


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <p>UKAS MEDICAL</p> <p>9315</p> <p>Accredited to ISO 15189:2012</p>	<h3>Hampshire Hospitals NHS Foundation Trust</h3>	
	<p>Issue No: 005</p>	<p>Issue date: 30 May 2019</p>
	<p>Cellular Pathology Department Romsey Road Winchester SO22 5DG</p>	<p>Contact: Jeff Morrison Tel: +44 (0) 1962 824 384 E-Mail: jeff.morrison@hhft.nhs.uk Website: www.hampshirehospitals.nhs.uk</p>
<p>Testing performed by the Organisation at the locations specified below</p>		

Locations covered by the organisation and their relevant activities

Laboratory locations:

Location details	Activity	Location code
<p>Address Cellular Pathology Department Royal Hampshire County Hospital Romsey Road Winchester SO22 5DG</p> <p>Local contact Jeff Morrison</p>	<p>Diagnostic cytology Routine Histology Immunocytochemistry Frozen sections OSNA Mortuary – body store, receipt and release Record keeping (body identification and audit trail)</p>	RHCH
<p>Cellular Pathology Department Basingstoke and North Hampshire Hospital Aldermaston Road Basingstoke RG24 9NA</p> <p>Local contact Jeff Morrison</p>	<p>Diagnostic cytology (receipt and reporting only) Pathologist and BMS cut up Frozen section OSNA Histopathology reporting Mortuary – body store, receipt and release Record keeping (body identification and audit trail)</p>	BNHH
<p>Microbiology Department Basingstoke and North Hampshire Hospital Aldermaston Road Basingstoke RG24 9NA</p> <p>Local contact Jeff Morrison</p>		BNHH (M)



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<p>HUMAN TISSUES AND FLUIDS</p> <p>Fixed and fresh cellular material and body fluid samples, prepared slides, fine needle aspirates and brushings:</p> <p>Unfixed Preparations: Body fluids, urine, CSFs, aspirates, washings, brushings, sputum Wet fixed preparations: brushings, EUSFNA Air Dried Preparations: Body fluids, CSFs FNA Wet Fixed Preparations: Breast, Thyroid, Lymph nodes, brushings, EUSFNA FNA Air Dried Preparations: Breast, Thyroid, Lymph nodes, EUSFNA</p>	<p><u>Diagnostic Cytopathology</u></p> <p>Examination of tissues to identify or exclude morphological and cytological abnormalities for the purposes of diagnosis</p>	<p><u>Diagnostic cytology</u> In-house documented procedures in conjunction with manufacturer's instructions using:</p>	RHCH
	<p>Visualisation of nuclear detail in cells present</p>	<p>Sample preparation (Mistral 3000E Centrifuge & Shandon Cytospin 4) LP-CYT-BCFPREP LP-CYT-URIPREP LP-CYT-CSFPREP LP-CYT-BRUPREP LP-CYT-ASPLAV LP-CYT-FNAPREP LP-CYT-SPUTUM</p> <p>LP-CYT-PAPSTAI Papanicolaou stain LP-CYT-H&ESTAIN H&E Stain LP-CYT-HEMAGUR Hema-Gurr Diff Quick stain LP-CYT-CENTRIF LP-CYT-CYTOCEN LP-CYT-SAFECAB LP-CYT-CVRSLPR LP-HIS-PRISMA</p>	RHCH
	<p>Morphological assessment and interpretation/diagnosis</p>	<p><u>Microscopy (qualitative analysis)</u> In-house procedures using light microscopy:</p> <p>LP-CYT-SCRNONG</p>	RHCH / BNHH



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<p>HUMAN BODY TISSUE AND FLUIDS</p> <p>Fixed, fresh and frozen tissue samples: excisional and incisional biopsies and surgical resections</p>	<p><u>Histopathological examination of tissues for the purpose of diagnosis</u></p> <p>Examination of tissues in order to identify or exclude morphological and cytological abnormalities for the purpose of clinical diagnoses.</p>	<p>Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):</p>	
		<p><u>Macroanalysis, Specimen dissection</u> using: LP-HIS-BMSCUTUP LP-HIS-CONCUT</p>	RHCH BNHH
		<p><u>Decalcification</u> using: LP-HIS-DECALC</p>	RHCH BNHH
		<p><u>Tissue processing</u> using: LP-HIS-VIP</p>	RHCH
		<p><u>Embedding</u> using: Sakura Tissue Embedding Centres LP-HIS-EMBEDHHFT</p>	RHCH
		<p><u>Frozen sections</u> using: Cryostat Thermo Shandon Cryostat LP-HIS-FROZENHHFT WI-HIS-FROZEN</p>	RHCH
		<p><u>Frozen sections</u> using: Cryostar NX50 (Thermo Fisher) LP-HIS-FROZENHHFT WI-HIS-NX50</p>	BNHH
<p><u>Microtomy and slide production</u> using: Leica rotary microtomes LP-HIS-MICROTOMY WI-HIS-MICROTOMY</p>	RHCH		



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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Histopathological examination of tissues for the purpose of diagnosis</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	
FFPE tissue samples and cell blocks	All tissue components: Nuclei Cytoplasm Muscle fibres Red blood cells Fibrin	<u>Automated Routine Haematoxylin & Eosin staining</u> using: Sakura H&E staining machine Sakura Prisma and Coverfilm LP-HIS-PRISMA	RHCH
FFPE tissue samples and cell blocks and frozen sections	All tissue components: Nuclei Cytoplasm Muscle fibres Red blood cells Fibrin	<u>Manual H&E staining</u> using: LP-HIS-PRISMA	RHCH
Frozen section	All tissue components: Nuclei Cytoplasm Muscle fibres Red blood cells Fibrin	<u>Manual H&E staining</u> using: LP-HIS-FROZ	BNHH
FFPE tissue samples and cell blocks	Special staining for identification of: Acid mucins and neutral mucins	<u>Automated Special staining</u> using: Sakura Prisma AB/PASD PAS/D LP-HIS-PRISMA	RHCH
FFPE tissue samples and cell blocks	Special staining for identification of: Acid mucins Acid mucins and neutral mucins Amyloid	<u>Special staining using manual techniques</u> Alcian blue WI-HIS-AB Alcian Blue Periodic Acid Schiff (AB PAS) +/- Diastase WI-HIS-ABPAS Congo Red WI-HIS-ALKCR	RHCH



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HUMAN BODY TISSUE AND FLUIDS (cont'd)	<u>Histopathological examination of tissues for the purpose of diagnosis</u> (cont'd)	Macroscopic and Microscopic examination using in house procedures in conjunction with manufacturer's instructions for the following methods (where relevant):	RHCH
FFPE tissue samples and cell blocks (cont'd)	Special staining for identification of: (cont'd)	<u>Special staining using manual techniques</u>	
	Elastin and connective tissue	EVG - Miller's Elastic Van Gieson WI-HIS-EVG	
	Helicobacter Pylori	Giemsa WI-HIS-GIEMSA	
	Differentiate between gram positive and negative bacteria	Gram WI-HIS-GRAM	
	Fungi	Grocott's Hexamine Silver WI-HIS-GROCOTT	
	Fibrin	Martius Scarlet Blue (MSB) WI-HIS-MSB	
	Connective tissue	Masson Trichrome WI-HIS-MASSTRI	
	Melanin	Masson – Fontana WI-HIS-MFONT	
	Tissue carbohydrates e.g. glycogen, mucins and fungal cell walls.	PAS +/- diastase WI-HIS-PAS	
	Demonstration of iron	Perls WI-HIS-PERLS	
	Reticulin	Reticulin - silver impregnation technique WI-HIS-RETIC	
	Copper associated protein / HepBsAg	Shikata's Orcein WI-HIS-SHITAKA	
	Mast cells	Toluidine Blue WI-HIS-TOLBLUE	
	Differentiation of collagen from other connective tissues	Van Gieson Stain WI-HIS-hvg	



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Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
FFPE tissue samples and cell blocks	<u>Immunohistochemistry (using antibodies) to detect the following for the purposes of clinical diagnosis</u>	<u>Automated</u> Immunohistochemical staining - Documented in house methods	RHCH
	Atypical mycobacteria	Wade Fite WI-HIS-WFITE	
	Acid fast Bacilli	Ziehl Neelsen (ZN) WI-HIS-ZN	
	High Molecular Weight Cytokeratin (Cytokeratins 1,5, 10 and 14	34BE12	
	Tumours of epithelial origin	AE1/AE3	
	Anaplastic lymphoma Kinase	ALK-1	
	B cells in mantle zone and interfollicular T cells.	BCL-2	
	Epithelial marker	BerEP4	
	Epithelioid malignancies of the ovary, cervical papillary serous carcinoma, adenocarcinoma of the endometrium, clear cell bladder adenocarcinoma, epithelioid mesothelioma	CA125	
	Mesothelial cell lining	Calretinin	
	Low molecular weight cytokeratin 7 and 8	CAM5.2	
	T-cells and T-cell neoplasms	CD3	
	B & T-cell malignancies	CD5	
T-cell lymphomas	CD8		
Early lymphoid cells and various non-lymphoid cells. Aids identification of Burkitt lymphoma and renal cell carcinomas	CD10		



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FFPE tissue samples and cell blocks (cont'd)	<u>Immunohistochemistry (using antibodies) to detect the following for the purposes of clinical diagnosis (cont'd)</u>	<u>Automated</u> Immunohistochemical staining - Documented in house methods	RHCH
	Reed- Sternberg cells – Hodgkin's lymphoma	CD15	
	B-cells (precursors and mature)	CD20	
	B-cells and monocytes	CD23	
	Reed Sternberg cells, activated B&T-lymphocytes	CD30	
	Endothelial cells	CD31	
	Endothelial cells, vascular and lymphocytic tumours	CD34	
	Leucocytes, lymphoid neoplasms	CD45 (LCA)	
	Lymphomas, small cell lung cancer	CD56	
	Megakaryocytic differentiation	CD61	
	Monocytes, macrophages	CD68	
	B-cell neoplasms	CD79A	
	GISTs, acute myeloid leukemia	CD117	
	Intestinal epithelial cells	CDX2	
	GI tract adenocarcinomas	CEA	
Cells of neuroendocrine origin	CHROMOGRANIN A		
Simple and non-keratinizing epithelia, tumours of epithelial origin	CK19		



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FFPE tissue samples and cell blocks (cont'd)	<p><u>Immunohistochemistry (using antibodies) to detect the following for the purposes of clinical diagnosis (cont'd)</u></p> <p>Normal and neoplastic epithelial cells – adenocarcinomas of colon, stomach, pancreas etc.</p> <p>Epithelial basal cells, mesothelioma</p> <p>Normal and neoplastic epithelial cells – adenocarcinomas of breast, lung, endometrium, thyroid</p> <p>Cell cycle regulation. Identification of mantle cell lymphomas</p> <p>Smooth and striated muscle cells Identification and classification of GIST</p> <p>Epithelial cells. Identification and classification of ductal breast carcinomas</p> <p>Secretory epithelial cells, Hodgkin's lymphoma, anaplastic large cell lymphoma</p> <p>Von Willebrand Factor – endothelial cells and megakaryocytes</p> <p>Platelets, Megakaryocytes, histiocytic cells, dermal dendritic cells. Identification dermatofibroma and some vascular neoplasms</p>	<p><u>Automated</u> Immunohistochemical staining - Documented in house methods</p> <p>CK20</p> <p>CK5/6</p> <p>CK7</p> <p>Cyclin D1</p> <p>Desmin</p> <p>DOG-1</p> <p>E-CADHERIN</p> <p>EMA</p> <p>Factor VIII</p> <p>Factor XIIIa</p>	RHCH



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FFPE tissue samples and cell blocks (cont'd)	<p><u>Immunohistochemistry (using antibodies) to detect the following for the purposes of clinical diagnosis (cont'd)</u></p> <p>Normal human hepatocytes, hepatocellular carcinomas</p> <p>Assessment of HER2 receptor status in invasive breast carcinomas</p> <p>Melanosomes. Identification and classification of malignant melanocytic lesions.</p> <p>IgG4-positive plasma cells. Diagnosis of IgG4 related sclerosing diseases</p> <p>Sex-cord stromal tumours</p> <p>Cells in active phases of the cell cycle. Classification of a variety of tumours</p> <p>Melanocytes. Identification and classification of melanomas</p> <p>Epithelial cells – broad reactivity (simple squamous to stratified squamous). Identification of epithelial cell neoplasms</p> <p>Normal epithelial cells. Identification and classification of neoplasms. Lung adenocarcinoma vs mesothelioma</p> <p>Cells of neuronal and neuroendocrine origin.</p>	<p><u>Automated</u> Immunohistochemical staining - Documented in house methods</p> <p>Hepatocyte Specific Antigen</p> <p>Her2 (4B5)</p> <p>HMB45</p> <p>IgG4</p> <p>Inhibin</p> <p>KI-67</p> <p>MELAN A</p> <p>MNF116</p> <p>MOC-31</p> <p>NSE</p>	RHCH



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FFPE tissue samples and cell blocks (cont'd)	<u>Immunohistochemistry (using antibodies) to detect the following for the purposes of clinical diagnosis (cont'd)</u>	<u>Automated</u> Immunohistochemical staining - Documented in house methods	RHCH
	Assessment of receptor expression invasive breast carcinomas	Oestrogen Receptor (ER)	
	Diagnosis of high grade cervical intraepithelial neoplasia, oropharyngeal squamous cell carcinomas	P16	
	Epithelial basal cells. Used for investigation of prostate, breast and lung neoplasms	P63	
	Some epithelial cells of kidney, thyroid and Müllerian organs. Identification and classification of ovarian serous carcinoma, clear cell renal carcinoma and papillary thyroid carcinoma	PAX-8	
	Germ cell neoplasms	PLAP	
	Assessment of receptor expression in invasive breast carcinomas	PR	
	Prostate secretory and ductal epithelial cells. Neoplasms of prostatic origin	PSA	
	Classification of tumours in central and peripheral nervous system, malignant melanocytic tumours, malignant melanoma metastasis	S100	
Glandular epithelium of prostate. Neoplasms of prostatic origin.	PSAP		



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FFPE tissue samples and cell blocks (cont'd)	<p><u>Immunohistochemistry (using antibodies) to detect the following for the purposes of clinical diagnosis (cont'd)</u></p> <p>Myocardial, skeletal and smooth muscle cells and myoepithelial cells</p> <p>Smooth muscle cells and myoepithelial cells. Classification of breast tumours.</p> <p>Neuroendocrine cells. Classification of neuroendocrine neoplasm</p> <p>Lung and thyroid tissue. Aids in classification of lung and thyroid tumours</p> <p>Thymocytes, resting T-cells, mature activated T-cells. Classification of T-cell neoplasms</p> <p>Cells of mesenchymal origin and tumours of mesenchymal origin</p> <p>Identification of Wilms' tumours, malignant mesothelioma</p> <p>Hepatocellular carcinomas, yolk sac tumours, mixed germ cell tumours</p> <p>Identification of human DNA mismatch repair proteins MLH-1, MSH-2, MSH-6 and PMS-2. Classification of colorectal tumours..</p>	<p><u>Automated</u> Immunohistochemical staining - Documented in house methods</p> <p>Smooth Muscle Actin</p> <p>Smooth Muscle Myosin</p> <p>Synaptophysin</p> <p>TTF-1</p> <p>UCHL-1 (CD45R0)</p> <p>Vimentin</p> <p>WT-1</p> <p>Alpha fetoprotein</p> <p>MLH-1 MSH2 MSH-6 PMS-2</p>	RHCH



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Slides prepared in house from samples listed above	<u>Morphological assessment and interpretation/diagnosis</u>	<u>Microscopy (qualitative analysis)</u> using: Leica/Nikon microscopes Consultant reporting SOP	BNHH RHCH
Lymph node tissue	Detect the expression level of Cytokeratin 19 mRNA normally not present in lymph node tissue	Manufacturer's instructions and documented in-house procedures with reference to: Homogeniser, vortex, centrifuge, Sysmex RD100i LP-HIS-OSNA Intraoperative analysis of sentinel lymph nodes using OSNA	BNHH RHCH
END			