


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <p>UKAS MEDICAL 8040</p> <p>Accredited to ISO 15189:2012</p>	<p>University College London Hospitals NHS Foundation Trust</p> <p>Issue No: 004 Issue date: 30 July 2021</p>	
	<p>Neurogenetics Unit Rare & Inherited Disease Laboratory NHS North Thames Genomic Laboratory Hub Level 4-6 Barclay House 37 Queen Square London WC1N 3BH</p>	<p>Contact: Vaneesha Gibbons Tel: 0207 829 8870 E-Mail: vaneeshagibbons@nhs.net Website: www.uclh.nhs.uk/neurogeneticslab</p>
<p>Testing performed at the above address only</p>		

DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of Sanger sequencing data</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype</p> <p>Variant detection in genes and variant confirmation and carrier/ predictive testing for the following neurological disorders:</p> <p>Andersen Tawil syndrome- KCNJ2</p> <p>Familial British Dementia- ITM2B</p> <p>Amyloidosis, Finnish Type (FAF)- GSN</p> <p>Charcot-Marie-Tooth Neuropathy, Type 1B and CMT2- MPZ</p> <p>CMT X-Linked - CX32/ GJB1</p> <p>Distal Hereditary Motor Neuropathy Type V; HMNV- BSCL2</p>	<p>Reporting of Sanger sequencing data using SeqScape software as described in NGENS ANA0001</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of Sanger sequencing data</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>Variant detection in genes and variant confirmation and carrier/ predictive testing for the following neurological disorders:</p> <p>Episodic ataxia: Type 1- KCNA1</p> <p>Familial Amyloid polyneuropathy; FAP- TTR</p> <p>Charcot-Marie-Tooth disease type 2K; CMT2K and Charcot-Marie-Tooth disease type 4A; CMT4A-GDAP1</p> <p>Charcot-Marie-Tooth disease type 2F; CMT2F and distal hereditary motor neuropathy; HMN2B- HSPB1</p> <p>Neuropathy, hereditary sensory and autonomic, type IA; HSAN1-SPTLC1</p> <p>Paramyotonia Congenita- SCN4A</p> <p>Hypokalemic periodic paralysis- CACNA1S & SCN4A</p> <p>Hyperkalemic periodic paralysis- SCN4A</p> <p>Leber Optic Atrophy; m.3460G>A; m.11778G>A; m.14484T>C</p> <p>Mitochondrial mutations in: MT-TK including m.8344A>G and MT-ATP6 including, m.8993T>G/C</p>	<p>Reporting of Sanger sequencing data using SeqScape software as described in NGENS ANA0001</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of Sanger sequencing data (including MLPA)</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>Sanger Sequencing and MLPA analysis for the following neurological disorders:</p> <p>SCN1A related epilepsy- SCN1A</p> <p>Charcot-Marie-Tooth Neuropathy, Type 2A2- MFN2</p> <p>Dopa Responsive Dystonia; DRD-GCH1</p> <p>Hereditary liability to pressure palsies- PMP22 del/PMP22 sequence mutations</p> <p>Charcot-Marie-Tooth disease demyelinating type 1A; CMT1A-PMP22 Dup/PMP22 sequence mutations</p> <p>Myotonia Congenita dominant and recessive forms- CLCN1</p> <p>Parkinson disease, juvenile, type 2 - PARK2</p>	<p>Reporting of Sanger sequencing data and MLPA data using SeqScape software as described in NGENS ANA0001 for Sanger sequencing and using Genemarker software as described in RGS ANA0205 and NGEN-LP-GeneMarkerMLPA for MLPA analysis</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of fragment analysis data (including MLPA)</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>DNA dosage analysis for:</p> <p>Autosomal dominant Parkinson's disease with a-synuclein rearrangements (PARK1/4)- SNCA</p> <p>Autosomal dominant Parkinson's disease-PARK8- LRRK2</p>	<p>Reporting of fragment analysis data (including MLPA) using Genemarker software as described in RGS ANA0205 and NGEN-LP-GeneMarkerMLPA</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of fragment analysis data (including MLPA)</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>Fragment analysis:</p> <p>Autosomal Dominant Spinocerebellar Ataxia :1,2,3,6,7, 12, 17- ATXN1; ATXN2; ATXN3; TBP, CACNA1A; ATXN7; PPP2R2B;</p> <p>Dentatorubropallidolysian Atrophy; DRPLA- ATN1</p> <p>Friedreich's Ataxia; FRDA- FXN</p> <p>Huntington Disease; HD- HTT</p> <p>Huntington Disease-Like 2; HDL2- JPH3</p> <p>Primary Torsion Dystonia; DYT1- TOR1A</p> <p>X- linked Bulbospinal Neuropathy- AR</p>	<p>Fragment sizing using GeneMapper software as described in SOPs NGENS ANA0004</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of fragment analysis data and blot image</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human mitochondrial genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>Detection of Chromosome-9 linked ALS/FTD- C9orf72</p>	<p>Reporting of fragment analysis data and Southern blot image using GeneMapper software as described in SOPs NGENS ANA0004 and NGENS DNG0001 for blot interpretation and reporting</p>
<p>Receipt of sequencing data and blot image</p> <p>Receipt of sequencing data</p>	<p>Large scale rearrangements of the mitochondrial genome.</p> <p>Detection of:</p> <p>Leber Optic Atrophy: m.3460G>A; m.11778G>A; m.14484T>C</p> <p>Mitochondrial common mutations: m.3243A>G, m.8344A>G, m.8993T>G/C</p>	<p>Reporting of fragment sizing data and blot image using NGENS DNG0018 and NGENS DNG0024</p> <p>Reporting of outcomes of targeted variant screening using SeqScape software as described in NGENS ANA0001 for Sanger sequencing and NGENS ANA0004 for GeneMapper</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of Sanger sequencing data and agarose gel eletrophoresis</p> <p>Receipt of NGS data</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human mitochondrial genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>DNA dosage analysis of mtDNA</p> <p>Mitochondrial whole genome sequencing</p>	<p>Reporting of Quantitative fluorescent PCR as described in NGENS DNG0019 and NGENS DNG0015</p> <p>SOP: Whole mtDNA NGS data analysis as described in NGENS DNG0028</p>



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<p>HUMAN BODY FLUIDS / TISSUES</p> <p>Receipt of Sanger data and MLPA data</p> <p>Data extracted following Next Generation Sequencing Analysis</p>	<p><u>Neurogenetics</u></p> <p>Analysis of the human genome (or part thereof) to detect pathogenic variation that results in a clinical phenotype (cont'd)</p> <p>Confirmation of pathogenic variation</p> <p>Variant interpretation</p>	<p>Using SeqScape software as described in NGENS ANA0001 for Sanger sequencing and using Genemarker software as described in RGS ANA0205 and NGEN-LP-GeneMarkerMLPA for MLPA</p> <p>Bioinformatic analysis with reference to: NGEN-LP-Alamut NGEN-LP-Custom NGS Panel Data Analysis for Pipeline NGEN-LP-NGS_Pipeline_Hands-on_Analysis</p>
END		