


Schedule of Accreditation

issued by

United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

| | | |
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|  9546 Accredited to ISO 15189:2022 | NHS Lothian | |
| | Issue No: 015 Issue date: 10 January 2025 | |
| | Royal Infirmary of Edinburgh 51 Little France Crescent Edinburgh EH16 4SA | Contact: Bernard Lawless E-Mail: Bernard.lawless@nhs.scot Website: www.edinburghlabmed.co.uk |
| Testing performed by the Organisation at the locations specified below | | |

Locations covered by the organisation and their relevant activities

Laboratory locations:

| Location details | Activity | Location code |
|---|--|---------------|
| Address: Royal Infirmary of Edinburgh 51 Little France Crescent Edinburgh EH16 4SA Local contact: Bernard Lawless | Microbiology: Molecular Virology Molecular Haematology Molecular Pathology | A |

Site activities performed away from the locations listed above:

| Location details | | Activity | Location code |
|--|--|--|---------------|
| Western General Hospital | Local contact: Bernard Lawless | Haematological Malignancy Diagnostic Service (HMDS) Immunophenotyping | B |
| Scottish Bacterial Sexually Transmitted Infection Reference Laboratory (SBSTIRL) | | National reference service for the management of sexually transmitted infection | C |
| Scottish E. coli O157/VTEC Reference Laboratory (SERL) | | Identification and typing of <i>E. coli</i> O157 and Shiga toxin-producing <i>E. coli</i> | D |
| Scottish HPV Reference Laboratory (SHPVRL) | | Screening and typing of Human Papillomavirus | E |
| Scottish Mycobacteria Reference Laboratory (SMRL) | | Identification, drug susceptibility testing and strain typing of <i>Mycobacterium tuberculosis</i> complex (MTBC) and non-tuberculous mycobacteria (NTM) | F |
| Blood Borne Virus Specialist Testing Service (BBVSTS) | | Sequencing of Blood Borne viruses | G |
| Viral Sequencing Service (VSS) - RIE | | Genome sequencing of viruses and other pathogens | H |
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DETAIL OF ACCREDITATION

| Materials/Products tested | Type of test/Properties measured/Range of measurement | Standard specifications/ Equipment/Techniques used | Location Code |
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| HUMAN TISSUES and FLUIDS | Microbiological examination activities for the purpose of clinical diagnosis | In-house documented procedures: | |
| Enrichment broths from the following sample types: Faeces and isolates of <i>E. coli</i> . Mixed cultures of bacterial growth submitted as slopes or pellets from centrifuged culture broth | DNA Extraction | Measurement Principle: Manual DNA extraction using SOP: SERL 50 | D |
| Nucleic Acid | Detection of stx1 stx 2 and rfbO157 genes | Real-Time PCR using BioRad C1000 Touch Thermal Cyclers using SOP: SERL 50 | D |
| Cultures isolates of <i>E. coli</i> | Identification of <i>E. coli</i> | Identification by BioMerieux API 20E, using SOP: SERL 58, SERL 53 | D |
| | Determination of MLST (7 loci), virulence gene profile, serotype (O:H antigen), cgMLST (2513 loci) and SNP analysis for outbreak investigation and surveillance of <i>E. coli</i> O157 and non-O157 STEC. | DNA extraction using the Qiagen DNeasy Blood and Tissue Kit using SOP: SERL137 | D |
| | | Use of the Qubit 3.0 Fluorometer using SOP: SERL144 | D |
| | | Nextera XT DNA Library Preparation using SOP: SERL 141 | D |
| | | Whole genome sequencing on the Illumina MiSeq using SOP: SERL 145 | D |
| | | Analysis of Whole Genome Sequencing Data using the UKHSA pipeline using SOP: SERL140 | D |
| | | Analysis of Whole Genome Sequencing Data using BioNumerics (BioMérieux) using SOP: SERL139 | D |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| Swabs, isolates | Culture and identification of <i>Neisseria gonorrhoeae</i> | Culture on agar followed by phenotypic identification using Gram stain, sugar utilisation, using SOPs: SBSTIRL39, SBSTIRL40 | C |
| Isolates | Antibiotic susceptibility of <i>Neisseria gonorrhoeae</i> | Antibiotic susceptibility testing by agar dilution method and E tests using SOP: SBSTIRL41 | |
| Swabs, urine, and aspirates | DNA Extraction | BioMerieux EasyMag or eMAG automated extractor, or manual method by Qiagen QIAamp DNA kit using SOPs: VIRM-3, SBSTIRL34 | A, C |
| DNA | Screening for <i>Chlamydia trachomatis</i> LGV serovars | Real-time PCR by ABI 7500 using SOP: SBSTIRL31 | C |
| Rectal, Genital, and Urines. | Screening for <i>Chlamydia trachomatis</i> LGV serovars | Real-Time PCR using the ABI7500 using SOP: SBSTIRL31 | C |
| Male urine, male anal swabs, female urine, cervical swabs, endocervical swabs, vaginal and high vaginal swabs | Detections of mutations in <i>Mycoplasma Genitalium</i> that correlate to macrolide and fluoroquinolone resistance | Automated extraction using EasyMag, eMAG, or manual extraction using QIAamp. PCR using Gel Electrophoresis visualisation or Sanger Sequencing using SOP SBSTIRL74, VSS-16 | C |
| Urine, Urogenital Swabs and Rectal Swabs | Detection of <i>Mycoplasma genitalium</i> and key mutations associated with macrolide resistance | ResistancePlus MG Flexible assay on the Cepheid GeneXpert using SOP: VIRM355 | A |
| Rectal, cervical, throat Swabs/Urine | Detection of <i>N. gonorrhoeae</i> DNA | Seegene Allplex ST14 Kit using SOP VIRM-337 | A |
| Hologic Liquid based cytology cervical samples (Thinprep Preservcyt) | Detection of high-risk HPV types: 16 & 18/45 and pool: 31,33,35,39,51,52,56,58,59,66 and 68. | Cepheid GeneXpert PCR Using SOP HPV112 | E |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| Liquid based cytology cervical samples (Thinprep Preservcyt) and Formalin Fixed Paraffin material – head and neck, and anogenital biopsies | Detection of HPV types: 19 high-risk types (16,18,26,31,33,35,39,45,51,52,53,56,58,59,66,68,69,73,82) and 9 low-risk HPV types (6,11,40,42,43,44,54,61,70) | SeeGene Allplex HPV28 kit SOP HPV157 Manual extraction using QIAamp DNA mini kit or automated extraction using Seegene STARMag 48 kit Combination of manual and automated stages and PCR set up using the Seegene Nimbus platform Detection using BioRad CFX and Seegene software | E |
| Positive Mycobacterium culture | DNA extraction | Genomic DNA extraction from a positive Mycobacterium culture for whole genome sequencing using SOP: SMRL139 | F |
| Nucleic acids obtained from Respiratory specimens | Molecular detection of <i>M. tuberculosis</i> complex (MTBC) and its resistance to rifampicin (RIF) | Real time nested PCR via Cepheid GeneXpert platform using SOP: SMRL69 | F |
| Mycobacteria growth indicator tube (MGIT) cultures | Detection of MTBC | Rapid immunochromatographic assay for MTBC antigen using SOP: SMRL19 | F |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| DNA | Mycobacteria Whole Genome Sequencing | Use of the Qubit 3.0 Fluorometer using SOP: SERL144 | F |
| | | Nextera XT DNA Library Preparation using SOP: SERL 141 | F |
| | | Whole genome sequencing on the Illumina MiSeq using SOP: SERL 145 | F |
| | | MTB phenotypic drug susceptibility testing based on WGS results using SOP: SMRL 151 | F |
| | | WGS result verification and reporting using SOP: SMRL 152, SERL149 | F |
| | | WGS data transfer to UKHSA server using FileZilla SOP: SMRL150 | F |
| Cultures | Susceptibility testing of <i>Mycobacterium tuberculosis</i> complex (MTBC) | BACTEC MGIT TM 960 fluorometric system using SOP: SMRL9 | F |
| Raw sputum or concentrated sediments prepared from sputum or MGIT culture | Detection of M. tuberculosis and resistance genes (katG, fabG1, oxyR-ahpC, inhA, gyrA, gyrB, rrs and eis) | Cepheid GeneXpert MTB/XDR assay using SOP SMRL 171 | F |
| Respiratory samples, pleural fluids, aspirates, pus, tissue, stools, CSF, and bone marrow. | Detection of Acid Fast Bacilli (AFB) | Fluorescent microscopy stained with Auramine Phenol using SOP: SMRL3 & SMRL17 | F |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| Respiratory samples, pleural fluids, aspirates, pus, tissue, stools, CSF, bone marrow, urine, whole & citrated blood culture | Culture, identification for Mycobacteria | Culture via solid and/or liquid media followed by BD BACTEC MGIT 960 or BD BACTEC Fx using SOP: SMRL17, SMRL6 | F |
| | | Kinyoun modification of Ziehl-Neelsen staining using SOP: SMRL21 | F |
| | <u>Molecular Microbiological examination activities for the purpose of clinical diagnosis</u> | In-house documented procedures: | |
| Nose and Throat Swabs in VTM or MSS | Detection of SARS CoV-2 virus RNA (COVID-19) specific gene sequences; • N2 and E | GeneXpert real-time system SOP: VIRM-299 | A |
| Nose and Throat Swabs in VTM or MSS | Detection of SARS-CoV-2 RNA (RdRp and N gene target sequences) | Real-Time PCR using the Abbott Resp-4-Plex assay on the Abbott Alinity M using SOP: VIRM 330 | A |
| Nose and Throat Swab in VTM or MSS | Detection of SARS-CoV-2 (N gene target sequences x2) | Real-Time PCR using the RP2 Panel on the Genmark ePlex using SOP: VIRM 314 | A |
| Dried Blood Spot | Detection of HCV RNA | In-house RT-PCR assay using EasyMag or eMAG automated extraction and ABI 7500 FAST using SOP VIRM-258, VSS-16 | A |
| Viral RNA | HCV genotyping | Sanger sequencing of HCV (various regions) via ABI 3500xl Genetic Analyser using SOP: BBV1, BBV4, BBV 33 | G |
| Viral RNA or DNA | HIV sequencing | Sanger sequencing of HIV (various regions) via ABI 3500xl Genetic Analyser using SOP: BBV1, BBV8, BBV5, BBV7, BBV11 | G |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Molecular Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| Plasma/serum/CSF | Viral nucleic acid extraction | Biomerieux EasyMag or eMAG automated nucleic acid extraction platform using SOP: VIRM-3, VSS-16 | G |
| Whole EDTA blood, Peripheral blood mononuclear cell's (PBMC's) | Viral DNA extraction | Manual DNA extraction via QIAGEN QIAamp DNA Blood Mini kit using SOP: BBV2 | G |
| Vesicle / Skin Swab | Detection of Orthopoxvirus DNA | EasyMag extraction platform and Altona RealStar Zoonotic Orthopoxvirus PCR kit 1.0 and ABI 7500 thermocycler. SOP: VIRM-358 | A |
| Vesicle / Skin Swab | Detection of Monkeypox virus DNA | In-house assay and EasyMag extraction platform and ABI 7500 thermocycler. SOP: VIRM-392 | A |
| Faeces | Detection of Norovirus (G1 and G2), Rotavirus, Adenovirus (F40/41), Astrovirus, Sapovirus (G1, 2, 4,5) nucleic acids | Hamilton STARlet, Seegene Allplex GI Plus kit and BioRad CFX 96 thermocycler. SOP: VIRM-337 | A |
| Respiratory secretions, respiratory swabs, genital swabs, vesicle fluids, vesicle swabs, tissues, whole blood, blood spots, urine, swabs, body fluids, faeces, vomit, CSF | Nucleic acid extraction | Biomerieux Easymag or eMAG Automated Extractors or Qiagen Manual Extraction SOP: VIRM-3, VIRM-71, VSS-16 | A |
| RNA/DNA Extracts | Detection of Influenza A, B, Respiratory Syncytial virus (RSV), Para-influenza type 1-3, Adenovirus, Mycoplasma pneumoniae, Rhinovirus, Human Metapneumovirus | Respiratory Multiplex PCR via ABI 7500 using SOP: VIRM-67, VIRM-5, VIRM-11, VIRM-19, VIRM-20 | A |



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| Respiratory samples | SARS-CoV-2 whole genome sequencing | SARS-CoV-2 whole genome sequencing (Oxford Nanopore) using Oxford Nanopore Technologies GridION using SOPs: VSS-3, VSS-9, VSS-11, VSS-12, VSS-15) | H |
| RNA/DNA Extracts | Typing of Swine Dublin H1N1 | ABI 7500 Real-Time PCR analyser using | A |
| | H1N1 H275Y Resistance testing | In-house RT-PCR assay using EasyMag or eMAG automated extraction and ABI 7500 FAST and associated SOP: | |
| | Influenza virus Typing and Confirmation (H1/H3/H5/H7) | SOP: VIRM-38 | |
| | Detection of Mumps | SOP VIRM-68 | |
| | Detection of Measles | SOP: VIRM-55, H1/H3 SOP VIRM-55, H5 SOP VIRM-30, H7 SOP VIRM-105 | |
| | Detection of Syphilis | SOP: VIRM-18, VIRM-62 | |
| | Detection of <i>Bordetella pertussis</i> | SOP: VIRM-61 | |
| | Detection of Enterovirus/Parecho | SOP: VIRM-40, VIRM-50 | |
| | Detection of Enterovirus Typing (EVD68) | SOP: VIRM-46, VIRM-48 | |
| | Detection of Herpes Simplex Virus (HSV), Varicella Zoster Virus (VZV), Adenovirus | SOP: VIRM-8, VIRM-13, VIRM-14, VIRM-28 | |
| | | SOP: VIRM-14 | |
| | | SOP: VIRM-20, VIRM-26, VIRM-31, VIRM-58, VIRM-104 | |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Molecular Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| RNA/DNA Extracts (cont'd) | Detection of Rotavirus | ABI 7500 Real-Time PCR analyser using SOP: VIRM-35, VIRM-70 | A |
| | Detection of Middle eastern respiratory syndrome (MERS) | SOP: VIRM-136 | A |
| | Detection of Bocavirus | SOP: VIRM-92 | A |
| | Detection of Coronavirus | SOP: VIRM-103 | A |
| | Detection of Bacterial Meningitis (<i>Neisseria meningitidis</i> , <i>Haemophilus pneumoniae</i> , and <i>Streptococcus pneumoniae</i>) | SOP: VIRM-47, VIRM-39 | A |
| | Detection of Viral Haemorrhagic Fever (Malaria, Ebola, Marburg, Lassa, Crimean Congo, and Dengue) | SOP: VIRM-111, VIRM113 | A |
| | Detection of Astrovirus and Sapovirus | SOP: VIRM-242 | A |
| | Detection of Parvovirus B19 | SOP: VIRM-123, VIRM-124 | A |
| | Detection of Hepatitis E | SOP: VIRM-12, VIRM-56 | A |
| | Detection of <i>Chlamydia pneumophila</i> | SOP: VIRM-114, VIRM-115, VIRM-116 | A |
| | Detection of Legionella (<i>L. pneumophila</i> /species) | SOP: VIRM-16, VIRM-49 | A |
| | Detection of Epstein-Barr Virus (EBV) | ABI 7500 Quantitative Real-Time PCR analyser using SOP: VIRM-2, VIRM27 | A |
| | Detection of Plasmodium spp | Altona Real Star Pan-Malaria assay using EasyMag, or eMAG automated extraction and ABI 7500 FAST using SOP VIRM-253, VSS-16 | A |



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| HUMAN TISSUES and FLUIDS (cont'd) | <u>Molecular Microbiological examination activities for the purpose of clinical diagnosis</u> (cont'd) | In-house documented procedures: | |
| Nose and Throat swabs in VTM or MSS | Detection of SARS CoV-2 virus RNA (COVID-19) specific gene sequences; <ul style="list-style-type: none"> E and S | ABI 7500 Real-Time PCR analyser using EasyMag, or eMAG extraction and Altona RealStar SARS-CoV-2 kit using ABI 7500 Fast Dx SOP: VIRM-290, VSS-16 | A |
| RNA/DNA Extracts | Detection of BK virus | Quantitative Real-time PCR via VELA Santosa SX using SOP: VIRM-177 | A |
| | Enterovirus Typing (VP1) | Sanger sequencing using ABI 3500xl Genetic Analyzer. | A |
| Clotted Blood/EDTA | CMV Quantitation | Vela Diagnostics Sentosa Real-time PCR using SOP: VIRM 119 | A |
| Stool | Detection of enteric parasites <ul style="list-style-type: none"> <i>Cryptosporidium</i> spp, <i>Giardia lamblia</i> <i>Entamoeba histolytica</i> | BioMerieux EasyMag, or eMAG automated extractor and Qiagen QIAamp DNA kit using SOP: VIRM-3, VSS-16 with detection by real-time PCR using the ABI 7500 thermocycler using SOP: SBSTIRL31 Reporting SOP: VIRM 247 | A |
| Boiled Crude lysate | Carbapenemase Producing/ Resistant Enterobacteriaceae | Biorad C1000 TouchReal-Time PCR analyser using SOP: VIRM-33, VIRM-73, VIRM-138, VIRM-139 | A |
| EDTA | Viral Load for HIV, HCV and HBV | Abbott Alinity System – Real-Time PCR extraction and amplification analyser using SOP: VIRM-302 | A |
| Urine | Detection of <i>Legionella pneumophila</i> serotype 1 antigen | BinaxNow Legionella kit by membrane immunochromatographic assay using SOP: VIRM-86 | A |



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| HUMAN BODY FLUIDS and TISSUES | <u>Immunophenotyping</u> <u>Testing for use in clinical diagnosis</u> | Documented In-house methods using: | |
| Whole Blood | <ul style="list-style-type: none"> • BD TBNK AssayCD3 • CD4 • CD8 • CD16 • CD19 • CD56 | BD FACS Lyric flow cytometer SOP: HAEM-W-927 and HAEM-W-928 | B |
| Whole Blood and Bone Marrow | Leukaemia/Lymphoma Immunophenotyping Antibodies listed in cocktails for CLL and Acute Leukaemia Panels: CD2, CD3, CD4, CD5, CD7, CD8, CD10, CD11c, CD13, CD15, CD16, CD19, CD20, CD22, CD23, CD25, CD33, CD34, CD45, CD56, CD79b, CD103, CD117, FMC7 HLADR, Oncomark reagent CD14/CD64, Kappa and Lambda Intracytoplasmic marker in panels: Oncomark reagent CD3, MPO, CD79a, TdT | BD FACS Lyric flow cytometer HAEM-W-927 and HAEM-W-929 | B |
| Whole Blood | Investigation of Paroxysmal Nocturnal Haemoglobinuria (CD59, CD235a, CD14, CD15, CD24, CD45. CD64, FLAER) | BD FACS Lyric flow cytometer HAEM-W-947 | B |



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| HUMAN BODY FLUIDS and TISSUES (cont'd) | <u>Molecular Haematology activities for the purpose of clinical diagnosis</u> | Documented In-house methods using: | |
| Whole Blood EDTA and NaCitrate | Extraction of nucleic acid | QiaSymphony SP: Purification of DNA according to SOP HAEM-R32 | A |
| DNA Extracted from whole blood/ bone marrow | Detection of JAK2 V617F mutation | Applied Biosystems Fast 7500 real time PCR Analyser using manufacturers' instructions and SOP: HAEM-R109 | A |
| | Detection of JAK2 exon 12 mutations and MPL exon 10 mutations | Real-time PCR amplification using the Rotorgene and analysis by High resolution melt curve analysis (HRM). SOP: HAEM R115, R126 | A |
| | Detection of JAK2 exon 12 mutations and MPL exon 10 mutations | PCR Amplification using thermal cyclers and direct DNA sequence analysis using Applied Biosystems 3500xl analyser using manufacturers' instructions SOP: HAEM R115, R126 | A |
| | Detection of prothrombin G20210A and Factor V Leiden mutations | Real-time amplification using the Applied Biosystems® Fast 7500 real time PCR analyser using SOP: HAEM-R19 | A |
| | Detection of CALR mutations. | PCR Amplification using thermal cyclers, and fragment analysis of PCR product by Applied Biosystems 3500xl analyser using manufacturers' instructions and SOP: HAEM-R113 | A |
| | Detection of CALR mutations. | PCR Amplification using thermal cyclers, and Sanger sequencing by Applied Biosystems 3500xl analyser using manufacturers' instructions and SOP: HAEM-R113 | A |
| DNA/RNA extracted from blood/bone marrow | NGS Myeloid panel | ThermoFisher Oncomine Myeloid Assay by NGS by Ion Chef and Ion Torrent S5 Sequencer using SOP HAEM-W-857 and HAEM-W-724 | A |
| | DPYD | DPYD testing using the Elucigene DPYD kit, SOP PATH-R-399 | A |



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| HUMAN BODY FLUIDS and TISSUES (cont'd) | <u>Molecular Haematology activities for the purpose of clinical diagnosis (cont'd)</u> | Procedures documented in manufacturer's equipment manuals in conjunction with documented in- house procedures by the following methods: | |
| DNA extracted from peripheral blood | DPYD variant testing | PCR and Sanger sequencing using ABI 3500X according to SOP- PATH-R-184 | A |
| DNA extracted from FFPE | Microsatellite Instability Testing | PCR/Fragment length analysis using Promega MSI Oncomate kit thermocyclers and Applied Biosystems 3500xl analyser using SOP PATH-R-473 | A |
| | NGS Caner Hotspot | ThermoFisher AmpliSeq Cancer Hotspot version 2 NGS by Ion Chef and Ion Torrent S5 sequencer using SOP PATH-R-358 and PATH-R-364 | A |
| Blood, Bone Marrow | | Automated extraction of genomic DNA using the Promega Maxwell RSC instrument using SOP: HAEM-W-948 | B |
| | | Automated extraction of RNA using the Promega Maxwell RSC instrument as described in SOP HAEM-W-581 and HAEM-W-937 | B |
| Extracted RNA | cDNA synthesis | cDNA synthesis using the VILO mastermix performed on Thermal Cyclers using SOP: HAEM-W-731 | B |



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| Synthesised cDNA | Detection of: <ul style="list-style-type: none"> FIPL1-PDGFR fusion gene | Reverse transcriptase PCR on Thermo cyclers and Gel electrophoresis SOP: HAEM-W-578 SOP: HAEM-W-286 | B |
| Synthesised cDNA | Detection of: <ul style="list-style-type: none"> BCR: ABL 1 fusion gene | Realtime RT-PCR ABI 7500 | B |
| Nucleic Acid | Detection of <ul style="list-style-type: none"> FLT3 internal tandem duplication and tyrosine kinase domain mutation | multiplex PCR on Thermal cyclers and capillary electrophoresis and the ABI 3500 genetic analyser using SOP: HAEM-W-586 | B |
| Synthesised cDNA | Relative Quantification of <ul style="list-style-type: none"> BCR: ABL 1 | Quantitative realtime RT-PCR using the ABI 7500 SOP: HAEM-W-205 (E13A2 and E14A2 fusion transcript) and SOP HAEM-W-586 (E1A2 fusion transcript) | B |
| | BCR: ABL 1 TKD mutation | Nested PCR and ABI 3500XL genetic analyser Sanger sequencing using SOP:HAEM-W-791 | B |
| Nucleic Acid | IGHV mutation status | PCR and Sanger sequencing using ABI 3500XL according to SOP: HAEM-W-796 | B |
| Nucleic Acid | NPM1 mutation | PCR and capillary electrophoresis using the ABI 3500XL genetic analyser using SOP: HAEM-W-835 | B |
| Nucleic Acid | Quantification of NPM1 mutation | Quantitative realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-907 | B |
| Blood/Bone Marrow | Isolation of human mononuclear cells | Centrifugation for sample separation sing Lymphoprep using HAEM-W-806 | B |
| Nucleic Acid | Quantification of Nucleic Acid | Use of Nanodrop ND1000 according to HAEM-R3 | B |
| Synthesised cDNA | PCR amplification | Veriti Thermocycler | B |



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| Materials/Products tested | Type of test/Properties measured/Range of measurement | Standard specifications/ Equipment/Techniques used | Location Code |
|--|---|---|---------------|
| HUMAN BODY FLUIDS and TISSUES (cont'd) | <u>Molecular Haematology (HMDS) Testing for use in clinical diagnosis (cont'd)</u> | Documented In-house methods using: | |
| Synthesised cDNA | Acute Myeloid Leukaemia (AML) and Acute Lymphoblastic Leukaemia (ALL) fusion gene Panel RUNX1::RUNX1T1 CBFB::MYH11 PML::RARA ETV6::RUNX1 TCF3::PBX1 BCR::ABL1 (E1A2) <ul style="list-style-type: none"> BCR::ABL1 (E13A2; E14A2) | Realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-619 (AML fusion panel) Realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-635 (ALL fusion panel) Realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-636 (ALL and AML fusion follow up) | B |
| Synthesised cDNA | Quantification of RUNX1::RUNX1T1 | Quantitative realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-915 | B |
| Synthesised cDNA | Quantification of PML::RARA | Quantitative realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-916 | B |
| Synthesised cDNA | Quantification of CBFB::MYH11 | Quantitative realtime RT-PCR using the ABI 7500 according to SOP HAEM-W-917 | B |
| Extracted DNA | T-cell Clonality tests <ul style="list-style-type: none"> TCRB TCRG TCRD | Multiplex PCR with Invivoscribe Identiclone Kits on Thermal cyclers, ABI 3500 Genetic Analyser and using SOP: HAEM-W-569, SOP:HAEM-W-570 | B |
| | B-cell Clonality tests <ul style="list-style-type: none"> IGH IGL IGK IGKDEL | | B |
| Extracted DNA | MYD88 L265P Mutation screening | Real-time PCR using the ABI 7500 by Allele Specific PCR using SOP: HAEM-W-720 | B |
| Extracted DNA | KIT D816V Mutation screening | Real-time PCR using the ABI 7500 by Allele Specific PCR using SOP: HAEM-W713 | B |



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| HUMAN BODY FLUIDS and TISSUES (cont'd) | <u>Molecular Haematology (HMDS) Testing for use in clinical diagnosis (cont'd)</u> | Documented In-house methods using: | |
| Extracted DNA | Detection of TP53 mutation | PCR and Sanger sequencing using ABI 3500XL according to SOP: HAEM-W-721 and HAEM-W-722 | B |
| | Detection of POLE mutation | | |
| | Detection of NPM1 mutation | | |
| | <u>Molecular Pathology tests to assist in detection of clinical abnormalities</u> | In house, manual methods and manufacturer's instructions documented | A |
| Plasma | DNA profiling for detection of abnormal gene sequences: | Manual Isolation of Cell-Free DNA from Plasma Using the COBAS cfDNA Sample Preparation Kit (MOLG-R-14) | A |
| FFPE various tissue samples | Extraction of nucleic acid | Documented in- house methods: Manual DNA extraction using one or a combination of the techniques below by In-house procedures using commercial kits and manual extraction SOP: PATH-R-120 | A |
| | | Extraction of DNA & RNA using the Promega Maxwell 16 Instrument and associated kits (PATH-R-379) | A |
| DNA extracted from FFPE and cell free DNA from plasma | EGFR | PCR amplification of DNA using Roche COBAS for mutation detection using in-house method SOP: PATH-R-174 | A |
| DNA extracted from FFPE | KRAS | Pyromark Q24 pyrosequencing for mutation detection using SOP: PATH-R-134, PATH-R-334 | A |
| | NRAS | Pyromark Q24 pyrosequencing for mutation detection using SOP- PATH-R-134, PATH-R-334 | A |
| | BRAF | Pyromark Q24 pyrosequencing for mutation detection BRAF using SOP- PATH-R-134, PATH-R-334 | A |



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| HUMAN BODY FLUIDS and TISSUES (cont'd) | <u>Molecular Pathology tests to assist in detection of clinical abnormalities</u> (cont'd) | In house, manual methods and manufacturer's instructions documented | |
| DNA extracted from FFPE | <ul style="list-style-type: none"> KIT PDGFRA EGFR KRAS TERT Promoter FOXL2 c.402C>G | PCR and Sanger sequencing using ABI 3500XL according to SOP: PATH-R-184 | A |
| | MGMT promoter methylation | Pyromark Q24 pyrosequencing for methylation analysis using SOPs PATH-R-134 and PATH-R-144 | A |
| | Molecular identity testing | Multiplex PCR followed by capillary electrophoresis using the ABI 3500 genetic analyser using SOP-PATH-R-149 | A |
| DNA extracted from FFPE | IDH 1 and IDH 2 somatic mutations | Pyromark Q24 pyrosequencing for mutation detection using SOP- PATH-R-134, PATH-R-334 | A |
| | <u>FISH analysis to assist in detection of clinical abnormalities</u> | In House manual methods and manufacturer's instructions documented | |
| FFPE tissue samples and cell blocks | ALK rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | DDIT3 using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | EWSR1 rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | FOXO1 rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |



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| HUMAN BODY FLUIDS and TISSUES (cont'd) | <u>FISH analysis to assist in detection of clinical abnormalities</u> (cont'd) | In House manual methods and manufacturer's instructions documented | |
| FFPE tissue samples and cell blocks | FUS rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | HER2 amplification for therapy stratification in gastric cancer | FISH Analysis Manual staining procedures (gastric): ICC-R41 Fluorescence Microscopy Interpretation and scoring: ICC-R46 | A |
| FFPE tissue samples and cell blocks | ROS1 rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | USP6 rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | MDM2 amplification by enumeration FISH | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |
| FFPE tissue samples and cell blocks | SS18 rearrangement using breakapart FISH probe | FISH Analysis Manual staining procedures: ICC-R41 Fluorescence Microscopy: ICC-R25 Interpretation and scoring: MOLG-R-107 | A |

END