


# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 Accredited to ISO 15189:2012	<b>Buckinghamshire Healthcare NHS Trust</b>	
	Issue No: 003    Issue date: 30 March 2022	
	Department of Cellular Pathology Wycombe Hospital Queen Alexandra Road High Wycombe HP11 2TT	Contact: Gail Bell Tel: +44 (0)1494 426054 E-Mail: gail.bell1@nhs.net Website: www.buckshealthcare.nhs.uk

Testing performed by the Organisation at the locations specified below

### Locations covered by the organisation and their relevant activities

#### Laboratory locations:

Location details	Activity	Location code	
<b>Address:</b> Wycombe Hospital Queen Alexandra Road, High Wycombe HP11 2TT	<b>Local contact</b>  Gail Bell	Histopathology, Gynaecological Cytopathology Diagnostic Cytopathology Rapid frozen section for intra-operative diagnosis Frozen sections for immunofluorescence studies Andrology (fertility and PVSA) OSNA (One Step Nucleic Acid Amplification) for the detection of breast cancer metastasis.	A
Stoke Mandeville Hospital Mandeville Road Aylesbury HP21 8AL United Kingdom	<b>Local contact</b>  Gail Bell	Rapid frozen section for intra-operative diagnosis	B



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**Site activities performed away from the locations listed above:**

Location details		Activity
Wycombe Hospital Mortuary Queen Alexandra Road, High Wycombe HP11 2TT	Gail Bell	Receipt and storage of bodies
Stoke Mandeville Hospital Mortuary Mandeville Road Aylesbury HP21 8AL	Gail Bell	Receipt and storage of bodies
Stoke Mandeville Hospital Mandeville Road Aylesbury HP21 8AL	Gail Bell	Storage of archived slides and PPFE blocks



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
<p><b>HUMAN TISSUES AND FLUIDS (cont'd)</b></p> <p>Body fluids: Fine needle aspirations and FNA material on slides Serous effusions Respiratory samples Bronchial washings Bronchial Brushings Transbronchial FNA) Bone aspirate Sputum Urine Gastrointestinal brushings Biliary brushing Synovial Fluid Cerebrospinal fluid Bile Fluid Cyst Fluids Hydrocele Fluid Nipple Discharge</p> <p>Slides prepared in-house from sample types listed above</p>	<p><u>Diagnostic Cytology</u></p> <p>Preparation and examination of cytological specimens to detect cytological abnormality to aid clinical diagnosis</p>	<p>Preparation/centrifugation In-house procedures: CPDC SOP 1 Diagnostic Cytology</p> <p>Shandon Cytospin 4 Centrifuge Thermo Scientific Sorvall ST 16</p> <p><u>Papanicolaou staining</u> Automated using in-house procedures: CPDC SOP 1 Diagnostic Cytology in conjunction with manufacturer's instructions using: Shandon Varistain Stainer 24-4</p> <p><u>Diff Quik Romanowski staining</u> Manual using in-house procedures: CPDC SOP 1 Diagnostic Cytology</p>	<p style="text-align: center;">A</p>



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HUMAN TISSUES AND FLUIDS (cont'd)  Slides prepared in-house from sample types listed above	<u>Diagnostic Cytology</u>  Morphological assessment and interpretation/diagnosis	Microscopy (qualitative analysis) In-house procedures: CPDC SOP 1 Diagnostic Cytology using microscopes as follows: Nikon Eclipse 80i Nikon Eclipse Ni-U Leica DMLB Olympus BX51 Olympus BX50 Nikon Eclipse 80i Leica DM4000 Nikon Eclipse E400	A
Synovial Fluids	Detection and identification of Calcium pyrophosphate and monosodium urate crystals	Polarising light microscopy (qualitative analysis) using in-house procedures: CPDC SOP 1 Diagnostic Cytology and: Nikon Eclipse E400 microscope	A



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<p>HUMAN TISSUES AND FLUIDS (cont'd)</p> <p>Formalin fixed tissues</p>	<p><u>Histopathology</u></p> <p>Preparation and examination to identify or exclude morphological &amp; cytological abnormalities</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p>Macroanalysis and dissection Manual method using: CP HCC SOP 1 CP HCC SOP2 CP HCC SOP3 CP HCC SOP5 CP HCC SOP7 CP HCC SOP8 CP HCC SOP9 CP HCC SOP10 CP HCC SOP11 CP HCC SOP12 CP HCC SOP13 CP HCC SOP14 CP HCC SOP16 CP HCC SOP17 CP HCC SOP18 CP HCC SOP19 CP HCC SOP20 CP HCC SOP21 CP HCC SOP22 CP HCC SOP23 CP HCC SOP24 CP HCC SOP25 CP HCC SOP26 CP HCC SOP27 CP HCC SOP28 CP HCC SOP29 CP HCC SOP30 CP HCC SOP31 CP HCC SOP32 CP HCC SOP33 CP HCC SOP34</p>	<p>A</p>



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<p>HUMAN TISSUES AND FLUIDS (cont'd)</p> <p>Formalin fixed tissues</p>	<p><u>Histopathology (cont'd)</u></p> <p>Preparation and examination to identify or exclude morphological &amp; cytological abnormalities</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p>CP HCC SOP35 CP HCC SOP36</p> <p><u>Decalcification</u> Manual method using: CP HCU SOP 6</p> <p><u>Tissue Processing</u> Automated method using: CP HCU SOP 14 Thermo Excelsior AS and ES tissue processors</p> <p><u>Embedding</u> Manual method using: CP HML SOP 1 and: Leica EG1150 Leica EG1160 Thermo Histostar</p> <p><u>Microtomy</u> Manual method using: CP HML SOP 3 CP HML SOP 4 CP HML SOP 5 and: Leica RM2135 Leica RM2235 Microm HM325 Thermo Finesse</p>	<p>A</p>



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology (cont'd)</u> Preparation and examination to identify or exclude morphological & cytological abnormalities (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	
Fresh tissue Frozen tissue	Intra-operative diagnosis Basophilic and eosinophilic tissue structures	<u>Rapid frozen section</u> Cryotomy and manual and automated Haematoxylin & Eosin (H&E) staining method using: CP HSS SOP 1 Appx 11 CP CG SOP 7 CP HCU SOP 20 and Thermo Cryostat Cryotome FSE Bright Cryostat OTF	A, B
Fresh skin and conjunctiva	Detection of:	<u>Manual immunofluorescent method using:</u> CP HML SOP 33 and Thermo Cryostat Cryotome FSE Olympus BH 2 microscope	A
	IgG	IgG	
	IgA	IgA	
	IgM	IgM	
	Complement (C3)	C3	



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology (cont'd)</u> Preparation and examination to identify or exclude morphological & cytological abnormalities (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	A
Formalin fixed, paraffin embedded (FFPE) tissue	Basophilic and eosinophilic tissue structures	<u>Routine Haematoxylin &amp; Eosin (H&amp;E) staining and coverslipping</u> Automated method using: CP HSS SOP 1 Appx 11 and: Gemini stainer AS	
	Detection of the following:	<u>Special staining</u> Manual methods using: CP HSS SOP 1 Appx 1-29 and the following stains:	
	Acid and sulphated mucins	Alcian Blue	
	Acidic and neutral mucins	Alcian Blue Periodic Acid-Schiff (AB PAS)	
	Elastic connective tissue fibres	Elastic van-Gieson (EVG) (Miller's)	
	Connective tissue and fibrin	Lendrum's Martius, Scarlet, Blue (MSB)	
	Campylobacter and Helicobacter sp.	Modified Giemsa stain	
	Organisms	Gram technique	
	Fungal hyphae and spores	Grocott Hexamine Silver	
	Pneumocystis sp.	Grocott	
	Chromatin	Haematoxylin and Eosin	
	Iron / haemosiderin	Perls Prussian Blue	





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HUMAN TISSUES AND FLUIDS (cont'd)  FFPE Tissue (unless otherwise stated)	<u>Histopathology (cont'd)</u>  Preparation and examination to identify or exclude morphological & cytological abnormalities (cont'd)  Detection of the following:  Melanin, lipofuscin and melanosis  Melanin and argentaffin granules  Muscle fibrin and connective tissue  Copper associated protein and Hepatitis B surface antigen  Glycogen, mucopolysaccharide components, fungi and basement membranes  Glycogen  Glycogen, acid and neutral mucins  Fibrin, muscle, and collagen  Reticulin fibres  Mast cells  Collagen connective tissue fibres  Leprosy bacilli  Acid and alcohol-fast bacilli	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)  <u>Special staining</u> Manual methods using: CP HSS SOP 1 Appx 1-29 and the following stains:  Mallory Bleach  Masson Fontana silver  Masson Trichrome  Orcein (Shikata)  Periodic Acid-Schiff (PAS) technique  Diastase Periodic Acid-Schiff (D PAS) technique  Periodic Acid Schiff Diastase Alcian Blue (DABPAS)  Picro Mallory  Reticulin (Gomori)  Thionin  Haematoxylin/Van-Gieson technique  Wade Fite technique  Ziehl-Neelsen (ZN) technique	A



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology (cont'd)</u> Preparation and examination to identify or exclude morphological & cytological abnormalities (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	A
FFPE tissue (cont'd)	Detection of the following:	<u>Immunohistochemistry</u> Automated methods using: CP HICC SOP 1 CP HICC SOP 2 CP HICC SOP 3 CP HICC SOP 4 and: Dako Autostainer and the following antibodies	
	Squamous epithelium. Stains all cell layers of epithelium. Prostate basal cells.	34BE12	
	Pan-keratin (Simple & complex epithelia)	AE1/AE3	
	Germ Cell tumours e.g. embryonal Ca and yolk sac tumour. Hepatocellular Ca.	Alpha Fetoprotein	
	Smooth Muscle cells and myoepithelial cells	Actin	
	B-catenin protein, binds to cytoplasmic tail of E-cadherin	B-catenin	
	For ID of follicular lymphoma	BCL2	
	Cell membrane glycoproteins expressed on healthy epithelia and in various carcinomas	BerEp4	



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HUMAN TISSUES AND FLUIDS (cont'd)	<u>Histopathology (cont'd)</u> Preparation and examination to identify or exclude morphological & cytological abnormalities (cont'd)	Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)	A
FFPE Tissue (cont'd)	Detection of (cont'd)	<u>Immunohistochemistry (cont'd)</u> Automated methods using: Dako Autostainer and the following antibodies	
	Endothelial cells in vascular and lymphatic spaces.	CD31	
	Endothelial cells in vascular and Lymphatic vessels	CD34	
	Lymphocytes, Histiocytes and macrophages	CD45	
	Peripheral nerves. ID of NK cells, NK-like T cells. Small cell lung Ca.	CD56	
	Islet cells of pancreas. Lymphocytes in tonsil. Squamous epithelial cells.	CD99	
	GIST tumours, mast cells and interstitial cells of Cajal in GI tract.	CD117	
	Enterocytes in the mucosa of appendix. Basal and luminal cells. For ID of GI adenocarcinomas and carcinoids	CDX2	
	Colonic epithelial cells. Expressed in colon Ca and lung adeno Ca.	CEA	
	Neuroendocrine cells in epithelial surface of colon. Nerve/ganglion cells.	CHROMO	



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<p>HUMAN TISSUES AND FLUIDS (cont'd)</p> <p>FFPE Tissue (cont'd)</p>	<p><u>Histopathology (cont'd)</u></p> <p>Preparation and examination to identify or exclude morphological &amp; cytological abnormalities (cont'd)</p> <p>Detection of (cont'd)</p> <p>Squamous epithelium in tonsil. Basal cells of the prostate.</p> <p>Epithelial cells in pancreatic ducts and in breast epithelial cells. For ID of tumours of unknown origin</p> <p>Basal layer of squamous epithelia in skin and prostate glands. Stratified squamous epithelia of tonsil and myoepithelial of breast.</p> <p>Neuroendocrine cells in the epithelial surface of the colon. Epithelial cells, neuron and ganglion cells.</p> <p>Squamous epithelial cells in tonsil.</p> <p>For ID of GI tumours, mucinous ovarian tumours.</p> <p>Cells infected with CMV</p> <p>Suprabasal layer of squamous epithelium in tonsil, scattered endothelial cells and fibroblasts.</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p><u>Immunohistochemistry (cont'd)</u></p> <p>Automated methods using: Dako Autostainer and the following antibodies</p> <p>CK5/6</p> <p>CK7</p> <p>CK14</p> <p>CHROMOGRANIN</p> <p>CK19</p> <p>CK20</p> <p>CMV</p> <p>CYCLIN D1</p>	<p>A</p>



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<p>HUMAN TISSUES AND FLUIDS (cont'd)</p> <p>FFPE Tissue (cont'd)</p>	<p><u>Histopathology (cont'd)</u></p> <p>Preparation and examination to identify or exclude morphological &amp; cytological abnormalities (cont'd)</p> <p>Detection of (cont'd)</p> <p>Normal proliferating cells; poorly differentiated squamous cell carcinoma, basal cells; abluminal cells</p> <p>Prostate adeno Ca</p> <p>ID of prostatic ductal epithelial cells</p> <p>Placental Alkaline Phosphatase in germ cell tumours, particularly seminomas; also, breast, lung, ovarian carcinomas</p> <p>Cellular progesterone receptor expression</p> <p>Prostate specific antigen</p> <p>ID of RCC vs non-renal cell tumours.</p> <p>Normal and neoplastic melanocytes; macrophages; adipocytes; nerve fibres; many tumour types</p> <p>Smooth muscle myosin</p>	<p>Macroscopic and Microscopic examination using in-house procedures (listed) in conjunction with manufacturer's instructions for the following methods (where relevant)</p> <p><u>Immunohistochemistry (cont'd)</u></p> <p>Automated methods using: Dako Autostainer and the following antibodies</p> <p>p63</p> <p>P504S</p> <p>PAP</p> <p>PLAP</p> <p>Progesterone receptor (PR)</p> <p>PSA</p> <p>RCC</p> <p>S100 protein</p> <p>SMM</p>	<p>A</p>



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FFPE Tissue (cont'd)	Detection of (cont'd)	<u>Immunohistochemistry (cont'd)</u> Automated methods using: Dako Autostainer and the following antibodies	
	Synaptic vesicle protein (neurones; neuroendocrine cells)	Synaptophysin	
	Stains cells lining blood and lymphatic vessels and syncytiotrophoblasts of placenta.	THROMBOMODULIN	
	ID of tumours of thyroid origin.	THYROGLOBULIN	
	Thyroid transcription factor-1 (Thyroid follicular & C cells)	TTF-1	
	Vimentin intermediate filament	Vimentin	
	ID of malignancies involving differentiated plasma cells.	VS398c	
	WT1 expression	WT1	
	<u>Molecular</u>	Manufacturer's instructions for Sysmex RD-210 one step nucleic acid amplification analyser in conjunction with documented in-house methods:	
Fresh tissue: Sentinel lymph nodes (axillary)	Expression of Cytokeratin 19 (CK19) mRNA	Detection and amplification of CK 19 mRNA  CP HOS SOP 1	A



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2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

**Buckinghamshire Healthcare NHS Trust**

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Testing performed by the Organisation at the locations specified

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
HUMAN TISSUES AND FLUIDS (cont'd)	<u>Andrology</u>	In house documented manual method in conjunction with BAS guidelines for the assessment of post vasectomy semen samples (2016)	A
Ejaculate	Post vasectomy examination: Presence or absence of spermatozoa	Manual preparatory technique and microscopy (reporting) <u>CPAND SOP 2</u>	
Ejaculate		<u>Semen Analysis</u> In house procedures following the guidelines below:  WHO Laboratory Manual for the Examination of Human Semen and Sperm-Cervical Mucus Interaction 5th Edition.	A
Ejaculate	Fertility screening:  Semen pH Semen volume  Sperm concentration Sperm morphology Sperm motility	Manual methods using in-house procedure: CPAND SOP1 Semen Analysis  Manual microscopy method using in-house procedure: CPAND SOP1 Semen Analysis Olympus BH-2  <u>Papanicolaou staining</u> Automated using in-house procedures: CPDC SOP 1 Diagnostic Cytology  in conjunction with manufacturer's instructions using: Shandon Varistain Stainer 24-4	

END