

# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <b>9997</b> Accredited to ISO 15189: 2022	<b>Advance Histopathology Laboratory Limited</b>	
	Issue No: 005    Issue date: 02 February 2026	
	<b>Laboratory</b> 47a Devonshire Street Lower Ground Floor London W1G 7AW	<b>Contact: Professor Gordon Stamp</b> Tel: +44 (0) 20 7636 9447 Fax: +44 (0) 20 7636 9929 E-Mail: g.stamp@ahlab.co.uk Website: www.ahlab.co.uk
Testing performed by the Organisation at the locations specified below		

### Locations covered by the organisation and their relevant activities

#### Laboratory locations:

Location details	Activity	Location code
<b>Address</b> Laboratory 47a Devonshire Street Lower Ground Floor London W1G 7AW	<b>Local contact</b> Professor Gordon Stamp	Histopathology  A

#### Site activities performed away from the locations listed above:

Location details	Activity	Location code
<b>Address</b> 41 Devonshire Street 2nd-floor London W16 7AJ	<b>Local contact</b> Professor Gordon Stamp	Reporting  B



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
HUMAN BODY TISSUE	<u>Histopathological examination activities for the purposes of clinical diagnosis</u>	Documented in house methods incorporating manufacturers' instructions where relevant:	
Formalin-fixed tissue samples	Examination of tissues in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis	Specimen Dissection Key SOPs: Specimen Reception AHL-Lab 62 Specimen Dissection AHL-Lab 30	A
		Automated Decalcification using the Sakura TDE 30 Decalcifier Key SOPs: AHL-Lab 243	A
		Manual Decalcification & Fixation using the Standard Analogue Shaker Key SOPs: AHL-Lab 243	A
		Tissue Processing using the Leica ASP300S tissue processor/ Sakura VIP6 AI/ Leica HistoCore Pegasus Plus tissue processor Key SOPs: AHL-Lab 67	A
		Embedding and Orientation using the Leica HistoCore Arcadia H embedder and Leica EG F Heated forceps Key SOPs: AHL-Lab 68	A



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Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
HUMAN BODY TISSUE (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	Documented in house methods incorporating manufacturers' instructions where relevant:	A
Fresh/frozen tissue	Examination of tissues in order to identify or exclude morphological and cytological abnormalities for the purpose of diagnosis	Frozen sections using the Leica CM1950 cryostat Key SOPs: AHL-Lab 73	A
Formalin Fixed paraffin wax blocks of processed tissue		Microtomy using the Histocore Biocut AS Microtomes. Slides printed using Primera slide printer and Eprexia Vega cassette printer Key SOPs: AHL-Lab 223, AHL-Lab 334	A
FFPE slides (Formalin fixed paraffin embedded) and frozen sections	Routine morphological staining for the detection of basophilic and eosinophilic structures	Manual and Automated Haematoxylin and Eosin (H&E) staining using the Leica ST5020 Multistainer, and coverslipping using the Leica CV5030 Fully Automated Glass Coverslipper Key SOPs: Manual AHL-Lab 249, Automated AHL-Lab 49	A
FFPE slides (Formalin fixed paraffin embedded)	Special stains for the detection of:	Documented in-house procedures for manual special stains using the following SOPs:	A
	Acid mucopolysaccharides Neutral mucopolysaccharides Mixed Mucins	Alcian Blue (AB) / Alcian Blue Periodic Acid Schiff (ABPAS) SOPs: AHL-Lab 33 / AHL-Lab 34	A
	Helicobacter pylori	Cresyl Fast Violet (CFV) AHL-Lab 242	A



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HUMAN BODY TISSUE (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	Documented in house methods incorporating manufacturers' instructions where relevant:	
FFPE slides (Formalin fixed paraffin embedded)	Special stains for the detection of:		
	Amyloid	Congo Red AHL-Lab 35	A
	Elastin Fibres Cell Granules Collagen	Millers Elastic Van Gieson (EVG) AHL-Lab 44	A
	Helicobacter Pylori Mast cells	Giemsa AHL-Lab 251	A
	Gram positive microorganisms Gram negative microorganisms	Gram AHL-Lab 36	A
	Fungal spores/hyphae/pneumocystis Collagen	Grocott Hexamine Silver AHL-Lab 37	A
	Melanin Argentaffin granules Chromaffin granules Nuclei	Masson Fontana AHL-Lab 41	A
	Melanin pigment	Melanin Bleach AHL-Lab 335	A
	Fibrin Muscle Collagen Erythrocytes Nuclei	Martius Scarlet Blue (MSB) AHL-Lab 40	A
Collagen Muscle fibres Erythrocytes	Masson Trichrome AHL-Lab 42	A	



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HUMAN BODY TISSUE (cont'd)	<u>Histopathological examination activities for the purposes of clinical diagnosis</u> (cont'd)	Documented in house methods incorporating manufacturers' instructions where relevant:	A
FFPE slides (Formalin fixed paraffin embedded) (cont'd)	Special stains for the detection of: (cont'd)	Documented in-house procedures for manual special stains using the following SOPs:	
	Hepatitis-B affected cells Copper-associated protein Elastin	Orcein AHL-Lab 336	A
	Haemosiderin Ferric Iron Nuclei	Perls' Prussian Blue AHL-Lab 31	A
	Neutral mucopolysaccharides Glycogen PAS positive substance Nuclei	Periodic Acid Schiff (PAS)/ Periodic Acid Schiff Diastase (dPAS) AHL-Lab 38	A
	Reticulin Fibres	Gordon and Sweet's Reticulin AHL-Lab 43	A
	Mast Cells	Toluidine Blue AHL-Lab 250	A
	Calcium Deposits	Von Kossa AHL-Lab 264	A
	Mycobacteria spp including TB	Ziehl Neelsen (ZN) AHL-Lab 39	A
Slides prepared as above	Morphological assessment and interpretation/diagnosis	Microscopy and reporting using Nikon and Olympus microscopes AHL-Lab 156-P	B
END			