


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <p>UKAS MEDICAL 8816</p> <p>Accredited to ISO 15189:2022</p>	<h3>NHS Fife</h3> <p>Issue No: 009 Issue date: 24 April 2025</p>	
	<p>Department of Cellular Pathology North Laboratories Hayfield Road Kirkcaldy Fife KY2 5AH</p>	<p>Contact: Derek Selbie Tel: +44 (0) 1592 643355 ext 28488 Fax: +44 (0) 1592 648170 E-Mail: Selbie Derek (NHS FIFE) (derek.selbie@nhs.scot)</p>
<p>Testing performed by the Organisation at the locations specified below</p>		

Locations covered by the organisation and their relevant activities

Laboratory locations:

Location details		Activity
<p>Address As Above</p>	<p>Local contact Derek Selbie</p>	<p>Routine Histology, Immunocytochemistry Diagnostic Cytology</p>

Location details		Activity
<p>Mortuary Victoria Hospital Hayfield Road Kirkcaldy Fife KY2 5AH</p>	<p>As above</p>	<p>Mortuary activities, Body Storage</p>
<p>Body Store Queen Margaret Hospital Dunfermline Fife KY12 0SU</p>	<p>As above</p>	<p>Body Storage</p>



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used
HUMAN BODY FLUIDS and TISSUES	<u>Diagnostic Cytology examinations to assist in clinical investigations</u>	In House documented procedure & manufacturer's instructions using series VLNL 015 – 021 and the following equipment:
Air Dried Preparations - Body fluids, CSFs	Processing and Staining of Diagnostic Specimens for detection of abnormal cells	Romanowsky Manual method using documented procedure VLNM 001
Wet Fixed Preparations – Body fluids, urine, CSFs, aspirates, EUSFNA	Processing and Staining of Diagnostic Specimens for detection of abnormal cells	Papanicolaou Automated method using Leica Autostainer and Leica CV5030 Coverslippers - ALAL 002 and ALAL 004.
FNA Wet Fixed Preparations Breast, Thyroid, Lymph nodes etc	Processing and Staining of Diagnostic Specimens for detection of abnormal cells	Leica CV5030 coverslipper – ALAL 002
FNA Air Dried Preparations - Breast, Thyroid, Lymph nodes etc	Processing and Staining of Diagnostic Specimens for detection of abnormal cells	Thermo IEC CL30 centrifuge – VLNL 001 Thermo Scientific Sorvall centrifuge-VLNL 001 Hologic Genesis Processor - VLNL 028 Thermo Cytospin 4 – VLNL 005
Endobronchial Ultrasound surgery samples (EBUS)	Preparation of tissue clots form EBUS samples	Receipt and preparation of EBUS specimens VLNL026 Thermo Scientific Sorvall centrifuge-VLNL 001



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<p>HUMAN BODY FLUIDS and TISSUES (cont'd)</p> <p>Frozen sections</p> <p>FFPE Tissue samples</p>	<p><u>Routine Histopathology examinations to assist in clinical investigations</u></p> <p><u>Frozen Section Histology Examinations & Touch Preparations to assist in clinical investigations</u></p> <p>Detection of abnormal cells</p> <p>Basophilic and eosinophilic cells</p> <p>Detection of abnormal cells</p>	<p>In House documented manual methods and manufacturer's instructions</p> <p>In House documented procedure & manufacturer's instructions using the following By microscopy manual methods using documentation VLHL 013 and VLHL 089</p> <p>Leica Cryostat – VLHL 007</p> <p>Manual method & documented procedure using VLHL 003 Routine staining (H&E)</p> <p>By microscopy supported by: Specimen Dissection Manual method using documented procedures in the range VLHL 023 - 97 MacroPath Digital Imager- VLHL 098</p> <p>Decalcification Manual method using VLHL083</p> <p>Tissue processing Documented procedure & manufacturer's instructions using Thermo Shandon Excelsior and Thermo Excelsior AS VLHL-020</p> <p>Tissue embedding Manual method, documented procedure & manufacturer's instructions using Leica EG1150 C & H Embedding centres - VLHL 002</p> <p>Tissue sectioning Manual method, documented procedure using Thermo Finesse, ME+ & Leica RM2255 Microtomes – VLHL 005</p>



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HUMAN BODY FLUIDS and TISSUES (cont'd)	<u>Routine Histopathology examinations to assist in clinical investigations</u> (cont'd)	In House documented manual methods and manufacturer's instructions
FFPE Tissue samples and Agar clots including EBUS	Basophilic and eosinophilic cells	Documented procedure & manufacturer's instructions using Leica Autostainer-ALAL004 and Leica CV5030 coverslipper Routine staining (H&E)
FFPE Tissue Samples	<u>Special staining of Histopathology examinations to assist in detection of clinical abnormalities</u>	In House documented procedure of manual methods using series VLHM 001 – 035 and Microscopy
	Acid Mucins	Alcian Blue pH 2.5
	Acid and Neutral Mucins	Alcian Blue PAS +/- Diastase
	Acid Fast Tubercle Bacilli	Ziehl Neelsen
	Amyloid	Congo Red (Highmans)
	Calcium	Von Kossa
	Connective tissue/Collagen/Muscle	Masson Trichrome Van Gieson
	Connective tissue/Collagen/Muscle/Fibrin	Martius Scarlet Blue (MSB)
	Elastic Fibres	Elastic Haematoxylin & Eosin Elastic Stain
	Fungi	Grocott Gomori Methenamine Silver
	Glycogen/Neutral Mucins/Fungi	Periodic Acid Schiffs (PAS) +/- Diastase
	Gram +ve & Gram -ve Organisms	Gram Stain
	Haemosiderin	Perls Prussian Blue
	Helicobacter/Micro-organisms	Modified Giemsa
	Hepatitis B affected Cells/Copper associated protein	Shikata's Orcein



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FFPE Tissue samples (cont'd)	Mast Cells	Toluidine Blue
	Melanin/Argentaffin/Chromaffin/some Lipofuscin	Masson Fontana
	Removal of Melanin	Melanin Bleach
	Mycobacterium Leprae	Wade-Fite
	Nuclei	Celestin Blue-Haematoxylin
	Reticulin Fibres	Gordon & Sweet's Reticulin
	Spirochetes	Warthin Starry
	<u>Immunocytochemistry of Histopathology examinations to assist in detection of clinical abnormalities</u>	In house documented procedures & manufacturer's instructions using Roche Ventana Benchmark Ultra system – SOP VLIL 003 & 005 and Leica CV5030 coverslipper Using antibodies to and microscopy;-
FFPE Tissue samples & Cell blocks/Agar clots including EBUS	Anaplastic Large Cell Lymphoma's (ALCL's)	ALK1
	Embryonic cells in normal tissues and germ tumours	AFP
	B cells, follicular lymphomas	Bcl-2
	B cells, lymphomas	Bcl-6 Protein
	B cells, plasma cells	Bob-1
	Ovarian epithelial malignancies, seminal vesical carcinoma, anaplastic lymphoma	CA125
	Parafollicular cells in thyroid and medullary thyroid carcinomas	Calcitonin
	Smooth muscle cells	Caldesmon
	Marker for mesotheliomas	Calretinin



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HUMAN BODY FLUIDS and TISSUES (cont'd)	<u>Immunocytochemistry of Histopathology examinations to assist in detection of clinical abnormalities (cont'd)</u>	In house documented procedures & manufacturer's instructions using Roche Ventana Benchmark Ultra system – SOP VLIL 003 & 005 and Leica CV5030 coverslipper Using antibodies to and microscopy;-
FFPE Tissue samples & Cell blocks/Agar clots including EBUS (cont'd)	<p>Mature B cells, follicular dendritic cells</p> <p>Various - mast cells, stem cells, Cajal cells, melanocytes, basal cells (skin) and mammary duct epithelium</p> <p>Plasma cells</p> <p>Granulocytes, Reed-Sternberg cells</p> <p>Langerhans cells</p> <p>T cells</p> <p>B cell</p> <p>Mature B cells, Follicular dendritic cell</p> <p>B cells, some T cell</p> <p>Reed-Sternberg cells, plasma cells</p> <p>Endothelial cells</p> <p>Myeloid cells</p> <p>Acute leukaemia, and differential diagnosis of poorly differentiated malignant tumours.</p> <p>T cells (helper & mature)</p> <p>T and B cell Lymphoma's</p> <p>Leucocytes</p> <p>T cells and the B1a subset of human B cells found in the follicular mantle zones</p>	<p>CD10</p> <p>CD117 (c-kit)</p> <p>CD138 (Syndecan-1)</p> <p>CD15 (Leu-M1)</p> <p>CD1a</p> <p>CD2, CD3</p> <p>CD20 (CD20cy)</p> <p>CD21</p> <p>CD23</p> <p>CD30 (Ki-1)</p> <p>CD31</p> <p>CD33</p> <p>CD34 (Qbend 10)</p> <p>CD4</p> <p>CD43</p> <p>CD45(LCA)</p> <p>CD5</p>



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<p>HUMAN BODY FLUIDS and TISSUES (cont'd)</p> <p>FFPE Tissue samples & Cell blocks/Agar clots including EBUS (cont'd)</p>	<p><u>Immunocytochemistry of Histopathology examinations to assist in detection of clinical abnormalities</u> (cont'd)</p> <p>Natural killer cells, neuroendocrine tissues</p> <p>Platelets, megakaryocytes, macrophages, endothelial cells</p> <p>T cells. Natural killer cells, thymocytes</p> <p>B cells</p> <p>T cells</p> <p>Wide range of tumours including Ewing's Sarcoma, & T cell lymphoma Intestinal epithelial cells, colorectal + other adenocarcinomas</p> <p>CEA glycoproteins</p> <p>Tissues of neuroendocrine origin</p> <p>Basal cells, filamentous keratins form glandular to stratified squamous, prostate adenocarcinoma</p> <p>Expressed in a range of tumours</p> <p>Pan cytokeratin (broad range)</p> <p>Epithelial cells</p> <p>Epithelial cells including ductal and glandular epithelia</p> <p>Adenocarcinomas, columnar epithelial cells</p> <p>Stratified squamous epithelium, basal cells, mesotheliomas</p>	<p>In house documented procedures & manufacturer's instructions using Roche Ventana Benchmark Ultra system – SOP VLIL 003 & 005 and Leica CV5030 coverslipper Using antibodies to and microscopy;-</p> <p>CD56(N-CAM) and</p> <p>CD57</p> <p>CD68 (PG M1 and KP1)</p> <p>CD7</p> <p>CD79a</p> <p>CD8 CD99 (MIC2) CDX2</p> <p>CEA</p> <p>Chromogranin A</p> <p>CK, HMW (34BE12)</p> <p>Cyclin D1</p> <p>Cytokeratin, MNF116</p> <p>Cytokeratin 14</p> <p>Cytokeratin 19</p> <p>Cytokeratin 20</p> <p>Cytokeratin 5/6</p>



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FFPE Tissue samples & Cell blocks/Agar clots including EBUS (cont'd)	Normal and neoplastic epithelia	Cytokeratin 7
	Pan cytokeratin (broad range)	Cytokeratin AE1/AE3
	Epithelial cells	Cytokeratin, Cam 5.2
	Cytomegalovirus	Cytomegalovirus
	Lymphatic Epithelium	D240
	Striated and smooth muscle cells	Desmin
	Gastro Intestinal Stromal tumours (GIST)	DOG-1
	Epstein-Barr Virus	EBV, LMP1
	Normal duct epithelial cells, ductal carcinoma	E-Cadherin
	Normal and neoplastic epithelium	EMA
	Epithelial cells, most adenocarcinomas	EP4
	Nuclei of cells containing a high level of oestrogen	Estrogen Receptor
	Luminal marker for breast epithelium	GATA 3
	Cytotoxic T Lymphocytes, Natural Killer cells	Granzyme B
	Anti-Glycophorin A for erythroid leukaemia.	Glycophorin A
	Placental trophoblasts	hCG
	Melanoma, melanocyte differentiation	HMB45
	Mitochondrial antigens of Hepatocytes	HSA



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FFPE Tissue samples & Cell blocks/Agar clots including EBUS (cont'd)	Sex chord stromal tumours	Inhibin
	Proliferating Cells	Ki-67
	Melanoma marker	Melan A
	Granulocytes and monocytes	Myeloperoxidase
	Lympho-haemopoietic tumours	MUM1
	Prostatic lesions	NKX3.1
	Peripheral nerves, neuroendocrine tumours	NSE
	B Cell lymphomas, germinal centre derived lymphomas	Oct 2
	Germ cell tumours, seminomas	OCT 3/4
	Various tumours	P16 Protein
	Squamous and Basal Cell Carcinoma's	P40
	Breast, colon, lung and urothelial carcinoma's	P53 (DO-7)
	Prostate basal cells, proliferative cells, glandular epithelium	P63
	B Cells	PAX-5
	Various Carcinomas	PAX-8
	Germ cell tumours, seminomas	Plac Alk Phos (PLAP)
	Nuclei of cells showing expression of Progesterone	Progesterone
	Metastatic prostatic tumours	Pros Acid Phos (PACP)



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FFPE Tissue samples & Cell blocks/Agar clots including EBUS (cont'd)	Prostate secretory and ductal epithelium Prostate adenocarcinomas Renal cell Carcinoma High Grade PIN Schwann cells, nerve processes, S100 +ve neoplasms Smooth muscle and myoepithelial tumors Melanocytic and schwannian tumors Neuroendocrine cells T & B Lymphocytes, Leukaemias, acute lymphocytic lymphomas Hairy Cell Leukaemia Thyroglobulin containing cells TTF-1 found in lung and thyroid (thyroid follicular cells) Cells of mesenchymal origin Epithelial cells and smooth muscle in fallopian tube. Wilms tumour Epstein-Barr Virus B cells and plasma cells B cell follicles	PSA Racemase (P504s) RCC S-100 SMA SOX10 Synaptophysin TdT TRAcP Thyroglobulin TTF-1 Vimentin Wilm's Tumour (WT1) EBER ISH Probe Kappa ISH Probe Lambda ISH Probe



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Slides prepared in-house as above	Morphological assessment and interpretation/diagnosis:	Using light microscopy and SOPs: SOP ALPM001
END		