


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 UKAS MEDICAL 8639 Accredited to ISO 15189:2012	Oxford University Hospitals NHS Foundation Trust	
	Issue No: 008 Issue date: 26 March 2024	
	Department of Microbiology John Radcliffe Hospital Headley Way Oxford OX3 9DU	Contact: Lucinda Barrett Tel: +44 (0)1865-220881 Fax: +44 (0)1865-220890 E-Mail: Lucinda.Barrett@ouh.nhs.uk Website: www.ouh.nhs.uk/microbiology/
Testing performed at the above address only		

DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS Routine samples: Cellular and fluid material contained on swabs Genital samples Semen Cellular and fluid material contained on swabs Blood	<u>Microbiology</u> <u>Microbiological examination activities for the purpose of clinical diagnosis</u> Isolation of microorganisms of clinical significance Screening for Methicillin-Resistant <i>Staphylococcus aureus</i> (MRSA) Isolation of microorganisms of clinical significance	In-house documented procedures following current Public Health England (PHE) Standards for Microbiological Investigations (SMIs) Manual or automated inoculation, incubation and image capture of culture media using: M-SOP-75 M-SOP-76 M-SOP-212 Kiestra InoquIA Kiestra ReadA & ReadA browser Manual or automated inoculation, incubation and image capture of chromogenic culture media as above and using: M-SOP-44 Liquid media culture using: M-SOP-17 M-SOP-138 BD Bactec FX



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd)	In-house documented procedures following PHE SMIs
Sterile site samples: Tissue Swabs Fluid Bone Heart valve (homograft)	Isolation of microorganisms of clinical significance	Liquid media culture using: M-SOP-72 M-SOP-40 BD Bactec FX
Cerebrospinal fluid (CSF)	Detection, identification and quantification (inc. differential) of: White blood cells Red blood cells Bacterial cells	Microscopy & staining (Gram and differential) using: M-SOP-29
CSF	Isolation of microorganisms of clinical significance	Manual inoculation and media culture using: M-SOP-29
Bronchoalveolar lavage Bronchial aspirate, brushings & washings Cellular and fluid material contained on swabs Endotracheal tube specimens Nasopharyngeal aspirate Pleural fluid Sputum	Isolation of microorganisms of clinical significance	Manual inoculation and media culture using: M-SOP-137 M-SOP-139 M-SOP-138
Breast milk Corneal grafts Perfusion fluids Islet preparation Bone graft	Sterility check, isolation of microorganisms of clinical significance	Manual inoculation and media culture using: M-SOP-73 M-SOP-74
Aspirates Corneal scrapes Eye (specimens from) Fluids Tissues	Detection, identification and quantification of: White blood cells Red blood cells Bacterial cells Yeast cells and fungal hyphae	Staining (inc. Gram) and microscopic examination using: M-SOP-46 M-SOP-29 M-SOP-138



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd)	In-house documented procedures following PHE SMIs
Urine	Detection, identification and quantification of: White blood cells Red blood cells Bacterial cells Casts	Automated urinalysis, image capture and recognition using: M-SOP-209 A.Menarini Diagnostics SediMAX system M-SOP-86 M-SOP-212 Manual microscopic examination using: M-SOP-20985 M-SOP-86
	Legionella antigen	STANDARD F Legionella Ag FIA M-SOP-221 SD Biosensor STANDARD F 200
Faeces	Isolation, identification and antimicrobial susceptibility of bacterial pathogens: <i>Campylobacter</i> spp. <i>Clostridium difficile</i> <i>Escherichia coli</i> O157 <i>Salmonella</i> sp. <i>Shigella</i> spp. <i>Vibrio</i> spp. <i>Yersinia</i> spp. <i>Pleisiomonas</i> spp.	Manual inoculation and media culture using: M-SOP-111
Cultures isolated in-house from all samples listed above	Identification of microorganisms of clinical significance	Manual and/or automated biochemical methods using: M-SOP-204 BD Phoenix Matrix-assisted laser desorption/ionisation using: M-SOP-202 BD Bruker Maldi-Tof



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd)	In-house documented procedures following PHE SMIs
Cultures isolated in-house from all samples listed above	Antifungal susceptibility testing of clinically significant Fungi	Sensitivity testing using: AIM Nephelometer for sensititre plate MIC Manual Yeastone Sensitre M-SOP-88
Bacterial cultures isolated in-house from all samples listed above	Antimicrobial susceptibility testing of clinically significant bacteria	Manual and/or automated antimicrobial susceptibility testing using EUCAST methodology BD Phoenix using in-house procedure M-SOP-204 Disc diffusion & gradient minimum inhibitory concentration determination using in-house procedure M-SOP-88 NG-Test Carba 5 M-SOP-221
Hair Nail Skin	Isolation and identification of clinically significant dermatophyte fungi	Manual inoculation, media culture, and macroscopic & microscopic examination (stained & unstained) using: M-SOP-128 M-SOP-131 M-SOP-132
Faeces	Detection of ova, cysts and parasites (OCP)	Direct microscopic examination and/or Formol-ether faecal concentration and microscopic examination using: M-SOP-111
Faeces	Detection of <i>Cryptosporidium</i> (confirmation of EIA positives), <i>Isospora</i> & <i>Cyclospora</i> oocysts	Cold ZN incident-light microscopic examination using: M-SOP-111
Urine	Detection of <i>Schistosoma haematobium</i> ova	Microscopic examination using in-house procedure: M-SOP-209



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd)	In-house documented procedures following PHE SMIs
Sellotape slide Perianal swab	Detection of <i>Enterobius vermicularis</i> larvae and ova	Microscopic examination using: M-SOP-111
Bile Duodenal aspirates Jejunal aspirates	Detection of OCP including: <i>Strongyloides stercoralis</i> larvae, <i>Giardia lamblia</i> trophozoites <i>Cryptosporidium</i> oocysts <i>Cyclospora cayetanensis</i> oocysts <i>Cystoisospora belli</i> oocysts	Microscopic examination (stained & unstained) using: M-SOP-111
Aspirates Cellular and fluid material contained on swabs Fluids Sputum Tissue Urine	Isolation and presumptive identification of Acid-alcohol fast bacilli (inc. <i>Mycobacterium</i> spp.)	Liquid culture, auramine phenol staining and incident-light fluorescence microscopy using: M-SOP-139 TBc ID kit M-SOP-221 BD Bactec MGIT 960
Bronchoalveolar lavage	Detection Aspergillus antigen	OLM AspLFD M-SOP-221
Nasopharyngeal aspirate Nasopharyngeal swab	Detection or exclusion of: Influenza A Influenza B Respiratory syncytial virus Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2)	Molecular diagnosis using: M-SOP-184 Cephied GeneXpert
Nose / Throat swabs	Influenza A antigen Influenza B antigen	STANDARD F Influenza A/B FIA M-SOP-221 SD Biosensor STANDARD F 200
Sera Culture isolates BALs Nose & Throat swabs Urine Respiratory samples	Detection of: <i>Mycobacterium tuberculosis</i> Galactomannan Aspergillus Carbapenamase Legionella Influenza A&B antigens	lateral flow rapid chromatographic immunoassay M- SOP-221



8639

Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd) Plasma Serum	<u>Serology</u> <u>Serological examination activities for the purpose of clinical diagnosis</u> Microbial serology for: HIV Ab/Ag combo HBsAg HBsAg (confirmation) Anti-HBs HB Core IgM Anti-HBc HBeAg Anti-HBe HAV IgM HAV IgG HCV Ab HCV Ag Rubella IgM Toxoplasma IgM CMV IgG Syphilis total Ab EBV EBNA IgG EBV VCA IgG EBV VCA IgM HTLV I & II Ab CMV IgM Toxoplasma IgG Rubella IgG SARS-CoV-2 Anti-N IgG SARS-Cov_2 Anti-S IgG	Documented in-house methods to meet the requirements of the Infectious Diseases in Pregnancy Screening Programme (IDPS) screening programme as defined in the 'NHS Infectious Diseases in Pregnancy Screening Programme: laboratory quality assurance evidence requirements'. Chemiluminescence using: M-SOP-54 Abbott Architect i2000SR



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 **Issue date:** 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Serological examination activities for the purpose of clinical diagnosis</u> (cont'd)	
Plasma Serum	Microbial serology for: HSV 1&2 IgG VZV IgG Parvovirus IgM Parvovirus IgG Measles IgG Mumps IgG Borrelia IgG HIV Ab / Ag HCV Ab	Chemiluminescence using: M-SOP-51 DiaSorin Liaison XL
Plasma Serum	Detection of antibodies to HIV1 & HIV2	Bio-Rad Geenius Immuno chromatographic assay M-SOP -221
Plasma Serum	Rapid plasma regain (RPR)	Manual flocculation method using: M-SOP-159 ASI RPR Card Test for Syphilis
Plasma Serum	<u>Molecular diagnostics</u> <u>Molecular diagnostic examination activities for the purpose of clinical diagnosis</u> Molecular diagnostics for: Quantitative HIV Quantitative HCV Quantitative HBV	Extraction, nucleic acid amplification and detection using: Abbott Alinity m M-SOP-226



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Molecular diagnostic examination activities for the purpose of clinical diagnosis</u> (cont'd)	
Faeces	Detection of genomic DNA from bacteria causing infectious gastroenteritis: Salmonella spp., Shigella spp. Enteroinvasive E.coli (EIEC), <i>Campylobacter spp.</i> (jejuni and coli) Shiga toxin-producing organisms (STEC, Shigella dysenteriae)	Automated DNA extraction, amplification & detection on the BD MAX platform. M-SOP- 214 Extraction, amplification and detection using: M-SOP-189 M-SOP-190 and
Whole Blood Plasma CSF Eye swabs Vesicle swabs Vesicle fluid Buccal swabs Urine Amniotic fluid Saliva swab Bronchoalveolar lavage Sputum	Detection of: Quantitative BKV Quantitative CMV Quantitative Adenovirus Quantitative Pneumocysts jirovecii Quantitative Parvovirus B19 Quantitative HEV HSV1 & HSV2 VZVChlamydia trachomatis	ELITe ELITech InGenius, RT PCR, using ELITe MGB assays M-SOP-222
Nose/Throat swabs	Detection of Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2)	ThermoFisher MagMax MVP II kit running on the KingFisher Flex (extraction) QuantStudio 5 (Amplification & Detection) using ThermoFisher TaqPath COVID-19 assay



8639

Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN TISSUES AND FLUIDS (cont'd) CSF (Excludes samples from EVD/shunt/neurosurgical patients)	<u>Molecular diagnostic examination activities for the purpose of clinical diagnosis</u> (cont'd) Detection of the following microorganisms using the Meningitis/Encephalitis panel: Escherichia coli K1 Haemophilus influenzae Listeria monocytogenes Neisseria meningitidis Streptococcus agalactiae Streptococcus pneumoniae Cryptococcus gattii/neoformans Cytomegalovirus Herpes simplex virus 1 Herpes simplex virus 2 Human herpesvirus 6 Enterovirus Human parechovirus Varicella zoster virus	Biomerieux Biofire film array, multiplexed nucleic acid molecular detection using Procedure: M-SOP-220



8639
Accredited to
ISO 15189:2012

Schedule of Accreditation
issued by
United Kingdom Accreditation Service
2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

Oxford University Hospitals NHS Foundation Trust
Issue No: 008 Issue date: 26 March 2024

Testing performed at main address only

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
<p>HUMAN TISSUES AND FLUIDS (cont'd)</p> <p>NPA BAL Throat Swab (Excludes samples from immunocompromised patients)</p>	<p><u>Molecular diagnostic examination activities for the purpose of clinical diagnosis</u> (cont'd)</p> <p>Detection of the following Viruses / Bacteria using the Respiratory 2.1 plus panel</p> <p>Adenovirus Coronavirus 229E Coronavirus HKU1 Coronavirus NL63 Coronavirus OC43 Middle east Respiratory Syndrome Coronavirus (MERS-CoV) Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Enterovirus/Rhinovirus Human metapneumovirus Influenza A Influenza A subtype H1 Influenza A subtype H3 Influenza A subtype 2009 H1 Influenza B Parainfluenza virus 1 Parainfluenza virus 2 Parainfluenza virus 3 Parainfluenza virus 4 Respiratory syncytial virus Bordetella parapertussis Bordetella pertussis Chlamydomphila pneumoniae Mycoplasma pneumoniae</p>	<p>Biomerieux Biofire film array, multiplexed nucleic acid molecular detection using Procedure: M-SOP-220</p>
Nasopharyngeal swabs: COPAN UTM, Sigma Virocult, Remel M4R	SARS-CoV-2 RNA Orf genes	Biomerieux Biofire film array, multiplexed nucleic acid molecular detection using Procedure: M-SOP-220
Nose / Throat Swabs or RNA eluate	SARS-CoV-2 Sequencing	Nanopore GridION M-SOP-228
END		