


# Schedule of Accreditation

issued by

## United Kingdom Accreditation Service

2 Pine Trees, Chertsey Lane, Staines-upon-Thames, TW18 3HR, UK

 <b>Accredited to ISO 15189:2012</b>	<b>HSL (Analytics) LLP</b>	
	Issue No: 002    Issue date: 01 February 2022	
	<b>Department of Parasitology</b> HSL Analytics (LLP) The Halo Building 1 Mabledon Place London WC1H 9AJ	<b>Contact: Spencer Polley</b> Tel: +44 (0) 20 7307 7373 E-Mail: <a href="mailto:spencer.polley@hslpathology.com">spencer.polley@hslpathology.com</a> Website: <a href="http://www.hslpathology.com">www.hslpathology.com</a>
<b>Testing performed by the Organisation at the locations specified below</b>		

### Locations covered by the organisation and their relevant activities

#### Laboratory locations:

Location details	Activity	Location code
<b>Address</b> Department of Clinical Parasitology The Halo Building 1 Mabledon Place London WC1H 9AJ	<b>Local contact</b> Spencer Polley  Tel: +44 (0) 7307 7373 <a href="mailto:spencer.polley@hslpathology.com">spencer.polley@hslpathology.com</a>	Clinical Parasitology  A

#### Site activities performed away from the locations listed above:

Location details	Activity	Location code
Specimen Reception / Hot-Lab 3rd Floor Mortimer Market Centre Capper Street London WC1E 6JB	Clinical Parasitology	B
Various locations adjacent to surgical procedures within University College London Hospital	Clinical Parasitology	C



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DETAIL OF ACCREDITATION

Materials/Products tested	Type of test/Properties measured/Range of measurement	Standard specifications/ Equipment/Techniques used	Location Code
HUMAN BODY TISSUES, FLUIDS AND ASSOCIATED SPECIMENS	<u>Clinical Parasitology</u> Parasitological examination activities for the purposes of clinical diagnosis. Detection of:	In house documented procedures based on equipment manuals and standard methods as specified:	
Blood,	Plasmodium falciparum, P. knowlesi, P. ovale spp, P. malariae and P. vivax, Trypanosoma spp., Babesia spp., and Borrelia spp.	General microscopy based techniques including wet preparation (inc thick and thin films), concentration methods and staining using Romanowsky, Giemsa & Rapid Fields stains PAR-MSY-001, PAR-MSY-004, PAR-MSY-003 and PAR-MSY-013	B
Slit skin smears	Leishmania spp.	General microscopy based techniques PAR-MSY-013	
Cyst fluid, sputum and bronchial lavage	Echinococcus granulosus and ova of parasites	Vital staining with Eosin and microscopy PAR-MSY-010	A
Blood, skin snips	Microfilaria	Direct microscopy, filtration and centrifugation followed by Giemsa & Haematoxylin stains and microscopy PAR-MSY-004	A
Clear adhesive tape from rectal area	Enterobius vermicularis	Direct microscopy PAR-MSY-014	A
Hair and skin scrapings, slides and matrices of non-human origin including carpet fibres	Eggs, body parts and whole parasites	Dissection and direct light microscopy PAR-MSY-008	A
Parasitic or non-parasitic organisms (whole or partial)	Differentiation of parasitic vs non parasitic helminths and anthropods	Direct light microscopy PAR-MSY-014 and PAR-MSY-009	A



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HUMAN BODY TISSUES AND FLUIDS	<u>Clinical Parasitology</u> (cont'd) Parasitological examination activities for the purposes of clinical diagnosis. Detection of:	In house documented procedures based on equipment manuals and standard methods as specified:	
Stool, duodenal aspirates, biopsies, string tests, rectal scrapes, rectal snips and corneal scrapes	Intestinal parasites, ova and cysts	General microscopy based techniques including wet preparation, concentration using Parasep and staining using Rowmanowsky, modified Ziehl-Neelson, Iodine, Modified Trichrome PAR-MSY-005, PAR-MSY-007, PAR-MSY-006 and PAR-MSY-010	A
Urine, semen and rectal snips	Helminth ova and parasites	Direct microscopy, filtration and centrifugation followed by microscopy PAR-MSY-011	A
Stool, liver aspirates, biopsies and corneal scrapes	Parasite DNA of intestinal protozoa: Entamoeba histolytica, Giardia intestinalis, Cryptosporidium spp., Enterocytozoon bienewisi, Encephalitozoon cuniculi, Encephalitozoon hellem, Encephalitozoon intestinalis or Pleistophora species	Extraction, amplification and detection of parasitic DNA by multiplex / SYBR Green Real Time PCR using QiaSymphony and Rotorgene PAR-MOL-001, PAR-MOL-005, PAR-MOL-006, PAR-MOL-008	A
Blood, bone marrow, blood spots	DNA of Plasmodium spp.	Extraction, amplification and detection of parasitic DNA by modified 'Snounou' PCR reaction using Qiagen and Proflex PAR-MOL-001, PAR-MOL-003 and PAR-MOL-004	A
Blood, slit skin smear, biopsies, bone marrow aspirate, bone marrow trephine, histological tissue sections and vitreous fluid	DNA of Leishmania spp.	Extraction, amplification and detection of parasitic DNA by PCR using Qiagen, Proflex and Rotorgene PAR-MOL-001, PAR-MOL-002 and PAR-MOL-004	A



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HUMAN BODY TISSUES AND FLUIDS (cont'd)	<u>Clinical Parasitology</u> (cont'd) Parasitological examination activities for the purposes of clinical diagnosis. Detection of:	In house documented procedures based on equipment manuals and standard methods as specified:	
Serum and CSF	Total IgG antibodies against: Schistosoma spp. Microfilaria spp. Echinococcus spp	In-house developed ELISA assays using Manual Techniques PAR-SER-006, PAR-SER-007 and PAR-SER-008	A
Serum and CSF	Total IgG2 antibodies against Echinococcus spp	In-house developed ELISA assays using Manual Techniques PAR-SER-009	A
Serum	Total IgG antibodies against: Toxocara spp. Strongyloides sp.	Commercial ELISAs assays using Gemini automated ELISA station PAR-SER-013 and PAR-SER-010	A
Plasma and serum	Human anti-parasite antibodies against Plasmodium spp and Trypanosoma cruzi.	Commercial ELISA assay using Gemini automated ELISA station PAR-SER-004 & PAR-SER-005	A
Serum, plasma, CSF	Taenia solium circulating antigen	In-house developed ELISA assays using Gemini automated ELISA station PAR-SER-012	A
Serum	Human anti-parasite antibodies to Trichinella spp. and Fasciola spp.	In-house developed IFAT PAR-SER-003	A
Serum	Human anti-parasite antibodies to Entamoeba histolytica	In-house developed CAP and IFAT methods PAR-SER-001	A
Serum	Human anti-parasite antibodies to Babesia microti, Plasmodium spp and Trypanosoma cruzi	In-house developed IFAT methods PAR-SER-002, PAR-SER-004 & PAR-SER-005	A
Serum	Human anti-parasite antibodies to Entamoeba histolytica	Bordier commercial ELISA assay using Gemini automated ELISA station SOP PAR-MP4-30	A



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HUMAN BODY TISSUES AND FLUIDS (cont'd)	<u>Clinical Parasitology</u> (cont'd) Parasitological examination activities for the purposes of clinical diagnosis. Detection of:	In house documented procedures based on equipment manuals and standard methods as specified:	
Serum	Human anti-parasite antibodies to Leishmania spp.	Manual method using Apacor rK39 Rapydtest and KIT Direct Agglutination Test PAR-SER-011	A
Serum and CSF	Human anti-parasite antibodies to Taenia solium and Toxocara sp.	Western Blot method using LDBIO kits. PAR-SER-012 & PAR-SER-013	A
EDTA Blood	P. falciparum and non P. falciparum (excl knowsleii) malarial antigens	Rapid Diagnostic Testing using CareStart Malaria Rapydtest. pLDH/HRP2 Combo (Pan/Pf) by lateral flow immunochromatography PAR-MSY-002	A
Blood, biopsy, bone marrow aspirate, bone marrow trephines and slit skin smear	Leishmania spp	Manual liquid culture method followed by observation, microscopy and Romanowsky stain PAR-MSY-013	A
Stool	Strongyloides spp. and Hookworm parasites	Manual culture method followed by observation PAR-MSY-006	A
Cyst fluids	Entamoeba histolytica and Echinococcus granulosus at remote sites	Direct microscopy and microscopy following vital staining with Eosin for viability PAR-MSY-010	C
Serum, Plasma and CSF	Human anti-parasite antibodies to African Trypanosoma spp.: T.b.gambiense  T.b.rhodesiense	Commercial IFAT (except where shown) PAR-SER-002  T.b.g IFAT (ITG)  T.b.r 1.1 and T.b.r.1.18 IFATS (ITG)	A



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Serum and CSF	T.b.rhodesiense	in-house IFAT	A
Stool (Hot)	Intestinal Entamoeba histolytica	Microscopy on wet preps PAR-MSY-005	A
END			