	PDL-1	
	Marker of normal throid/lung tumours	TTF-1	
		Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies: CPATH-SOP-HP-61 Use of Leica Bond III Immunostainer	
	A1-AT deiciency	A1-AT	
	Pituitary hormone	ACTH	
	Epithelial marker	AE1/3	
	Primary liver tumours/yolk sac tumours	AFP	
	Prostate cancer	AMACR	
	Prostate/breast marker	ANDROGEN R	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Lewy bodies /Parkinsons disease	A-SYN	
	Melanocytic lesions	BAP1	
	Neuronal damage	BAPP	
	Marker for apoptosis	BCL-2	
	Epithelial marker	BER-EP4	
	Senile plaques in Alzheimers disease	B-AMYLOID	
	Fibromatous lesions	B-CATENIN	
	Germ cell tumours	B-HCG	
	Marker of breast tumours	BRST-2 (GCDFP15)	
	Marker of ovarian tumours	CA125	
	Tumours of GI tract and pancreas	CA19.9	
	Marker of thyroid tumours	CALCITONIN	
	Marker of smooth muscle	CALDES	
	Marker for mesothelioma	CALRET	
	Marker of rejection in renal/liver transplants	C4D	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Langerhan cells	CD1A	
	T lymphocytes	CD3	
	T helper lymphocytes	CD4	
	T supressor lymphocytes	CD8	
	Lymphoid/endometrial malignancies	CD10	
	Hogkins lymphoma	CD15	
	B lymphocytes	CD20	
	Lymphoma marker	CD21	
	Hogkins lymphoma	CD30	
	Endothelial marker	CD31	
	Endothelial marker	CD34	
	Lymphocyte marker	CD45	
	Marker of nerve/neuroendocrine tumours	CD56	
	Histiocytes/macrophages	CD68	
	B lymphocytes	CD79a	
	Ewings sarcoma	CD99	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Mesothelioma marker	CD141	
	Histiocytes/macrophages	CD163	
	GI tract tumours	CDX2	
	Epithelial marker	CEA-M	
	Epithelial marker/liver tumour marker	CEA-P	
	Neuroendocrine cells	CHROMO	
	Epithelial marker	CK5	
	Epithelial marker	CK5/6	
	Epithelial marker	CK7	
	Epithelial marker/mallory bodies	CK8/18	
	Epithelial marker	CK14	
	Epithelial marker	CK19	
	Epithelial marker	CK20	
	Gastrointestinal stromal tumour marker	C-KIT (CD117)	
	Cytomegalovirus infection	CMV	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Marker of basement membrane	COLL IV	
	Marker of lymphatic channels	D2-40	
	Pan muscle marker	DES	
	Gastrointestinal stromal tumour marker	DOG-1	
	Cell adhesion molecule	E-CAD	
	Epidermal growth factor receptor	EGFR	
	Epithelial marker	EMA	
	Estrogen receptor/ prognostic breast marker	ER	
	Vascular tumours	ERG	
	Pituitary hormone	FSH	
	Marker of gastrin producing tumours	GASTRIN	
	Glial cells benign and malignant	GFAP	
	Pituitary hormone	GH	
	Pancreatic tumour typing	Glucagon	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Pancreatic tumour typing	Glucagon	
	Adenoma v HCC	Glutamine Synthetase	
	Liver adenoma/HCC	Glypican-3	
	Hepatitis B infected cells	HEP B c	
	Hepatitis B infected cells	HEP B s	
	Marker of normal/malignant hepatocytes	HEPAR 1	
	Marker of Her2 amplification in breast tumours.	HER 2	
	Melanoma marker	HMB45	
	Epithelial marker	HMC	
	Marker for H pylori	HP	
	Marker for teratoma	HPL	
	Herpes simplex 1 infection	HSV 1	
	Brain tumours	IDH1	
	Lymphoma markers	IgA IgG IgM	
	IgG4 disease marker	IgG4	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Granulosa cell tumours Mucinous epithelial ovarian tumours	Inhibin	
	Paediatric tumours	INI-1	
	Pancreatic tumours	Insulin	
	Lymphoma marker	KAPPA	
	Proliferation marker	KI67	
	Lymphoma marker	Lambda	
	Pituitary hormone	LH	
	Head and Neck tumour	Mammaglobin	
	Melanoma marker	MEL-A	
	Mismatch repair protein	MLH 1 MSH 2 MSH6	
	Epithelial marker	MNF	
	MUC4 in tumours	MUC4	
	Marker for rhabdomyosarcoma	MYOD 1	
	Marker for rhabdomyosarcoma	MYOGENIN	
	Neuronal marker	N52	



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Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Neuronal marker	NEU N	
	Neuronal marker	NFP	
	Neuroectodermal marker	NSE	
	Epithelial gynae tumours	PAX-8	
	Marker of CIN	P16	
	Marker of wild type P53 in malignancy	P53	
	Marker of molar pregnancy	p57	
	Marker in breast/prostate tumours	p63	
	Infection with parvo virus	PARVO	
	Marker of neuroblastoma	Phox 2B	
	Marker of germ cell tumours	PLAP	
	Mismatch repair protein	PMS2	
	Polyoma virus infection I renal transplant patients	POLYOMA SV40	
	Pituitary hormone	PROLACTIN	
	Progesterone receptor/prognostic breast marker	PR	



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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	BEX
Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):	
	Marker of normal/malignant prostate	PSA	
	Marker of normal/malignant prostate	PSAP	
	Marker of parathyroid tumours	PTH	
	Marker of ovarian malignancy	R1(inhibin)	
	Marker of primary renal carcinoma	RCC	
	Nerve sheath, neural marker	S100	
	Marker of serotonin producing tumours	SEROT	
	Pancreatic tumours	SOMAT	
	Smooth muscle marker	SMA	
	Smooth muscle marker	SMM	
	Marker of neural cells/neuroendocrine tumors	SYNAPT	
	Stat6 Expression	STAT6	
	Tangles in Alzheimers disease	TAU	
	Marker of normal/malignant thyroid	THYROG	



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**Leeds Teaching Hospitals NHS Trust**  
**Issue No: 016 Issue date: 10 June 2025**

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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	
Tissue and cell blocks (cont'd)	<u>Histopathology – identification of (cont'd):</u>  Pituitary hormone  Tangles, plaques and inclusions in dementia  Marker of Mallory bodies.  Mesenchymal tumours  Wilms tumour/mesothelioma  Desmoplastic cell tumour  <u>Histopathology</u>	Automated Immunohistochemical staining, Leica Bond III Immunostainer using the following monoclonal, polyclonal and cocktail antibodies (cont'd):  TSH  UBIQ brain  UBIQ liver  Vimentin  WT-1  WT-1 C19	BEX
Tissue (Breast primary)	HER2  <u>Histopathology – Enzyme histochemistry</u> <u>Immuno fluorescence (IF) for identification/demonstration of:</u>	Automated Immunohistochemical staining, CPATH-SOP-HP-58: Use of Ventana Benchmark Ultra for HER2 Antibody: anti-HER-2/neu(4B5)	BEX
Tissue (Rectal)	Presence/absence of ganglion cells and acetylcholinesterase when Hirschsprung's disease is suspected	CPATH-SOP-HP-19 Acetylcholinesterase	BEX



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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	
Tissue (Renal)	<u>Histopathology – Enzyme histochemistry</u> <u>Immuno fluorescence (IF) for identification/demonstration of:</u> (cont'd)		
	Immunoglobulins and complement	CPATH-SOP-HP-20 Immunofluorescence staining of renal biopsies Using antibodies IgA IgG IgM C3C C1q Kappa Lambda	BEX
Tissue (Skin)	Immunoglobulins and complement	CPATH-SOP-HP-21 Sectioning & Immunofluorescence Staining of Skin/ Oral/Neuro frozen sections Using antibodies in IgA IgG IgM C3C Fibrinogen	BEX
	<u>Histopathology –</u> Morphological assessment and interpretation/diagnosis:	Using light microscopy and SOPs:	BEX
Slides prepared in house as above (Breast)		CPATH-SOP-MED-5 Dissection and Reporting of Breast Specimens	
Slides prepared in house as above (Gastrointestinal)		CPATH-SOP-MED-6 Reporting of colonic resection specimens with vascular pathology  CPATH-SOP-MED-7 Reporting of ileo-colic resection specimens  CPATH-SOP-MED-8 Reporting of small intestinal resection specimens  CPATH-SOP-MED-9 Reporting of Colorectal Cancer Specimens	



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**Issue No: 016 Issue date: 10 June 2025**

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<p>HUMAN BODY TISSUE AND FLUIDS (cont'd)</p> <p>Slides prepared in house as above (Thoracic, skin)</p>	<p><u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u></p> <p><u>Histopathology –</u> Morphological assessment and interpretation/diagnosis: (cont'd)</p>	<p>Macroscopic and Microscopic examination:</p> <p>Documented in house methods incorporating manufacturers' instructions where relevant:</p> <p>Using light microscopy and SOPs:</p> <p>CPATH-SOP-MED-10 Reporting of resections for pancreatic, ampullary and bile duct carcinoma</p> <p>CPATH-SOP-MED-11 Reporting of stomach resection specimens</p> <p>CPATH-SOP-MED-12 Reporting of oesophageal resection specimens</p> <p>CPATH-SOP-MED-47 Reporting of endomucosal resection specimens (EMR)</p> <p>CPATH-SOP-MED-39 Reporting of liver resections not transplant</p> <p>CPATH-SOP-MED-13 Reporting of skin specimens, biopsies and resections</p> <p>CPATH-SOP-MED-14 Reporting of lung resection specimens</p> <p>CPATH-SOP-MED-15 Reporting of Thoracic lung biopsies and segmental excisions</p> <p>CPATH-SOP-MED-48 Reporting of Soft Tissue tumours specimens, core biopsies, excisions, amputations</p>	BEX



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**Issue No: 016 Issue date: 10 June 2025**

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<p>HUMAN BODY TISSUE AND FLUIDS (cont'd)</p> <p>Slides prepared in house as above (Head &amp; neck)</p> <p>Slides prepared in house as above (Neuropathology)</p>	<p><u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u></p> <p><u>Histopathology</u> – Morphological assessment and interpretation/diagnosis (cont'd):</p>	<p>Macroscopic and Microscopic examination:</p> <p>Documented in house methods incorporating manufacturers' instructions where relevant:</p> <p>Using light microscopy and SOPs:</p> <p>CPATH-SOP-MED-16 Reporting of oral mucosal specimens for tumours</p> <p>CPATH-SOP-MED-17 Reporting of thyroid specimens</p> <p>CPATH-SOP-MED-18 Reporting of small mucosal specimens</p> <p>CPATH-SOP-MED-19 Reporting of small bone specimens</p> <p>CPATH-SOP-MED-20 Reporting of salivary gland specimens for tumours</p> <p>CPATH-SOP-MED-21 Reporting of radical neck dissection specimens</p> <p>CPATH-SOP-MED-22 Reporting of maxillectomy specimens for tumours</p> <p>CPATH-SOP-MED-23 Reporting of laryngectomy specimens for tumours</p> <p>CPATH-SOP-MED-24 Reporting of Globes</p> <p>CPATH-SOP-MED-25 Reporting the examination of orbital exenterations</p>	BEX



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**Issue No: 016 Issue date: 10 June 2025**

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<p>HUMAN BODY TISSUE AND FLUIDS (cont'd)</p> <p>Slides prepared in house as above (Urology)</p> <p>Slides prepared in house as above (Urology)</p> <p>Slides prepared in house as above (Skin)</p>	<p><u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u></p> <p><u>Histopathology</u> – Morphological assessment and interpretation/diagnosis (cont'd):</p>	<p>Macroscopic and Microscopic examination:</p> <p>Documented in house methods incorporating manufacturers' instructions where relevant:</p> <p>Using light microscopy and SOPs:</p> <p>CPATH-SOP-MED-26 Reporting of wedge excisions - eyelid lesions</p> <p>CPATH-SOP-MED-27 Reporting of Temporal Artery Specimens</p> <p>CPATH-SOP-MED-28 Reporting of Trabeculectomy Specimens</p> <p>CPATH-SOP-MED-29 Reporting a penetrating keratoplasty (Cornea)</p> <p>CPATH-SOP-MED-34 Prostatic biopsies, transurethral or open prostatic resections and radical prostatectomies</p> <p>CPATH-SOP-MED-35 Adult renal parenchymal tumours</p> <p>CPATH-SOP-MED-36 Penile biopsies and resections</p> <p>CPATH-SOP-MED-37 Testicular biopsies and resections, retroperitoneal lymph node dissections</p> <p>CPATH-SOP-MED-38 Urinary bladder, ureteric, renal pelvic and urethral specimen</p> <p>CPATH-SOP-HP-21 Screening of skin biopsies by Immunofluorescence</p>	



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<p>HUMAN BODY TISSUE AND FLUIDS (cont'd)</p> <p>Digital images from slides prepared in house as above</p>	<p><u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u></p> <p><u>Histopathology</u> – Morphological assessment and interpretation/diagnosis (cont'd):</p>	<p>Macroscopic and Microscopic examination:</p> <p>Documented in house methods incorporating manufacturers' instructions where relevant:</p> <p>Using digital pathology (slide scanners and viewing software):</p> <p>Slide Scanning CPATH-SOP-HP-166 Scanning Digital Cases Using Leica GT450 Scanners</p> <p>Slide Reporting CPATH-SOP-HP-64 Reporting digital slides using Sectra IDs&amp;</p>	



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**Issue No: 016 Issue date: 10 June 2025**

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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	
Tissue	<u>Muscle / Nerve Histopathology</u>	Frozen section preparation (Microm Cryostat): CPATH-SOP-23 Cryostat Section Preparation for Histochemistry  Sample preparation techniques: CPATH-COSHH-32 Tissue Quenching for Histochemistry CPATH-SOP-SPH-12 CPATH-SOP-SPH-13 Specimen Handling for Muscle, Nerve, Rectal, Skin and Conjunctival Biopsies	SJUH
	Identification of basophilic and eosinophilic structures	CPATH-SOP-HP-4 Haematoxylin and Eosin Staining Method (cryostat sections)	SJUH
	<u>Muscle / Nerve Histopathology</u> - Identification of:	Manual special staining techniques:	SJUH
	Glycogen	CPATH-SOP-HP-8 Periodic Acid Schiff (PAS) and Diastase-PAS Staining Method for Cryostat Sections	
	Mitochondria and their distribution	CPATH-SOP-HP-3 Gomori Trichrome CPATH-SOP-HP-16 Succinate Dehydrogenase Demonstration	



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**Issue No: 016 Issue date: 10 June 2025**

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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	
Tissue (Muscle)	<u>Muscle / Nerve Histopathology</u> - Identification of normal /abnormal muscle fibre elements	CPATH-SOP-HP-15 Immunocytochemical Sarcolemmal Staining Method ( Real) Using the following monoclonal, polyclonal and antibodies:  Dystrophin 1 Dystrophin 2 Dystrophin 3 Alpha Sarcoglycan BetaSarcoglycan Gamma Sarcoglycan Delta Sarcoglycan Emerin Dysferlin Merosin Spectrin	SJUH
Tissue (Surgical and MH muscle)	Identification of nucleated cells  <u>Muscle / Nerve Histopathology - identification /demonstration of:</u>  Acid Phosphatase  Fibres with abnormal COX activity  Myoadenylate Deaminase  Adenosine triphosphatase	MHC-1 (HLA-ABC)  Enzyme histochemical staining CPATH-SOP-SPH-12 Specialist Histopathology Staining Panels  CPATH-SOP-HP-1 Acid Phosphatase – Naphthol AS-BI Phosphate  CPATH-SOP-HP-16 Combined Demonstration of Cytochrome Oxidase & Succinate Dehydrogenase CPATH-SOP-HP-2 Cytochrome Oxidase Demonstration  CPATH-SOP-HP-5 Myoadenylate Deaminase Demonstration  CPATH-SOP-HP-6 Myosin ATPase Staining Method	SJUH



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**Leeds Teaching Hospitals NHS Trust**  
**Issue No: 016 Issue date: 10 June 2025**

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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Examination activities in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis (cont'd)</u>	Macroscopic and Microscopic examination:  Documented in house methods incorporating manufacturers' instructions where relevant:	
Tissue (Muscle)	<u>Muscle / Nerve Histopathology - identification /demonstration of:</u>	Manual tinctorial Stain techniques for:	SJUH
Tissue (Surgical and MH muscle) (cont'd)	NADH	CPATH-SOP-HP-7 NADH - Tetrazolium Reductase Demonstration	
	PFK and Aldolase activity	CPATH-SOP-HP-9 Phosphofructokinase and Aldolase Demonstration	
	Myophosphorylase in muscle	CPATH-SOP-HP-13 Phosphorylase Demonstration	
	Presence and distribution of lipid in muscle	CPATH-SOP-HP-10 P-Phenylene diamine Staining Method  CPATH-SOP-HP-14 Sudan Black B Staining Method	
	Myelin in nerves	CPATH-SOP-SPH-12 Palade Fixation/Staining Method	
Slides prepared in house as above	<u>Muscle / Nerve Histopathology - Morphological assessment and interpretation/diagnosis</u>	CPATH-SOP-MED-3 Reporting of muscle biopsy specimen examination and handling	BEX
		CPATH-SOP-MED-4 Reporting of peripheral nerve biopsy specimen examination	BEX
FFPE sections prepared in house as above	<u>Molecular Pathology</u>  Detection of BRAF mutation at codon 600	RT-PCR CPATH-SOP-HP-60 Idylla™ System	BEX



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**Issue No: 016 Issue date: 10 June 2025**

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HUMAN BODY FLUIDS	<u>Andrological examination activities for the purposes of clinical diagnosis</u>		
Semen	<u>Post vasectomy:</u>  Detection of sperm	Based on 2016 Laboratory guidelines for post vasectomy semen analysis: Association of Biomedical Andrologists, the British Andrology Society and the British Association of Urological Surgeons) Manual method using large volume chambers/ manual microscopy OR wet prep and centrifugation and documented in-house procedure CPATH-SOP-AND-5 Post vasectomy testing	SEA
Semen	<u>Fertility testing:</u>     Volume  pH  Sperm morphology excluding TZI scoring  Sperm concentration   Sperm motility	Based on WHO laboratory manual for the examination and processing of human semen (Sixth edition 2021) and manual method / documented in-house procedures: CPATH-SOP-AND-1 Infertility testing CPATH-SOP-AND-2 Sperm density CPATH-SOP-AND-3 Motility testing CPATH-SOP-AND-4 Sperm morphology  Volume/weight  Colorimetric strips  Stained slides (H&E) microscopy (Bright field)  Improved Neubauer counting chambers and Microscopy (Phase contrast)  Microscopy (Phase contrast)	SEA



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**Leeds Teaching Hospitals NHS Trust**  
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<p><b>HUMAN TISSUE AND FLUIDS</b></p> <p>Human DNA / RNA Blood, saliva, tissue, paraffin embedded tissue amniotic fluid, chorionic villus sampling (CVS) Extracted DNA from external sources</p>	<p><b><u>Genetics</u></b></p> <p><u>Molecular genetic examinations for the purpose of clinical diagnosis (including predictive and prenatal) for a range of disorders:</u></p> <p><u>DNA/RNA Extraction, quantification and quality check for subsequent in-house analysis (see below), referral to specialist centres and long-term storage</u></p>	<p>In house documented methods incorporating manufacturers' instructions where relevant</p> <p>Using a combination of the methods listed below, supported by:</p> <ol style="list-style-type: none"> <li>1. DNA Extraction by: Automated EZ1 robot: (LG-SR-SOP-1)</li> <li>2. Automated Chemagic and Chemagic 360i: ((LG-SR-SOP-2))</li> <li>3. Manual extraction of cfDNA from plasma with Qiagen QIAamp kit: (LG-SR-SOP-3))</li> <li>4. Automated extraction using Maxwell RSC 48 extraction robot SOP LG-SR-SOP-4</li> <li>5. DNA Quantification by Nanodrop spectrophotometer and Tecan plate reader F200 (PicoGreen) or Qubit: LG-SR-SOP-5</li> <li>6. Amplification by Polymerase Chain Reaction/PCR (Thermocyclers: LG-PCR-SOP-12, Tecan Robot: LG-NGS-SOP-1)</li> <li>7. Sanger DNA sequencing (Applied Biosystems (AB) Analysers 3730, 3500XL: LG-PCR-SOP-1)</li> <li>8. Multiplex Ligation-dependent Probe Amplification, (AB, Thermocyclers: LG-PCR-SOP-12)</li> <li>9. Gel Electrophoresis (LG-PCR-SOP-3)</li> <li>10. Fragment analysis (AB) : LG-PCR-SOP-4) (Applied Biosystems (AB) Analysers 3730, 3500XL: LG-PCR-SOP-1)</li> </ol>	<p>BEX</p> <p>ASH CSB</p>



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**Leeds Teaching Hospitals NHS Trust**  
**Issue No: 016 Issue date: 10 June 2025**

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HUMAN TISSUE AND FLUIDS (cont'd)	<u>Molecular genetic examinations for the purpose of clinical diagnosis (including predictive and prenatal) for a range of disorders</u> (cont'd)	<p>In house documented methods incorporating manufacturers' instructions where relevant</p> <p>11.NGS Library preparation: (Sureselect – custom targeting and clinical exomeillumina , Sureselect Library Prep: LG-NGS-SOP-2</p> <p>Agilent Bravo: LG-NGS-SOP-2 Sciclone SureSelect Library Preparation: LG-NGS-SOP-11 Tecan Liquid Handling SOP: LG-NGS-SOP-1</p> <p>Agilent Bioanalyser: LG-NGS-SOP-14</p> <p>Covaris LE220: LG-NGS-SOP-12 Quantitative DNA analysis (PicoGreen): LG-NGS-SOP-20</p> <p>Next Generation Sequencing: sequencing and dosage analysis:</p> <p>12. NovaSeq 6000</p> <p>13. CFEU2 (Cystic Fibrosis Base - Yourgene) (Thermocyclers, AB : LG-PCR-SOP-5) 14. MFI-Y Plus (Male Factor Fertility Base- Yourgene) (Thermocyclers, AB: LG-PCR-SOP-6)</p>	<p>CSB BEX</p> <p>CSB</p> <p>ASH</p>



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<p>HUMAN TISSUE AND FLUIDS (cont'd)</p> <p>Human DNA obtained from:</p> <p style="margin-left: 20px;">A. blood, saliva, tissue, paraffin embedded tissue</p> <p style="margin-left: 20px;">B. amniotic fluid, chorionic villus sampling (CVS)</p>	<p><u>Molecular genetic examinations for the purpose of clinical diagnosis (including predictive and prenatal)</u> (cont'd)</p>	<p>In house documented methods incorporating manufacturers' instructions where relevant</p>	<p>ASH or ASH &amp; CSB, or ASH, CSB &amp; BEX (see techniques 1,2,4-8, 10, 11,12, above)</p>
<p>A, B</p>	<p>Li-Fraumeni syndrome</p>	<p>LG-IC-SOP-6</p>	
<p>A</p>	<p>Selected Gene Panel (Agilent Custom Design) including: Ataxia/Neuropathy panel BOB (Breast, ovarian, bowel) panel Cancer panel Connective tissue disorders panel Core (FH and CF) panel Dystonia/Paroxysmal (DYT) panel Gastrohepatology panel Haematology panel HSP/Degenerative disorders panel LGMD and MH panel Respiratory diseases panel Somatic BRCA panel YNExome panel (includes mitochondrial disorder targets)</p>	<p>LG-IC-SOP-4LG-NGS-SOP-6LG-NGS-SOP-6aLG-NGS-SOP-2</p>	<p>ASH or ASH &amp; CSB, or ASH, CSB &amp; BEX (see techniques 1.2.4-8,10, .11.12 above)</p>
<p>A, B</p>	<p>Variant Confirmation</p>	<p>LG-RD-SOP-33</p>	



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HUMAN TISSUE AND FLUIDS (cont'd)	<u>Molecular genetic examinations for the purpose of clinical diagnosis (including predicative and prenatal)</u> (cont'd)	In house documented methods incorporating manufacturers' instructions where relevant	
A,B	Cystic Fibrosis	LG-PCR-SOP-5	ASH or ASH & CSB, or ASH, CSB & BEX (see techniques 1,2,4-8, 10-13 above)
A (Blood only)	Y Chromosome Microdeletions	LG-PCR-SOP-6	ASH, CSB & BEX (See techniques 2, 5, 6, 10, 14 above)
A,B	DNA Profiling	LG-PCR-SOP-7	
Blood (EDTA anticoagulant), amniotic fluid, chorionic villus or solid tissue	Detection of aneuploidy of chromosomes 13, 18, 21	QF-PCR analysis involving DNA extraction followed by molecular analysis of DNA dosage for rapid aneuploidy using ABI 3500 or ABI 3730 gene scanner (LG-CF-SOP-24, LG-CF-SOP-30; LG-PP-SOP-1, LG-PP-SOP-15, LG-PP-SOP-17)	ASH & BEX (See techniques 1, 2, 4, 5, 6, 10)
Paraffin embedded Tissue	Human identity testing for detection of specimen mix up	Microsatellite PCR test to identify DNA variation at multiple genomic loci known to be unique to individuals using Thermal cycler, ABI 3500XL or ABI 3730 genetic analyser, LG-SO-SOP-3	ASH & BEX (See techniques 4, 5, 6, 10 above)



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HUMAN TISSUE AND FLUIDS (cont'd)	<u>Molecular genetic examinations for the purpose of clinical diagnosis (including predicative and prenatal)</u> (cont'd)	In house documented methods incorporating manufacturers' instructions where relevant	
Peripheral Blood	Detection of variants of DPYP gene indicating vulnerability to 5FU toxicity. Eleucie DPYD detection from the following alleles: c.1679T>G c.1905+1G>A c.2846A>T c.483+18G>A c.1129-5923C>G c.1236G>A.	Genotyping: Yourgene DPYD kit and analysis on using ABI 3500XL or ABI 3730 genetic analyser LG-IC-SOP-15	ASH & BEX (See techniques 4, 5, 6, 7, 9 above)
FFPE Tumour	Detection of methylation of the MLH1 gene (by DNA sequencing test)	Pyrosequencing involving DNA extraction followed by MLH1 methylation using Pyromark Q48 LG-PCR-SOP-11, LG-PCR-SOP-10, LG-NGS-SOP-5	ASH



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HUMAN TISSUE AND FLUIDS (cont'd)	<u>Molecular genetic examinations for the purpose of clinical diagnosis (including predicative and prenatal)</u> (cont'd)	In house documented methods incorporating manufacturers' instructions where relevant	
Blood plasma	Mutation testing of circulating tumour DNA - targeted mutation testing of the EGFR gene	Separation of blood plasma from whole blood, DNA extraction from blood plasma. Targeted EGFR mutation detection by real-time PCR using the Qiagen Rotagene Q and Therascreen Plasma RGQ PCR kit (LG-SO-SOP-24)	ASH & BEX (See techniques 3, 5 above)
Bone marrow, Blood	DNA extraction for the purposes of external Whole Genome Sequencing	Automated extraction using Chemagic and Chemagic 360i: (LG-SR-SOP-2)	BEX
HUMAN BODY FLUIDS/TISSUE	<u>Cytogenetics examination activities for the purpose of clinical diagnosis</u>	In house documented methods incorporating manufacturers' instructions where relevant	
Amniotic fluid, chorionic villus, solid tissue, blood (lithium heparin), bone marrow	Karyotype	Culture and karyotype analysis using G-banding LG-CF-SOP-3 and light microscope using SOPs:  LG-CF-SOP-4, LG-CF-SOP-6, LG-PP-SOP-11, LG-PPSOP-4 (amniotic fluid)  LG-CF-SOP-5, LG-CF-SOP-6, LG-PP-SOP-11,, LG-PPSOP-4 (chorionic villus)  Culture and karyotype analysis using G-banding LG-CF-SOP-3 and image analysis via automatic slide scanner /Cytovision using SOPs:  LG-CF-SOP-7, LG-CF-SOP-6, LG-PP-SOP-5, LG-PP-SOP-12, LG-PP-SOP-4 (solid tissue)	ASH



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2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

**Leeds Teaching Hospitals NHS Trust**  
**Issue No: 016 Issue date: 10 June 2025**

**Testing performed by the Organisation at the locations specified**

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<b>HUMAN BODY FLUIDS/TISSUE (cont'd)</b>	<u>Cytogenetics examination activities for the purpose of clinical diagnosis (cont'd)</u>	In house documented methods incorporating manufacturers' instructions where relevant	
Amniotic fluid, chorionic villus, solid tissue, blood (lithium heparin), bone marrow (cont'd)	Karyotype (cont'd)	LG-CF-SOP-1, LG-CF-SOP-14, LG-PP-SOP-9, LG-PP-SOP-3, LG-PP-SOP-5, LG-PP-SOP-12,, LG-PP-SOP-2 (blood) LG-CF-SOP-2, LG-PP-SOP-5, LG-PP-SOP-12,, LG-SO-SOP-6 (bone marrow)	ASH
Blood (lithium heparin anticoagulant)	Karyotype	Culture and karyotype analysis using G-banding and image analysis via automatic slide scanner /Cytovision LG-CF-SOP-1, LG-CF-SOP-3, LG-PP-SOP-3, LG-PP-SOP-5, LG-PP-SOP-12,, LG-PP-SOP-2	ASH
Bone marrow	Karyotype	Culture and karyotype analysis using G-banding and image analysis via automatic slide scanner /Cytovision LG-CF-SOP-2, LG-CF-SOP-3, LG-PP-SOP-5, LG-PP-SOP-12,, LG-SO-SOP-6	ASH
Illumina Bead Array GTC Files received from ISO 15189 accredited laboratory 9028	Interpretation of data to detect microscopic chromosomal imbalance (gains and losses) expressed as changes to copy number in: Postnatal disorders, prenatal diagnosis, neoplastic genetics	SNP array analysis using Illumina BlueFuse Multi software and SOP LG-RD-SOP-18	ASH
Fixed cells/Formalin Fixed Paraffin Embedded tissue (FFPE) from: Peripheral Blood Bone Marrow Amniotic fluid CVS- Chronic villus samples Solid tissues Solid tumours Buccal smears	Detection of chromosomal aberrations loci changes including confirmation of array CGH findings and rearrangements in the diagnosis of haematological malignancy, bone marrow failure syndromes, constitutional disorders, solid tumours	Pre analytical preparation of tissues for Fluorescent in situ hybridisation procedures (FISH) using Hybrite, Water bath, hot-plate and SOPs: LG-CF-SOP-9 (FFPE slides) (fixed suspension samples) followed by:	BEX



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<p>HUMAN BODY FLUIDS/TISSUE (cont'd)</p> <p>Fixed cells/Formalin Fixed Paraffin Embedded tissue (FFPE) from: Peripheral Blood Bone Marrow Amniotic fluid CVS- Chronic villus samples Solid tissues Solid tumours Buccal smears (cont'd)</p>	<p><u>Cytogenetics examination activities for the purpose of clinical diagnosis (cont'd)</u></p> <p>Break-apart probes Fusion product probes Single copy probes to detect gain or loss of a specific locus</p>	<p>In house documented methods incorporating manufacturers' instructions where relevant</p> <p>Fluorescent in situ hybridization (FISH) analysis of metaphase and interphase cells using commercial (and including probe validation/verification) using Cyto- vision analysis system and Fluorescent microscopy as defined in SOP LG-SO-MISC-4 v1.16 in conjunction with SOPs LG-CF-SOP-10, LG-CF-SOP-9, LG-SO-SOP-14, LG-CF-SOP-12, LG-CF-SOP-11</p>	BEX
END			